CLH report

Proposal for Harmonised Classification and Labelling

Based on Regulation (EC) No 1272/2008 (CLP Regulation), Annex VI, Part 2

Substance Name: Mandipropamid

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Part A.

1 PROPOSAL FOR HARMONISED CLASSIFICATION AND LABELLING

1.1 Substance

Table 1: Substance identity

Substance name:	Mandipropamid
EC number:	-
CAS number:	374726-62-2
Annex VI Index number:	-
Degree of purity:	≥93 %
Impurities:	No relevant impurities

1.2 Harmonised classification and labelling proposal

Table 2: The current Annex VI entry and the proposed harmonised classification

	CLP Regulation	Directive 67/548/EEC (Dangerous Substances Directive; DSD)
Current entry in Annex VI, CLP Regulation	No entry	No entry
Current proposal for consideration by RAC	Aquatic acute 1 – H400 Aquatic chronic 2 – H411	N; R50/53
Resulting harmonised classification (future entry in Annex VI, CLP Regulation)	Aquatic acute 1 – H400 Aquatic chronic 2 – H411	N; R50/53

1.3 Proposed harmonised classification and labelling based on CLP Regulation and/or DSD criteria

Table 3: Proposed classification according to the CLP Regulation

CLP Annex I ref	Hazard class	Proposed classification	Proposed SCLs and/or M- factors	Current classification 1)	Reason for no classification ²⁾
2.1.	Explosives	1			conclusive but not sufficient for classification
2.2.	Flammable gases	1			conclusive but not sufficient for classification
2.3.	Flammable aerosols	1			conclusive but not sufficient for classification
2.4.	Oxidising gases	1			conclusive but not sufficient for classification
2.5.	Gases under pressure	1			conclusive but not sufficient for classification
2.6.	Flammable liquids	1			conclusive but not sufficient for classification
2.7.	Flammable solids				Data conclusive, but not sufficient for classification
2.8.	Self-reactive substances and mixtures	-			Data lacking
2.9.	Pyrophoric liquids	-			Data conclusive, but not sufficient for classification
2.10.	Pyrophoric solids	-			inconclusive
2.11.	Self-heating substances and mixtures	-			inconclusive
2.12.	Substances and mixtures which in contact with water emit flammable gases	-			Data conclusive, but not sufficient for classification
2.13.	Oxidising liquids	-			Data conclusive, but not sufficient for classification
2.14.	Oxidising solids	1			Data conclusive, but not sufficient for classification
2.15.	Organic peroxides	-			Data conclusive, but not sufficient for classification
2.16.	Substance and mixtures corrosive to metals	-			Data conclusive, but not sufficient for

					classification
3.1.	Acute toxicity - oral	No classification	-	-	conclusive, but not sufficient for classification
	Acute toxicity - dermal	No classification	-	-	conclusive, but not sufficient for classification
	Acute toxicity - inhalation	No classification	-	-	conclusive, but not sufficient for classification
3.2.	Skin corrosion / irritation	No classification	-	-	conclusive, but not sufficient for classification
3.3.	Serious eye damage / eye irritation	No classification	-	-	conclusive, but not sufficient for classification
3.4.	Respiratory sensitisation	No classification	-	-	conclusive, but not sufficient for classification
3.4.	Skin sensitisation	No classification	-	-	conclusive, but not sufficient for classification
3.5.	Germ cell mutagenicity	No classification	-	-	conclusive, but not sufficient for classification
3.6.	Carcinogenicity	No classification	-	-	conclusive, but not sufficient for classification
3.7.	Reproductive toxicity	No classification	-	-	conclusive, but not sufficient for classification
3.8.	Specific target organ toxicity –single exposure	No classification	-	-	conclusive, but not sufficient for classification
3.9.	Specific target organ toxicity – repeated exposure	No classification	-	-	conclusive, but not sufficient for classification
3.10.	Aspiration hazard	No classification	-	-	conclusive, but not sufficient for classification
4.1.	Hazardous to the aquatic environment	Aquatic Acute 1, H400 Aquatic Chronic 2, H411	1		
5.1.	Hazardous to the ozone layer				Data Lacking

Labelling: Signal word: - Warning

Hazard statements:

Very toxic to aquatic life H400

Toxic to aquatic life with long lasting effects H411

¹⁾ Including specific concentration limits (SCLs) and M-factors
2) Data lacking, inconclusive, or conclusive but not sufficient for classification

Precautionary statements:

P101	If medical advice is needed, have product container or label at hand.
P102	Keep out of reach of children.
P270	Do no eat, drink or smoke when using this product.
P273	Avoid release to the environment
P391	Collect spillage
P501	Dispose of contents/container to

Suppl. Hazard:

EUH401 To avoid risks to human health and the environment, comply with the instructions for use.

Proposed notes assigned to an entry:

Table 4: Proposed classification according to DSD

Hazardous property	Proposed classification	Proposed SCLs	Current classification 1)	Reason for no classification ²⁾
Explosiveness	-			Data conclusive, but not sufficient for classification
Oxidising properties	-			Data conclusive, but not sufficient for classification
Flammability	-			Data conclusive, but not sufficient for classification
Other physico-chemical properties [Add rows when relevant]	1			-
Thermal stability	1			Data conclusive, but not sufficient for classification
Acute toxicity	No classification	-	-	conclusive, but not sufficient for classification
Acute toxicity – irreversible damage after single exposure	No classification	-	-	conclusive, but not sufficient for classification
Repeated dose toxicity	No classification	-	-	conclusive, but not sufficient for classification
Irritation / Corrosion	No classification	-	-	conclusive, but not sufficient for classification
Sensitisation	No classification	-	-	conclusive, but not sufficient for classification
Carcinogenicity	No classification	-	-	conclusive, but not sufficient for classification
Mutagenicity – Genetic toxicity	No classification	-	-	conclusive, but not sufficient for classification
Toxicity to reproduction – fertility	No classification	-	-	conclusive, but not sufficient for classification
Toxicity to reproduction – development	No classification	-	-	conclusive, but not sufficient for classification
Toxicity to reproduction – breastfed babies. Effects on or via lactation	No classification	-	-	conclusive, but not sufficient for classification
Environment 1) Including SCLs	N R50/53			

¹⁾ Including SCLs

<u>Labelling:</u> <u>Indication of danger:</u> N Dangerous for the Environment

R-phrases: R50/53 Very toxic to aquatic organisms, may cause long-term adverse

effect in the aquatic environment.

<u>S-phrases:</u> S2 Keep out of the reach of children

S13 Keep away from food, drink and animal feeding stuffs

S20/21 When using do not eat, drink or smoke

²⁾ Data lacking, inconclusive, or conclusive but not sufficient for classification

- S56 Dispose of this material and its container to hazardous or special waste collection point.
- S57 Use appropriate container to avoid environmental contamination.
- S60 This material and its container must be disposed of as hazardous waste.
- S61 Avoid release to the environment. Refer to special instructions/safety data sheets.

2 BACKGROUND TO THE CLH PROPOSAL

2.1 History of the previous classification and labelling

Mandipropamid is a new mandelamide fungicide. The compound was applied as new active substance under Council Directive 91/414/EEC, with Austria as Rapporteur Member State. In accordance with Article 36(2) of the CLP Regulation, mandipropamid should now be considered for harmonised classification and labelling. Therefore, this proposal considers all physical and chemical properties, human health and environmental endpoints. This Annex VI dossier presents a classification and labelling proposal based mainly on the information presented in the assessment of mandipropamid under Directive 91/414/EEC. This assessment (DAR) was based on one full data package submitted by one company.

Mandipropamid is not currently listed in Annex VI of Regulation EC 1272/2008 (CLP Regulation). Following evaluation of the data this proposal seeks to propose classification for the environment. No classification for physical and chemical properties and human health is proposed.

2.2 Short summary of the scientific justification for the CLH proposal

No classification and labelling has been proposed regarding physical and chemical properties by Austria as Rapporteur Member State for mandipropamid

Regarding classification criteria for Mandipropamid for aquatic environment hazards acute category 1 (very toxic to aquatic organisms) is proposed.

Regarding environment (considering 2nd ATP criteria) following classification will be proposed:

DSD: N, R50/53 (DSD)

CLP: Aquatic Acute 1, H400, M=1; Aquatic Chronic 2, H411

Aquatic Acute classification is based on:

• EC50 value for *Crassostrea virginica* = 0.97 mg/L (Palmer et al 2005c), resulting in N, R50 (DSD) and Aquatic Acute 1, H400, M = 1 (CLP)

Aquatic chronic classification is based on:

- Mandipropamid is not considered as ready biodegradable/rapid degradable. Therefore a R53 (DSD) classification is proposed.
- chronic aquatic toxicity studies

 Based on the non rapid degradability and on the toxicity to Daphnia (Grade 2003) with

 a NOEC= 0.28 mg/L. a classification with <u>Aquatic Chronic 2, H411 (CLP)</u> is proposed.

2.3 Current harmonised classification and labelling

Mandipropamid has not been previously discussed or agreed at TC C&L (Dir. 67/548/EEC); no harmonised classification and labelling exist.

- 2.3.1 Current classification and labelling in Annex VI, Table 3.1 in the CLP Regulation
- 2.3.2 Current classification and labelling in Annex VI, Table 3.2 in the CLP Regulation
- 2.4 Current self-classification and labelling

No current self-classification and labelling based on CLP Regulation criteria.

- 2.4.1 Current self-classification and labelling based on the CLP Regulation criteria
- 2.4.2 Current self-classification and labelling based on DSD criteria
- 3 JUSTIFICATION THAT ACTION IS NEEDED AT COMMUNITY LEVEL

No need for justification (mandipropamid is a pesticide)

Part B.

SCIENTIFIC EVALUATION OF THE DATA

1 IDENTITY OF THE SUBSTANCE

1.1 Name and other identifiers of the substance

Table 5: Substance identity

EC number:	
EC name:	Mandipropamid
CAS number (EC inventory):	
CAS number:	374726-62-2
CAS name:	Benzeneacetamide, 4-chloro-N-[2-[3-methoxy-4-(2-propyn-1-yloxy)phenyl]ethyl]-α-(2-propyn-1-yloxy)-
IUPAC name:	2-(4-chlorophenyl)-N-{2-[3-methoxy-4-(prop-2-yn-1-yloxy)phenyl]ethyl}-2-(prop-2-yn-1-yloxy)acetamide
CLP Annex VI Index number:	
Molecular formula:	C ₂₃ H ₂₂ ClNO ₄
Molecular weight range:	411.9 g/mol

Structural formula:

1.2 <u>Composition of the substance</u>

Table 6: Constituents (non-confidential information)

Constituent	Typical concentration	Concentration range	Remarks
Mandipropamid	94.7 %	95.7 %	97.3 %

Current Annex VI entry: No entry available.

Table 7: Impurities (non-confidential information)

Impurity	Typical concentration	Concentration range	Remarks
-			

Table 8: Additives (non-confidential information)

Additive	Function	Typical concentration	Concentration range	Remarks
-				

1.2.1 Composition of test material

<u>Human health hazard assessment:</u> purity of tested technical material in the range from 96.5% to 98.5%.

Environmental hazard assessment: purity of tested technical material in the range from 96.1% to 99.5%

1.3 Physico-chemical properties

Table 9: Summary of physico - chemical properties

Study	Method	Material	Results	Conclusion/Comment	Reference
B.2.1.1 Melting point,	OECD 102 capillary method with	pure substance	96.4° C to 97.3 °C	Acceptable	Das R., (2002a) (NOA446510/ 0024)
freezing point or solidification point (IIA 2.1.1)	photocell detection GLP	990 g/kg		EEC/A1 is based on OECD 102	
B.2.1.2 Boiling point	OECD 103	pure	Thermal decomposition starts at about 200 °C	Acceptable	Das R., (2003a)
(IIA 2.1.2)	GLP	990 g/kg		EEC/A2 is based on OECD 103	(0500 015011001)
B.2.1.3	OECD 103	pure	Thermal decomposition starts at about 200 °C	Acceptable	Das R., (2003a)
I emperature of decomposition or sublimation	DSC GLP	substance 990 g/kg		EEC/A2 is based on OECD 103	(NOA446510/ 0038)
(IIA 2.1.3)	OECD 113	tech.	Stable in nitrogen or air, no thermal decomposition	Acceptable	Vehling H., (2005)
	DSC and TGA GLP	substance 952 g/kg	or weight loss attributable to reaction/decomposition at room temperature		(NOA446510/ 0401)
B.2.1.4	OECD 109	pure	1.24 x 10 ³ kg/m ³ at 22 °C	Acceptable	Füldner H., (2003)
Relative density	pycnometer	substance	corresponds to a relative density $= 1.24$		(NOA446510/ 0031)
(IIA 2.2)	-air comparison GLP	990 g/kg		EEC/A3 is based on OECD 109	
B.2.1.5	OECD 104	bnre	$< 9.4 \text{ x } 10^{-7} \text{ Pa}$ at 20 °C	Acceptable	Geoffroy A., (2003)
Vapour pressure	gas saturation	substance			(NOA446510/0064)
(IIA 2.3.1)	GLP	990 g/kg	$< 9.4 \text{ x } 10^{-7} \text{ Pa}$ at $50 ^{\circ}\text{C}$	EEC/A4 is based on	Das R., (2006d)
			The formation of the fo	OECD 104	(Doc. 10115200)
			HPLC was used for the determination of the concentration of mandipropamid condensed in the		
			U-tube, based on the method of analysis used for the		
			determination of active substance in technical grade		
			material. The reference solutions were prepared with different		
			The second secon		

> CII/	Method	Material	Results			Conclusion/Comment	Reference
			concentrations in order to take into account the expected concentrations of mandipropamid in the test solutions.	to take into acco s of mandipropar	ount the mid in the		
B.2.1.6 Volatility, Henry's law constant (IIA 2.3.2)	calculation	I	$< 9.2 \times 10^{-5}$ Pa m ³ / mol at 25 °C: values used for calculation: vapour pressure at 25 °C: $< 9.4 \times 10^{-7}$ Pa water solubility at 25 °C: 4.2 mg/L	Pa m³ / mol at 25 °C: ior calculation: ure at 25 °C: < 9.4 x 10^{-7} F ity at 25 °C: 4.2 mg/L	e,	Acceptable	Baker S., (2005) (NOA446510/ 0445)
B.2.1.7 Appearance: physical state (IIA 2.4.1)	Visual assessment GLP Visual assessment GLP	pure substance 990 g/kg tech.	light beige powder			Acceptable Acceptable	Das R., (2002b) (NOA446510/ 0025) Das R., (2005a) (NOA446510/ 0376)
B.2.1.9 Appearance: odour (IIA 2.4.2)	Organoleptic GLP	952 g/kg pure substance 990 g/kg	odourless			Acceptable	Das R., (2002b) (NOA446510/ 0025)
	Organoleptic GLP	tech. substance 952 g/kg	odourless			Acceptable	Das R., (2005a) (NOA446510/ 0376)
B.2.1.10 Spectra of the active substance (IIA 2.5.1)	GLP	pure substance 990 g/kg	CUV/VIS Solution Wave-neutral neutral Emgth [nm] 223 Emol x cm 20144 (methanol) 276 2724 acidic 223 20144 (methanol/1 N HCl 223 20313 (90+10) 276 2845 (90+10) 276 2845 (90+10) 276 2845 (90+10) 276 2864 (90+10) 276 2864 (90+10) 276 2864 Spectra were found to be in agreement with the proposed chemical structure	Wave- length [nm] [1] 223 276 223 276 223 276 ect to the IR, NM be in agreement w	E [L/mol x cm] 20144 2724 20313 2845 19414 2864 MR and MS with the	Acceptable	Oggenfuss P., (2004) (NOA446510/ 0082)

Study	Method	Material	Results	Conclusion/Comment	Reference
B.2.1.10.1 Optical purity				Not relevant	
B.2.1.11 Spectra of relevant impurities (IIA 2.5.2)			Not required as no impurities of toxicological or ecotoxicological concern were identified	Acceptable	Tier II
B.2.1.12 Solubility in water	OECD 105 flask method	pure substance	4.2 mg/L in pure water at 25 $^{\circ}$ C	Acceptable	Das R., (2003b) (NOA446510/0026)
(IIA 2.6)	GLP	990 g/kg	HPLC was used for the determination of the concentration of mandipropamid, based on the method of analysis used for the determination of active substance in technical grade material. The reference solutions were prepared with different concentrations in order to take into account the expected concentrations of mandipropamid in the test solutions.	EEC/A6 is based on OECD 105 Although column elution method is required for substances which water solubility is < 10 ⁻² g/L the notifier justifies the use of the flask method that the evaporation of organic solvent was not complete and the crystal structure of mandipropamid may be altered during the deposition onto the carrier material of the column. Effect of pH is not required since there is no dissociation in water in the environmentally relevant pH-range (see B.2.1.18)	Das R., (2006a) (Doc. 10115199)
		Metabolite CGA 380775 (CA 3584) pure substance 982 g/kg	The solubility in aqueous buffered solutions at 25 °C has been determined to be: 230 mg/L 230 mg/L pH 7.0 280 mg/L pH 9.0	Acceptable	Das R., (2004a) (CGA380775/0001)
		Metabolite CGA 380778 (R 730383) pure	The solubility in pure water at 25 $^{\circ}$ C is 100 mg/L	Acceptable	Das R., (2004b) (CGA380778/0001)

			Mesuits	Conclusion/Comment	Reference
		substance 980 g/kg			
		Metabolite SYN	The solubility in aqueous buffered solutions at 25 °C has been determined to be:	Acceptable	Das R., (2004c) (SYN500003/0001)
		500003 (R 740990.	45 g/L pH 5.0		
		CA 4013)			
		pure substance			
	•	990 g/kg			(1000 C) H
		Metabolite SYN	The solubility in aqueous buffered solutions at 25°C has been determined to be:	Acceptable	Das R., (2005b) (SYN504851/0005)
		504851 (R	2		
		740991) piire	> 500 g/L pH 7.0 and 9.0		
		substance			
		970 g/kg			
		Metabolite	The solubility in pure water at 25 °C is 26 mg/L	Acceptable	Das R., (2004d)
		535839 pure			(SYNSSS839/0001)
		substance			
	,	960 g/kg			
		Metabolite	The solubility in pure water at 25 °C is 14 mg/L	Acceptable	Das R., 2005c
		536638 (R			(1000/00000011)
		290539)			
		pure			
		substance 980 g/kg			
	•	Metabolite	The solubility in aqueous buffered solutions at 25 °C	Acceptable	Das R., (2005d)
		NOA	determined to be:		(CA4011/0005)
		458422 (CA 4011) pure	51 mg/L pH 5.0 51 mg/l rH 7.0		
		substance			
		990 g/kg			
	CIPAC MT 157.3	tech.	The solubility in different solvents at 25 °C was	Acceptable	Das R., (2005e)
solvents	JLF	substance 952 g/kg	led to be:		(INOA440310/0373) Das P (2006k)
(IIA 2.7)		3.72 gl ng	aceune 300 g/L		Das K., (2000b)

Study	Method	Material	Results	Conclusion/Comment	Reference
			dichloromethane 400 g/L ethyl acetate 120 g/L hexane 42 mg/L octanol 66 g/L toluene 29 g/L HPLC was used for the determination of the concentration of mandipropamid, based on the method of analysis used for the determination of active substance in technical grade material. The reference solutions were prepared with different concentrations in order to take into account the expected concentrations of mandipropamid in the test solutions.		(Doc. 10115182)
B.2. l.14 Partition coefficient n-octanol/water (IIA 2.8.1)	OEDC 10/ (shake flask method) GLP	pure substance 990 g/kg Metabolite CGA 380775 (CA 3584) pure a	ctanol/water partition pure water was deter = 1600 (± 42) C was used for the defentration of mandipro od of analysis used fo e substance in technic eference solutions we entrations in order to rede concentrations of olutions. ctad concentrations of concentration	Acceptable EEC/A8 is based on OECD 107 Acceptable	Das R., (2003c) (NOA446510/ 0027) Das R., (2006c) (Doc. 10115098) Das R., (2005f) (CGA380775/0003)
		substance 982 g/kg Metabolite CGA 380778 (R	Pow = 120 (± 3.1) log Pow = 2.1 at pH 5.0 Pow = 120 (± 3.0) log Pow = 2.1 at pH 7.0 Pow = 100 (± 1.5) log Pow = 2.0 at pH 9.0 The octanol/water partition coefficient (Pow) at 25 °C was determined to be: Pow = 360 (± 11) log Pow = 2.6	Acceptable	Das R., (2004e) (CGA380778/0002)

Study	Method	Material	Results	Conclusion/Comment	Reference
		pure substance 980 g/kg			
		Metabolite SYN 500003 (R 740990,	The octanol/water partition coefficient (Pow) at 25 $^{\circ}$ C was determined to be: Pow = 1.8 (± 0.051) log Pow = 0.27 at pH 5.0	Acceptable	Das R., (2005g) (SYN500003/0006)
		CA 4013) pure substance 990 g/kg	$= 0.057 (\pm 0.0020) \log Pow =$ $= 0.026 (\pm 0.00089) \log Pow =$		
		Metabolite SYN 504851 (R	The octanol/water partition coefficient (Pow) at 25 °C was determined to be:	Acceptable	Das R., (2005h) (SYN504851/0008)
		740991) pure substance 970 g/kg	Pow = $4.9 (\pm 0.053)$ log Pow = 0.69 at pH 5.0 Pow = $0.14 (\pm 0.0083)$ log Pow = -0.86 at pH 6.8 6.8 Pow = $0.055 (\pm 0.0011)$ log Pow = -1.3 at pH 9.0		
		Metabolite SYN 535839 pure substance 960 o/ko	The octanol/water partition coefficient (Pow) at 25 $^{\circ}$ C was determined to be: Pow = 9000 (\pm 400) log Pow = 4.0	Acceptable	Das R., (2004f) (SYN535839/0002)
		Metabolite SYN 536638 (R	octanol/water partition as determined to be:	Acceptable	Das R., (2005i) (SYN536638/0002)
		pure substance 980 g/kg	$Pow = 4000 (\pm 59)$ $log Pow = 3.6$		
		Metabolite NOA 458422 (CA	The octanol/water partition coefficient (Pow) at 25 °C was determined to be:	Acceptable	Das R., (2005j) (CA4011/0007)
		4011) pure substance 990 g/kg	Pow = $600 (\pm 18)$ log Pow = 2.8 at pH 5.0 Pow = $550 (\pm 6.7)$ log Pow = 2.7 at pH 7.0 Pow = $560 (\pm 25)$ log Pow = 2.8 at pH 9.1		
Effect of pH (4-10) on the			Not relevant as the active substance shows no pH dependency	(see B.2.1.18)	

Study	Method	Material	Results	Conclusion/Comment	Reference
n-octanol/water partition co-efficient (IIA 2.8.2)					
B.2.1.15 Hydrolysis rate (IIA 2.9.1)	OECD 111 GLP	14C labelled NOA 446510, 989 g/kg radio-chemical purity	The hydrolytic behaviour of NOA 446510 was investigated in diluted aqueous buffer solutions at pH 4, 7 and 9 using ¹⁴ C labelled active substance. The recovery for all samples was between 92.7 and 105.7% of the applied radioactivity. No degradation of the test substance was observed under all conditions. Therefore, NOA 446510 is hydrolytically stable at pH 4, 7 and 9.	Acceptable For details see B.8.4 Fate and behaviour	Buckel T., (2002) (NOA446510/ 0018)
B.2.1.16 Direct phototransformation (IIA 2.9.2)	OECD, Proposal "Phototrans- formation of Chemicals in Water – Direct and Indirect Photolysis" Draft, Aug. 2000 GLP	14C labelled NOA 446510 1980 Bq μḡ 1 radio- chemical purity: >99%	The photolytic degradation was evaluated in sterile buffer solution at pH 7, under a xenon arc light, at 25°C using ¹⁴ C-labelled test substance at a concentration of 1 µg mL ⁻¹ . The samples were irradiated for periods up to the equivalent of 17 days summer sunlight. Duplicate "dark" control samples were also prepared and maintained at 25 °C and analysed at the same time period as the irradiated samples. The estimated half-life was 34 hours of continuous irradiation. At least 16 degradates were formed, none of which represented >5% of the applied radioactivity at any point during the study. With further irradiation these degradates were broken down further to at least 10 highly polar degradates. The mean mass balance from irradiated samples was 102.9% of the applied radioactivity, of which up to 16.2% was characterised as ¹⁴ CO ₂ .	Acceptable For details see B.8.4 Fate and behaviour	Hand L and Towers J., (2003) (NOA446510/ 0041)
B.2.1.17 Quantum yield (IIA 2.9.3)	OECD, Proposal "Phototrans- formation of Chemicals in Water – Direct and Indirect Photolysis" Draft, Aug. 2000 GLP	pure substance 990 g/kg	The quantum yield of direct photolysis was found to be $\Phi = 0.37$, at 300 nm wavelength of applied light. Photolytic half-life in shallow waters was estimated for geographic latitudes of 30° N, 40° N and 50° N for all seasons. Summer half-lives between 30 and 60 days were calculated for 30° N and 50° N respectively.	Acceptable For details see B.8.4 Fate and behaviour	Schmidt E., (2004) (NOA446510/ 0118)

Study	Method	Material	Resulfs	Conclusion/Comment	Reference
B.2.1.18 Dissociation constant (pKa)	OECD 112 spectrophotometric	pure substance 990 g/kg	No pKa was found at various pH values in the range of 1.0 to 12.0 of a solution of NOA 446510 in water.		Martin N., (2003) (NOA446510/ 0029)
(IIA 2.9.4)	GLP	Metabolite CGA	pKa = 10.34 at 20 °C	Acceptable	Martin N., (2004a) (CGA380775/0002)
		380775 (CA 3584) pure			
		substance 982 g/kg			
		Metabolite		Acceptable	Martin N., (2004b)
		CGA 380778 (R	pKa = 11.64 at 20 °C		(CGA380 / /8/0004)
		730383) puire			
		substance 980 g/kg			
		Metabolite		Acceptable	Martin N., (2004c)
		SYN 500003 (B	$pKa = 2.76$ at $20 ^{\circ}C$		(SYN500003/0002)
		740990,			
		CA 4015)			
		substance			
		990 g/kg			
		Metabolite	100	Acceptable	Richner D., (2005a)
		504851 (R	$p_{Na} = 2.91$ at 20 °C		(+000/100+0011g)
		740991)			
		pure			
		substance 970 g/kg			
		Metabolite		Acceptable	Martin N., (2004d)
		535839 pure	No pKa was found in the pH range of 1.0 to 12.0		(SYN555859/0005)
		substance			
		960 g/kg			
		Metabolite		Acceptable	Martin N., (2005)
		NIN	No pKa was found in the pH range of 1.0 to 12.0		(51/05/56658/0003)

Study	Method	Material	Results	Conclusion/Comment	Reference
		536638 (R 290539) pure substance 980 g/kg			
		Metabolite NOA 458422 (CA 4011) pure substance 990 g/kg	pKa = 10.37 at 20 °C	Acceptable	Richner D., (2005b) (CA4011/0006)
B.2.1.19 Stability in air, photochemical oxidative degradation (IIA 2.10)	Calculation with Atmospheric Oxidation Program based on Atkinson method		The atmospheric oxidation of NOA 446510 by hydroxyl radicals was estimated by calculation according to Atkinson. The estimated half-life is 1.4 hours.	Acceptable	Widmer H., (2003) (NOA446510/0035)
B.2.1.20 Flammability (IIA 2.11)	EEC A10 GLP	tech. substance 952 g/kg	Preliminary Test: the test substance melted and charred but did not ignite According to EEC A10 a full test is not required	Acceptable Not classified as highly flammable in terms of its burning characteristics	Jackson W.A., (2005a) (NOA446510/0405)
B.2.1.21 Auto-flammability (IIA 2.11.2)	BEC A16 GLP	tech. substance 952 g/kg	No ignition was detected below the melting point	Acceptable Compound is not considered as auto- flammable under the test conditions	Jackson W.A., (2005b) (NOA446510/ 0403)
B.2.1.22 Flash point (IIA 2.12)			Not relevant NOA 446510 is a solid with a melting point > 40 °C		
B.2.1.23 Explosive properties (IIA 2.13)	EEC A14 GLP	tech. substance 952 g/kg	The substance did not explode when exposed to heat, mechanical shock or friction	Acceptable Compound is not considered as explosive under the test conditions of EEC/A14 Although dust explosion does not cover this annex point, the MSDS for	Jackson W.A., (2005c) (NOA446510/ 0404)

Study	Method	Material	Results	Conclusion/Comment	Reference
				mandipropamid technical indicates that the compound is capable of forming flammable dust clouds in air, which can produce a dust cloud explosion, if ignited.	
B.2.1.24 Surface tension (IIA 2.14)	OECD 115 GLP	tech. substance 952 g/kg	$\sigma = 72.8 \text{ mN} / \text{ m}$ 90 % of the saturation concentration at 20 °C	Acceptable Mandipropamid is not regarded as surface active.	Richner D., (2005c) (NOA446510/ 0426)
B.2.1.25 Oxidizing properties (IIA 2.15)	EC A17 GLP	tech. substance 952 g/kg	Not an oxidising substance. The maximum overall burning rate is 2.4 mm/s for the 5% test substance mixture. This is lower than the max. burning rate of the reference mixture (3.4 mm/s) containing of 60% Ba(NO ₃)2/Cellulose.	Compound is not considered as oxidizing under the test conditions	Jackson W.A., (2005d) (NOA446510/ 0402)
B.2.1.2.26 pH (IIA 2.16)				This is not an EC data requirement	
Storage stability (IIA 2.17.1)				This is not an EC data requirement	
Stability (temperature, metals) (IIA 2.17.2)				This is not an EC data requirement	
Other/special studies (IIA 2.18)				This is not an EC data requirement	

According to Directive 91/414/EEC, granulometry is not required for active substances. Thus, no study considering this end-point has been provided.

2 MANUFACTURE AND USES

2.1 Manufacture

Not relevant for Classification and Labelling.

2.2 Identified uses

Mandipropamid is a new mandelamide fungicide agriculture for foliar application on vegetables and grapes.

3 CLASSIFICATION FOR PHYSICO-CHEMICAL PROPERTIES

No classification required.

4 HUMAN HEALTH HAZARD ASSESSMENT

4.1 Toxicokinetics (absorption, metabolism, distribution and elimination)

4.1.1 Non-human information

<u>Absorption</u>: The percentage absorption of radioactivity over 48 hours after dosing was calculated from the amounts found in urine, bile, cage wash and carcass of bile duct cannulated rats. The extent of absorption was similar in both sexes, but more extensive at the low dose level (3 mg/kg bw) than at the high dose level (300 mg/kg bw). At the low dose level, absorption was 74 % in males and 67 % in females. At the high dose level, absorption was 30 % in males and 45 % in females.

<u>Distribution</u>: Following a single oral dose of either 3 or 300 mg/kg of the test substance to both male and female rats, tissue concentrations were considered to be low in all cases and were also consistently lower in female tissues than in male tissues. The highest residues were present in the liver and kidneys 8 hours after application, however, these values declined rapidly. The half life of elimination in the tissues investigated was approximately 24 hours or lower. When repeated doses of 3 mg/kg were administered to male rats, levels of radioactivity found in tissues were also generally low. Tissue accumulation profiles for liver and kidneys indicate that the concentration reached a plateau by 4 days. Elimination from these organs was rapid after termination of dosing. No potential for bioaccumulation was identified.

Excretion: In non-cannulated low dose rats, about 70 % of test substance were excreted within 48 hours, with males excreting most of the radioactivity by faeces and females excreting roughly half of the radioactivity by faeces and the other half by urine. In the high dose animals when absorption was saturated, large proportions of test substance were found in the faeces (about 80 % after 48 hours) while only 2-10 % were found in urine in both sexes. Excretion was almost complete within 7 days of dosing, by which time all excised tissue and carcass residues were very low (<0.5 % of the administered dose). Biliary elimination was important at both dose levels and in both sexes, accounting for 73 % and 55 % of a 3 mg/kg dose and 28 % and 22 % of a 300 mg/kg dose in males and females, respectively. Residues in expired air were near to or below the limit of detection.

Metabolism: The principle steps in metabolism involved loss of one or both propargyl groups, followed by glucuronidation and O-demethylation to produce 6 major metabolites. There were no sex or dose related differences in the qualitative metabolic profile. However, a sex difference was observed in the major route of excretion and relative proportions of metabolites excreted via certain routes. In females, the major route of excretion was via the urine, and NOA 458422 glucuronide was identified as the major urinary metabolite. In males, the major route of excretion was via the faeces, with NOA 458422 being the major metabolite. The substance was extensively metabolised at the low dose level of 3 mg/kg, with 21 % and 12 % of the parent found in the faeces of non-cannulated males and females, respectively. In bile duct-cannulated rats, the amount of unchanged parent compound was 13 % in males and 22 % in females. At the high dose level, where absorption of a gavage dose was less extensive, a high proportion of the parent compound was detected unchanged in the faeces (>70 %). This was considered to represent the unabsorbed dose, since no

unchanged parent was detected in the bile. Similarily, no unchanged parent was present in urine. Repeated daily dosing, investigated in the male rat only, had no effect on the metabolism of mandipropamid, and similar metabolite profiles were obtained 24 hours after the first and fourteenth consecutive daily doses.

4.1.2 Human information

No data available.

4.1.3 Summary and discussion on toxicokinetics

Absorption, distribution, excretion and metabolism (toxicokinetics) (Annex IIA, point 5.1)

Rate and extent of oral absorption ‡	70 % absorbed (rat study, 3 mg/kg bw), based on excretion via urine and bile
Distribution ‡	Uniformly distributed; highest levels found in liver and kidney
Potential for accumulation ‡	no evidence for bioaccumulation
Rate and extent of excretion ‡	rapid, mainly via faeces (70–80% within 48 hours)
Metabolism in animals ‡	loss of one or both propargyl groups, followed by glucuronidation and O-demethylation
Toxicologically relevant compounds ‡ (animals and plants)	Mandipropamid
Toxicologically relevant compounds ‡ (environment)	Mandipropamid

4.2 Acute toxicity

Table 11: Summary table of relevant acute toxicity studies

Type of Study	Species	Vehicle	Results	Reference
Acute Oral	rat	corn oil	LD ₅₀ > 5000mg/kg bw	Moore G (2004)
Acute Dermal	rat	corn oil	$LD_{50} > 5050 mg/kg bw$	Kuhn J (2005)
Acute Inhalation	rat	clean dry air	LC ₅₀ > 5.19 mg/l/4h	Kilgour J (2003)
Skin Irritation	rabbit	moistened with deionized water	Not irritant	Johnson I (2004a)
Eye Irritation	rabbit	test substance was used as supplied	Not Irritant	Johnson I (2004b)
Skin sensitisation (LLNA)	mouse	dimethylformamide (DMF)	Not a sensitiser	Johnson I (2004c)

4.2.1 Non-human information

4.2.1.1 Acute toxicity: oral

After oral application to female rats, the acute oral LD_{50} of mandipropamid techn. was greater than 5000 mg/kg/bw.

All animals survived and gained bodyweight during the 14 days following dosing. Apart from anogenital staining noted for one rat five hours post-dosing, all animals appeared active and healthy over the 14-day observation period. There were no other signs of gross toxicity or abnormal behaviour. No gross abnormalities were noted at examination *post mortem*.

4.2.1.2 Acute toxicity: inhalation

Nose-only exposure for 4 hours to a particulate concentration of 5.19 mg/l resulted in no deaths and signs of mild irritation to the respiratory tract from which the animals made a rapid recovery. It is concluded that the LC₅₀ of mandipropamid exceeds 5.19 mg/l.

During and immediately following exposure, abnormalities generally associated with restraint (wet fur, stains around the nose) were observed in all animals. Other signs noted were slight salivation and signs of respiratory tract irritation (increased breathing depth and abnormal respiratory noise). All female animals had completely recovered by day 2 and in males only abnormal respiratory noise remained in 2 males on day 2 and in one male on day 3. All animals had fully recovered by day 4 of the study.

There were not treatment-related findings at necropsy.

4.2.1.3 Acute toxicity: dermal

The acute dermal LD_{50} of mandipropamid techn. to male and female rats was greater than 5050 mg/kg bodyweight.

There were no signs of systemic toxicity or skin irritation. All animals showed normal weight gain during the study. There were no treatment-related findings at examination *post mortem*.

4.2.1.4 Acute toxicity: other routes

No information on other routes.

4.2.2 Human information

No information available from case reports, epidemiological studies, medical surveillance, reporting schemes and national poisons centres.

4.2.3 Summary and discussion of acute toxicity

Mandipropamid is of low acute oral, dermal and inhalative toxicity in rats (rat oral $LD_{50} > 5000$ mg/kg bw, dermal $LD_{50} > 5050$ mg/kg bw, $LC_{50} > 5.19$ mg/L air/4h.

4.2.4 Comparison with criteria

All estimated LD_{50} values are above the criteria for classification and labelling (both DSD and CLP).

4.2.5 Conclusions on classification and labelling

No classification and labelling is proposed regarding acute toxicity.

4.3 Specific target organ toxicity – single exposure (STOT SE)

4.3.1 Summary and discussion of Specific target organ toxicity – single exposure

No <u>specific</u>, non lethal, <u>target organ toxicity</u> after single exposure was observed in acute toxicity studies. In addition, no human data are available that would support classification for this endpoint. No classification as STOT-SE under the CLP Regulation is proposed.

4.3.2 Comparison with criteria

No effects observed in acute toxicity studies would trigger criteria for classification and labelling STOT SE.

4.3.3 Conclusions on classification and labelling

No classification and labelling is proposed regarding specific target organ toxicity after single exposure.

4.4 Irritation

4.4.1 Skin irritation

Table 12: Summary table of relevant skin irritation studies

Method	Results	Remarks	Reference
Dermal irritation study	Rabbit (New Zealand White albino)	slight erythema and slight desquamation in one animal for 4 days.	Johnson, I.R.; 2004

4.4.1.1 Non-human information

Two male and one female rabbits were dermally exposed for four hours to 500 mg of mandipropamid techn. There were no signs of ill-health in any animal during the study. Very slight erythema was seen in one animal for 4 days. Slight desquamation was also seen in the same animal on day 4 only. There were no other signs of skin irritation. All signs of irritation had completely resolved within 7 days of application.

4.4.1.2 Human information

No information available from case reports, epidemiological studies, medical surveillance, reporting schemes and national poisons centres.

4.4.1.3 Summary and discussion of skin irritation

According to the results of the rabbit skin irritation study, mandipropamid is <u>not irritant</u> to the intact shaved rabbit skin.

4.4.1.4 Comparison with criteria

Estimated skin irritation scores are below the criteria for classification and labelling (according to both DSD and CLP).

4.4.1.5 Conclusions on classification and labelling

No classification and labelling is proposed for mandipropamid regarding skin irritation.

4.4.2 Eye irritation

Table 13: Summary table of relevant eye irritation studies

Method	Results	Remarks	Reference
Eye irritation study	Rabbit (New Zealand White albino)	Slight iritis, redness and chemosis	Johnson, I.R.; 2004

4.4.2.1 Non-human information

Two male and one female rabbits received approximately 100mg mandipropamid techn. into the conjunctival sac of the left eye. There were no signs of ill health in any animal during the study. Application into the eye caused practically no or slight initial pain (class 1-2 on a 0-5 scale). There

were no corneal effects. Slight iritis was seen in two animals approximately 1 hour after application. Conjunctival effects were seen in all animals and consisted of slight or moderate redness and slight or mild chemosis for up to 4 days, and a slight discharge approximately 1 hour after application. Additional signs of irritation comprised lachrymatory discharge and dried secretion around the periorbital skin. All signs of irritation had completely resolved within 7 days of application.

4.4.2.2 Human information

No information available from case reports, epidemiological studies, medical surveillance, reporting schemes and national poisons centres.

4.4.2.3 Summary and discussion of eye irritation

According to the results of the rabbit eye irritation study, mandipropamid is not irritant.

4.4.2.4 Comparison with criteria

Estimated eye irritation scores (24 - 72 hours) are below the criteria for classification and labelling (according to both DSD and CLP).

4.4.2.5 Conclusions on classification and labelling

No classification and labelling is proposed for mandipropamid regarding eye irritation.

4.4.3 Respiratory tract irritation

4.4.3.1 Non-human information

There is no specific information regarding the ability of mandipropamid to cause irritation to the respiratory tract during the acute inhalation toxicity study.

4.4.3.2 Human information

No information available from case reports, epidemiological studies, medical surveillance, reporting schemes and national poisons centres.

4.4.3.3 Summary and discussion of respiratory tract irritation

No classification is proposed for respiratory tract irritation.

4.4.3.4 Comparison with criteria

4.4.3.5 Conclusions on classification and labelling

4.5 Corrosivity

Based on the data from the skin and eye irritation studies it can be concluded that mandipropamid is not corrosive.

4.6 Sensitisation

4.6.1 Skin sensititsation

Table 15: Summary table of relevant skin sensitisation studies

Method	Results	Remarks	Reference
Local Lymph Node Assay	Not sensitising		Johnson, I.R.; 2005

4.6.1.1 Non-human information

Mandipropamid techn. was assessed for its skin sensitisation potential using the mouse Local Lymph Node Assay (LLNA). Under the conditions of this test, application of mandipropamid at concentrations of 10%, 25% or 50% w/v in DMF resulted in a less than 3-fold isotope incorporation at all three concentrations. Mandipropamid is therefore unlikely to be a sensitiser.

4.6.1.2 Human information

No information available from case reports, epidemiological studies, medical surveillance, reporting schemes and national poisons centres.

4.6.1.3 Summary and discussion of skin sensitisation

According to the results of the skin sensitisation study (mouse Local Lymph Node Assay), mandipropamid is <u>not sensitising</u>. According to classification criteria, classification and labelling is not warranted.

4.6.1.4 Comparison with criteria

Effects observed in the skin sensitisation study in an mouse Local Lymph Node Assay are below the criteria for triggering classification and labelling (according to both DSD and CLP).

4.6.1.5 Conclusions on classification and labelling

No classification and labelling is proposed for mandipropamid regarding skin sensitisation.

4.6.2 Respiratory sensitisation

No data on respiratory sensitisation available.

4.7 Repeated dose toxicity

Table 17: Summary table of relevant repeated dose toxicity studies

Method	Dose levels	• NOAEL	Remarks (Relevant effects at the LOAEL)	Reference
Wistar rats, 90 days oral	0, 100, 500, 3000, 5000 ppm/diet (equivalent to 0, 8.2, 41.1, 260 and 435 mg/kg bw (♂); 0, 8.9, 44.7, 260 and 443 mg/kg bw (♀)	500 ppm (41.1mg/kg bw ♂; 44.7 mg/kg bw ♀)	-bodyweight ↓ -bodyweight gain ↓ -haematological and clinical chemical findings -liver weight ↑ -periportal hypertrophy/eosinophilia -kidney weight ↑ tubular basophilia	Pinto P; 2005a
Beagle dogs, 90 days oral	0, 5, 25, 100, 400 mg/kg bw/d (capsule)	25 mg/kg bw/d	-haematological and clinical chemical findings -liver weight ↑ -porphyrin deposition	Brammer A; 2005a
Beagle dogs, 1 year oral	0, 5, 40, 400 mg/kg bw/d (capsule)	5 mg/kg bw/d	-bodyweight ↓ -haematological and clinical chemical findings -liver weight ↑ -porphyrin deposition	Brammer A; 2005b
Wistar rats, 28 days dermal	0, 250, 500, 1000 mg/kg bw/d	1000 mg/kg bw/d	No toxicologically significant changes at the highest dose tested	Lees D; 2005

4.7.1 Non-human information

4.7.1.1 Repeated dose toxicity: oral

The short-term toxicity of mandipropamid has been tested in a 90-day oral toxicity study in rats. Mandipropamid has also been tested in 90-day and 1 year oral toxicity studies in dogs via capsule administration.

In a 90 day dietary toxicity study in rats, toxicity was demonstrated at the two highest dose levels (3000 and 5000 ppm), manifest as reduced bodyweight in males. The bodyweights at these two dose levels continued to diverge from the control group throughout the study. At 3000 and 5000 ppm there were decreases in a number of red blood cell parameters, indicating red blood cells as a target. Also at these dose levels there were elevations in plasma albumin, total protein, cholesterol and γ -glutamyl transferase and increased liver weights with associated histopathological change (increased periportal hypertrophy/eosinophilia), indicating the liver to be a target organ. At 500 ppm, relative liver weight in males was slightly higher than controls. Increased kidney weight (3000 and 5000 ppm) and increased incidence of tubular basophilia (5000 ppm) were also noted in males.

The NOAEL was considered to be 500 ppm, equivalent to 41.1 mg/kg bw/d for males and 44.7 mg/kg bw/d for females.

In a 90 day study in dogs, oral administration of mandipropamid resulted in clear evidence of liver toxicity in animals dosed at 400 and 100 mg/kg bw/d. Liver toxicity was characterised by increased liver weight, marked elevations in liver enzymes (alkaline phosphatase and alanine aminotransferase) and porphyrin deposition within the liver.

Other treatment related effects included reductions in white blood cell count and neutrophil count (females at 400 mg/kg bw/d), increases in plasma cholesterol (both sexes at 100 and 400 mg/kg bw/d) and decreases in plasma aspartate aminotransferase activity (both sexes at 400 and males at 100 mg/kg bw/d).

The NOAEL of mandipropamid in this study was considered to be 25 mg/kg bw/d.

In a 1 year study in dogs, oral administration of mandipropamid resulted in clear evidence of toxicity in dogs dosed at 400 and 40 mg/kg bw/d. Effects included reduced bodyweights at 400 mg/kg bw/d and liver toxicity at both dose levels, characterised by increased liver weight, marked elevations in liver enzymes (alkaline phosphatase and alanine aminotransferase) and porphyrin deposition within the liver.

Other treatment related effects included reductions in MCV and MCH (both sexes at 400 mg/kg bw/d), increases in platelets (males at 40 and 400 mg/kg bw/d) and decreased activated partial thromboplastin time (males at 400 mg/kg bw/d).

The NOAEL of mandipropamid in this study was considered to be 5 mg/kg bw/d.

4.7.1.2 Repeated dose toxicity: inhalation

No data available.

4.7.1.3 Repeated dose toxicity: dermal

The short-term toxicity of mandipropamid has been tested in a 28 day percutaneous toxicity study in rats.

Dermal administration of mandipropamid at dose levels up to 1000 mg/kg bw/day for 21 days in a 28 day period to male and female rats produced no evidence of systemic toxicity. There was an increased incidence of signs of slight skin irritation at 250, 500 and 1000 mg/kg/day, including erythema, oedema and desquamation at the application site.

4.7.1.4 Repeated dose toxicity: other routes

No data available.

4.7.1.5 Human information

No information available from case reports, epidemiological studies, medical surveillance, reporting schemes and national poisons centres.

4.7.1.6 Other relevant information

No data available.

4.7.1.7 Summary and discussion of repeated dose toxicity

Repeat dose toxicity studies in mice, rats and dogs confirmed the liver as target organ of mandipropamid.

4.7.1.8 Summary and discussion of repeated dose toxicity findings relevant for classification according to DSD

Effects observed in the subchronic studies in rat, mouse and dog do not trigger the criteria for classification and labelling for repeated dose toxicity.

4.7.1.9 Comparison with criteria of repeated dose toxicity findings relevant for classification according to DSD

Effects observed in the subchronic studies in rat, mouse and dog do not trigger the criteria for classification and labelling for repeated dose toxicity.

4.7.1.10 Conclusions on classification and labelling of repeated dose toxicity findings relevant for classification according to DSD

Effects observed in the subchronic studies in rat, mouse and dog do not trigger the criteria for classification and labelling for repeated dose toxicity.

4.8 Specific target organ toxicity (CLP Regulation) – repeated exposure (STOT RE)

Effects observed in the subchronic studies in rat, mouse and dog do not trigger the criteria for classification and labelling for repeated dose toxicity.

4.9 Germ cell mutagenicity (Mutagenicity)

The mutagenicity of mandipropamid has been adequately investigated in vitro and in vivo.

Table 18: Summary table of relevant in vitro and in vivo mutagenicity studies

Type of Study	Test system	Dose levels	Results	Reference
In vitro studies				
Bacterial reverse mutation	Salmonella typhimurium (TA1535, TA1537, TA98 and TA100) and two strains of Escherichia coli (WP2P and WP2PuvrA)	100 to 5000 μg/plate	Negative	Callander R (2005)
In vitro cytogenetics	human lymphocytes	2.5 to 100 μg/ml	Negative	Fox V (2002)
Mammalian cell gene mutation (mouse lymphoma)	L5178Y mouse lymphoma cells	1-4119μg/ml	Negative	Clay P (2002)
In vivo studies				
Rat bone marrow micronucleus	bone marrow of male rats	2000mg/kg	Negative	Fox V (2005)
Unscheduled DNA synthesis – rat liver	liver of male rats	2000 mg/kg	Negative	Clay (2005)

4.9.1 Non-human information

4.9.1.1 In vitro data

In vitro, mandipropamid was negative in both bacterial (Ames test) and mammalian cells (L5178Y TK^{+/-} mouse lymphoma) for gene mutation. The L5178Y TK^{+/-} assay was also negative for clastogenicity. In the *in vitro* cytogenetic assay using primary human lymphocyte cultures, mandipropamid was examined for evidence of chromosomal damage up to dose levels limited by cytotoxicity to the cells (100μg/ml). Mandipropamid showed no evidence of induced chromosomes aberrations in this assay either in the presence or absence of S9-mix.

4.9.1.2 In vivo data

In vivo, mandipropamid was found to be non-clastogenic in the rat bone marrow micronucleus assay at the limit dose of 2000mg/kg. There was also no evidence for any induction of DNA damage or repair in the rat liver by mandipropamid using the UDS assay.

4.9.2 Human information

No information available from case reports, epidemiological studies, medical surveillance, reporting schemes and national poisons centres.

4.9.3 Other relevant information

No data available.

4.9.4 Summary and discussion of mutagenicity

Mandipropamid has been examined in a range of *in vitro* and *in vivo* genotoxicity assays, including endpoints of gene mutation, chromosomal damage and DNA repair.

All the results were negative, showing that mandipropamid has no genotoxic potential *in vitro* and *in vivo*.

4.9.5 Comparison with criteria

Effects observed in the *in vitro* and *in vivo* mutagenicity studies do not trigger the criteria for classification and labelling for mutagenicity.

4.9.6 Conclusions on classification and labelling

There is no evidence of genotoxic potential of mandipropamid, therefore, no classification is proposed.

4.10 Carcinogenicity

Table 19: Summary table of relevant carcinogenicity studies

Method	Dose levels	NOAEL	Remarks (Relevant effects at the LOAEL)	Reference
Wistar rats, 2 years oral	0, 50, 250, 1000 ppm/diet (equivalent to 0, 3, 15.2 and 61.3 mg/kg bw (♂) and 0, 3.5, 17.6 and 69.7 mg/kg bw (♀)	250 ppm (15.2 mg/kg bw ♂; 17.6 mg/kg bw ♀)	-bodyweight ↓ -bodyweight gain ↓ -haematological and clinical chemical findings -liver weight ↑ -periportal hypertrophy/eosinophilia -chronic progressive nephropathy; osteo-renal syndrome	Pinto P; 2005b
C57BL/1 0J _f CD-1 mice, 80 weeks oral	0, 100, 500, 2000 ppm/diet (equivalent to 0, 10.6, 55.2 and 222.7 mg/kg bw (\circlearrowleft) and 0, 13.2, 67.8 and 284.6 mg/kg bw (\updownarrow)	500 ppm (55.2 mg/kg bw ♂; 67.8 mg/kg bw ♀)	-bodyweight ↓ -bodyweight gain ↓ -liver weight ↑	Milburn G.; 2005a

4.10.1 Non-human information

4.10.1.1 Carcinogenicity: oral

The chronic toxicity and carcinogenic potential of mandipropamid was investigated in rats when administered orally in the diet for a period of up to 105 weeks. The highest dose of 1000 ppm, equivalent to 61.3 mg/kg bw for males and 69.7 mg/kg bw for females, caused decreases in mean cell volume (MCV) and mean cell haemoglobin (MCH) in both male and female rats, indicating red blood cells as a target.

Some clinical chemistry parameters were affected at this dose level too.

Increased liver weights were observed at 1000 ppm. Histopathologically, an increase in the incidence of periportal eosinophilia in the liver in both sexes at a dose level of 1000 ppm and in females also at > 250 ppm was observed at week 53. This confirms the liver as a target organ.

In the kidneys of males, chronic progressive nephropathy associated with an increased incidence of an osteo-renal syndrome was observed at 1000 ppm.

Mandipropamid was not carcinogenic in the rat. There were no treatment-related increases in the incidence of tumours and no trend towards increased numbers of tumours with dose. The NOAEL for mandipropamid was considered to be 250 ppm (15.2 mg/kg bw/day in males and 17.6 mg/kg bw/day in females), based on low body weight, liver and kidney toxicity and changes in red blood and clinical chemistry parameters in the 1000 ppm groups.

The carcinogenic potential of mandipropamid in mice was investigated when administered orally via the diet for at least 80 weeks.

There was a treatment-related reduction in bodyweight and bodyweight gain in both sexes at the highest dose level of 2000 ppm, equivalent to 222.7 mg/kg bw for males and 284.6 mg/kg bw for females.

Increases in liver weights at 2000 ppm in both sexes and at 500 ppm in males confirmed the liver as target organ of mandipropamid.

There were no treatment-related increases in the incidence of tumours and no trend towards increased number of tumours with dose.

500 ppm, equivalent to 55.2 mg/kg bw for males and 67.8 mg/kg bw for females can be considered as NOAEL for mandipropamid in this study.

4.10.1.2 Carcinogenicity: inhalation

No data available.

4.10.1.3 Carcinogenicity: dermal

No data available.

4.10.2 Human information

No information available from case reports, epidemiological studies, medical surveillance, reporting schemes and national poisons centres.

4.10.3 Other relevant information

No data available.

4.10.4 Summary and discussion of carcinogenicity

Based on the results of two submitted studies in rats and mice, mandipropamid can be regarded to have no oncogenic potential.

4.10.5 Comparison with criteria

No oncogenic effects were observed in studies conducted with mandipropamid, neither in rat nor in mouse carcinogenicity studies (according to both DSD and CLP).

4.10.6 Conclusions on classification and labelling

Mandipropamid can be regarded to have no oncogenic potential.

4.11 Toxicity for reproduction

Table 20:	Summary	table of relevant re	eproductive to	oxicity studies

Study; Reference	Dose levels	NOAEL	Relevant effects
Multigeneration, rats Milburn G.; 2005b	0, 50, 250, 1500 ppm/diet (equivalent to 0, 4, 20 and 120 mg/kg bw	parental: 250 ppm (20 mg/kg bw) reproductive: 1500 ppm (120 mg/kg bw) developmental: 250 ppm (20 mg/kg bw)	Parental and offspring: -bodyweight ↓ -liver weight ↑
Developmental toxicity, rats Moxon M.; 2005a	0, 50, 200, 1000 mg/kg bw	maternal: 200 mg/kg bw developmental: 1000 mg/kg bw	maternal: plasma total protein ↓, total bilirubin ↓ albumin/globulin ratio ↑ developmental: no effects
Developmental toxicity, rabbits Moxon M.; 2005b	0, 50, 250, 1000 mg/kg bw	maternal: 1000 mg/kg bw developmental: 1000 mg/kg bw	maternal: no effects developmental: no effects

4.11.1 Effects on fertility

4.11.1.1 Non-human information

Effects on fertility were investigated in a multigeneration study in rats. Dietary administration of mandipropamid at a dose level of 1500 ppm for two successive generations did result in decreased bodyweights in F1 males during the pre-mating period. F0 and F1 females in the 1500 ppm group producing F1A and F2A litters respectively had slightly lower bodyweights on days 15 and/or 22 post partum, but these differences were no longer evident on day 29.

Bodyweight of F1A and F2B pups in the 1500 ppm group was reduced from day 15 onwards, but there were no effects on bodyweight in F2A pups.

The liver was identified as the target organ, increases in liver weight were seen in both sexes, both generations, in parents and in pups. The effects were confined to the 1500 ppm dose group.

There were no effects on implantation data or reproductive performance and no microscopic changes were observed in the reproductive system that could be related to mandipropamid.

The parental NOAEL for systemic toxicity can be considered at 250 ppm, equivalent to approximately 20 mg/kg bw/d. For pup developmental effects the NOAEL can be considered at 250 ppm also. The NOAEL for effects on reproduction was considered to be 1500 ppm, equivalent to approximately 120 mg/kg bw/d.

4.11.1.2 Human information

No information available from case reports, epidemiological studies, medical surveillance, reporting schemes and national poisons centres.

4.11.2 Developmental toxicity

4.11.2.1 Non-human information

Developmental toxicity tests of mandipropamid were performed in rats and rabbits.

Mandipropamid administered at a dose level of 50, 200 or 1000 mg/kg/day had no effect on the clinical condition of rats, on maternal bodyweight or food consumption and no treatment-related findings were observed at examination post mortem. Plasma total protein and total bilirubin were lower and albumin/globulin ratio higher in the rats given 1000 mg/kg/day. The maternal NOAEL can be considered at 200 mg/kg/day.

There was no effect of mandipropamid on the number, growth or survival of the foetuses in utero. There was no effect of mandipropamid on foetal development. Although major observations affecting the sternum were seen only in foetuses in the mandipropamid treated groups the incidence of foetuses affected was very small and not dose-related. Also, there were no minor changes in the appearance or ossification of the sternebrae to indicate that mandipropamid adversely affected this area of the skeleton and there was no evidence for an effect of mandipropamid on other ossification centres of the skeleton. The low incidence of major observations affecting the sternum was therefore considered to be incidental to treatment with mandipropamid. The NOAEL for developmental effects can be set at 1000 mg/kg bw/d.

In rabbits, there were no adverse effects of 50, 250 or 1000 mg/kg/day mandipropamid on the clinical condition, bodyweight or food consumption of the pregnant female rabbits. Therefore the NOAEL for maternal toxicity can be considered at 1000 mg/kg bw/d.

No effect on the number, growth, survival or development of the foetuses in utero has been observed.

The group mean values of foetuses with an incompletely ossified odontoid and incompletely ossified 5th sternebra observed in the intermediate and high dose groups were within the historical controls means. Therefore these differences are considered not to be related to treatment.

The NOAEL for developmental toxicity can be considered at 1000 mg/kg bw/d.

4.11.2.2 Human information

No information available from case reports, epidemiological studies, medical surveillance, reporting schemes and national poisons centres.

4.11.3 Other relevant information

No data available.

4.11.4 Summary and discussion of reproductive toxicity

Dietary administration of mandipropamid at a dose level of 1500 ppm for two successive generations did result in decreased bodyweights in F1 males during the pre-mating period. F0 and F1 females in the 1500 ppm group producing F1A and F2A litters respectively had slightly lower bodyweights on days 15 and/or 22 post partum, but these differences were no longer evident on day 29.

Bodyweight of F1A and F2B pups in the 1500 ppm group was reduced from day 15 onwards, but there were no effects on bodyweight in F2A pups.

The liver was identified as the target organ, increases in liver weight were seen in both sexes, both generations, in parents and in pups. The effects were confined to the 1500 ppm dose group.

There were no effects on implantation data or reproductive performance and no microscopic changes were observed in the reproductive system that could be related to mandipropamid.

The parental NOAEL for systemic toxicity can be considered at 250 ppm, equivalent to approximately 20 mg/kg bw/d. For pup developmental effects the NOAEL can be considered at 250 ppm also. The NOAEL for effects on reproduction was considered to be 1500 ppm, equivalent to approximately 120 mg/kg bw/d.

In a teratogenicity study in rats, mandipropamid administered at dose levels of 1000 mg/kg/day had effects on plasma total protein and total bilirubin levels and the albumin/globulin ratio. The maternal NOAEL can be considered at 200 mg/kg/day.

Although major observations affecting the sternum were seen only in foetuses in the mandipropamid treated groups the incidence of foetuses affected was very small and not dose-related. Also, there were no minor changes in the appearance or ossification of the sternebrae to indicate that mandipropamid adversely affected this area of the skeleton and there was no evidence for an effect of mandipropamid on other ossification centres of the skeleton. The low incidence of major observations affecting the sternum was therefore considered to be incidental to treatment with mandipropamid. The NOAEL for developmental effects can be set at 1000 mg/kg bw/d.

In a teratogenicity study in rabbits, there were no adverse effects of 50, 250 or 1000 mg/kg/day mandipropamid on the clinical condition, bodyweight or food consumption of the pregnant female rabbits. Therefore the NOAEL for maternal toxicity can be considered at 1000 mg/kg bw/d.

The group mean values of foetuses with an incompletely ossified odontoid and incompletely ossified 5th sternebra observed in the intermediate and high dose groups were within the historical controls means. Therefore these differences are considered not to be related to treatment.

The NOAEL for developmental toxicity can be considered at 1000 mg/kg bw/d.

4.11.5 Comparison with criteria

No effects on fertility or development were observed in studies conducted with mandipropamid, neither in a rat multigeneration study, nor in rat and rabbit developmental studies (according to both DSD and CLP).

4.11.6 Conclusions on classification and labelling

There is no evidence of effects on reproduction and development caused by mandipropamid, therefore, no classification is proposed.

4.12 Other effects

No other data available.

4.12.1 Non-human information

No other data available.

4.12.1.1 Neurotoxicity

Table 21: Summary table of relevant neurotoxicity studies

Study; Reference	Dose levels	NOAEL	Relevant effects
Acute neurotoxicity Milburn G.; 2005c	0, 200, 600, 2000 mg/kg bw	systemic: 2000 mg/kg bw neurotox.: 2000 mg/kg bw	No adverse effects of treatment and no evidence of neurotoxicity
Subchronic neurotoxicity Pinto P.; 2005c	0, 100, 500 and 2500 ppm/diet (7.4, 37.3, 192.5 mg/kg bw/d ♂ and 8.4, 41, 206.7 mg/kg bw/d ♀)	neurotox.: 2500 ppm (192.5 mg/kg bw/d ♂, 206.7 mg/kg bw/d ♀) systemic tox.: 500 ppm (37.3 mg/kg bw/d ♂, 41 mg/kg bw/d ♀)	no neurotoxic effects -bodyweight ↓ (♂) -bodyweight gain ↓ (♂) -liver weight ↑ (♂♀)

Mandipropamid has been assessed for potential neurotoxicity in an acute and a subchronic neurotoxicity study in the rat and was shown to have no neurotoxic potential in these studies.

In an acute neurotoxicity study, rats received a single oral dose of 0, 200, 600 or 2000 mg/kg bw mandipropamid via gavage. Detailed clinical observations, bodyweights and food consumption and a full range of functional assessments (FOB, including grip strength, tail flick, landing foot splay), brain weight and neuropathology revealed no treatment related effects of mandipropamid.

The NOAEL for systemic and neurotoxic effects in this study was established at 2000 mg/kg bw mandipropamid.

In a subchronic neurotoxicity study, mandipropamid was administered to groups of 12 male and 12 female rats at dose levels of 0, 100, 500 and 2500 ppm (equivalent to 7.4, 37.3 and 192.5 mg/kg bw/d for males and 8.4, 41 and 206.7 mg/kg bw/d for females) in the diet for 90 consecutive days. The highest dose of 2500 ppm resulted in toxicity characterised by reduced bodyweight and bodyweight gain in males and increased liver weight in both sexes.

A comprehensive battery of neurobehavioural tests and neuropathological examination of the central and peripheral nervous system showed no effects of treatment at doses of up to 2500 ppm mandipropamid.

The NOAEL for neurotoxic effects in this study was established at 2500 ppm for male and female rats (192.5 and 206.7 mg /kg bw/d for males and females respectively). The NOAEL for systemic toxicity can be considered at 500 ppm (37.3 and 41 mg/kg bw/d for males and females respectively).

4.12.1.2 Immunotoxicity

No data available.

4.12.1.3 Specific investigations: other studies

No data available.

- 4.12.1.4 Human information
- 4.12.2 Summary and discussion
- 4.12.3 Comparison with criteria
- 4.12.4 Conclusions on classification and labelling

CLH REPORT FOR MANDIPROPAMID

5 ENVIRONMENTAL HAZARD ASSESSMENT

5.1 Degradation

Table 22: Summary of relevant information on degradation

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Method	Results											Remarks	Reference
Hydrolysis ECCD 94/37/EC (1994), ECCD 95/36/EC (1995), OECD 111 (1981), US EPA: N: 161-1 (1982), BBA Merkblatt Nr. 55, Teil I und II (1980)	No degradati to 9.	on of mandi	No degradation of mandipropamid occurred, therefore, mandipropamid can be considered hydrolytic stable in a pH range of 4 to 9.	1, therefore,	mandiprop	amid can t	oe conside	ered hydro	lytic stabl	e in a pH 1	ange of 4		Buckel, T., 2002 NOA 446510 / 0018
Photolysis	Parent: (No metabolites > 10 % AR)	metabolites	> 10 % AR)										*Hand, L. H.,
* ECCD 94/37/EC (1994), ECCD 95/36/EC (1995), US EPA: N: 161-2 (1982), OECD (2000), SETAC (1995)	*Sterilized buffered solution, pH 7.0: DT ₅₀ = 1.4 days [14 C-methoxy-Ph]-label DT ₅₀ = 6.7 days [14 C-Cl-Ph]-	ays [¹⁴ C-label] ays [¹⁴ C-label]		*Natural summer light, 40° N: $DT_{50} = 1.6$ days [14 C-methoxy-Ph]-label $DT_{50} = 8.0$ days [14 C-Cl-Ph]-label	it, 40°N: Cl-Ph]-	**Sterilized natural water, pF 7.4: DT ₅₀ = 1.0 days [14 C- methoxy-Ph]-label DT ₅₀ = 0.9 days [14 C-Cl-Ph]-	ed natural) days [¹⁴ Ph]-label) days [¹⁴	**Sterilized natural water, pH 7.4: DT ₅₀ = 1.0 days [14 C- methoxy-Ph]-label DT ₅₀ = 0.9 days [14 C-Cl-Ph]-		**Natural summer light, 40° N: DT ₅₀ = 1.9 days [14 C- methoxy-Ph]-label DT ₅₀ = 1.5 days [14 C-Cl-	ler light, [2004;NOA 44610 / 0041
** ECCD 95/36/EC (1995), US EPA: N: 161-2 (1982), OECD (2000)	label					label			PnJ-label	lbel			** Harrison, C. L., 2004;NOA 446510 / 0197
Readily biodegradable OECD 301F (1992)	NO The degr	radation of 1 ion: Mandip	NO The degradation of mandipropamid technical was < 5 % after 28 days Conclusion: Mandipropamid is not considered readily biodegradable.	thnical was residered read	vas < 5 % after 28 days readily biodegradable.	28 days radable.							Wallace, S. J., 2002
Degradation in water /	Condi-	Lohol	Cerefore		Water		17	Sediment	nent	0L	Total		*Grosjean, J.,
	tions	Label	System	DegTs0	Degradation	DT ₅₀ DT ₆	9	Degradation	DegTon	DegTs ₀	Degradation		2005 NOA
OECD 308 (2002),		M. Dt	Calwich Abbey	Stable	Stable	3.44		4.41	14.7	10.3	28.7		446510 / 0388
SELAC (1995), HS EDA: N (1982)	Aorobio	Me-rn	Swiss Lake	234	<i>LLL</i>	4.93	30.4	5.72	19.0	14.1	41.3		**Hurt, A. D., Bremler: V. M.
US-EFA: N (1982)	Acronic	לם כ	Calwich Abbey	Stable	Stable	69.0	2.30	4.86	16.2	5.93	17.2		Bramley, Y. M.,
		CI-FII	Swiss Lake	Stable	Stable	14.1	46.8	69.7	25.6	25.9	61.9		Orosjean, J., Davison K
		Mo Dh	Calwich Abbey	Stable	Stable	96.0	3.18	3.04	10.1	4.55	11.7		2005 NOA
	An-	1410-1 11	Swiss Lake	Stable	Stable	8.33	27.7	4.84	16.1	15.7	37.4		446510 / 0395
	aerobic	Cl.Ph	Calwich Abbey	Stable	Stable	6.75	22.4	3.95	13.1	12.76	30.3		
			Swiss Lake	34.3	114	20.2	72.5	5.54	18.4	23.7	76.0		

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Method	Results										Remarks	Reference
Degradation in an outdoor			Water	_		Sediment	nent	Total	a			Oliver, R. G.,
aquatic sediment system		Degradation	ation	Dissipation	ıtion	Degradation	ation	Degradation	ation			Webb, J.,
No regulatory guideline		DegT ₅₀ [days]	DegT ₉₀ [days]	DT ₅₀ [days]	DT ₉₀ [days]	DegT ₅₀ [days]	DegT ₉₀ [days]	DegT ₅₀ [days]	DegT ₉₀ [days]			Edwards, P. A., 2005
existing, following ouidelines were taken into	CI-Ph	10.6	35.1	2.50	11.8	3.18	10.6	5.86	16.9			
consideration: OECD	Me-Ph	14.4	47.9	3.19	12.0	1.94	6.45	5.53	15.0			
(2002), SETAC (1995), US- EPA: N (1982)												
Degradation in soil Rate of Degradation in soil												Berdat, T.,
Laboratory studies	The laboratory soil degradation rate of mandipropamid was investigated in total 25 experiments using 5 soils with a wide range	radation rate	of mandipro	opamid wa	s investiga	ated in total	25 experin	nents using.	5 soils with	a wide range		Nicollier, G.
	of soil properties (pH, organic C, texture, origin) under varying test conditions: Under aerobic conditions, 19 – 25 °C incubation temperature, soil moisture contents close to field capacity and relevant application rates (150 – 675 g ai ha-1), mandipropamid degraded with an overall half-life time in a range of 12.6 – 93.1 days based on SFO kinetics (n = 15, R2 > 0.95). The degradation rate in the lab was strongly depending on the application rate, at higher dose rates degradation significantly slowed down. Based on averaged results for each soil type (n = 5), DT50 values were in a range of 30.6 – 85.7 days with an arithmetic mean of 53.0 days. Under anaerobic conditions degradation of mandipropamid was significantly slower, DT50 was in a range of 158 – 179 days (based on SFO and DFOP kinetics, n = 2), mean DT50 = 169. Respective DT90 value was 758 days. No degradation rates for anaerobic metabolites could be obtained owing to their low occurrence.	organic C, tes ns, 19 – 25 ° (n = 15, R2 > dation signifi – 85.7 days vions degrada OP kinetics, 1	cture, origin C incubation 11), mandipr c 0.95). The cantly slow with an arith tion of man $n = 2$, mear ined owing 1	n temperational desperational descriptional	trying test- ire, soil m graded wi in rate in tl 3ased on a in of 53.0 1 was sign 69. Respe	conditions: oisture con th an overa he lab was veraged res days. ifficantly sk ctive DT9C ce	tents close I Il half-life ti strongly dep sults for eac wer, DT50	to field caps ime in a ran pending on t h soil type (was in a ra 758 days. N	city and rele ge of 12.6 – he applicati n = 5), DT5 nge of 158 – o degradati	evant 93.1 days on rate, at 0 values - 179 days		2005 a - b
Field studies ECCD 94/37/EC (1994),	The field dissipation rate of mandipropamid (as one single broadcast application of the formulation to bare soils) was investigated in 10 studies:	te of mandip es:	ropamid (as	one single	broadcast	application	ı of the forr	nulation to l	oare soils) w	as		Evans, P. 2003a 2003b
ECCD 99/36/EC (1935)	Dissipation of mandipropamid followed partly SFO (7 trials) and partly FOMC (3 trials) kinetics. Based on best fit (adjusted R2) DT50 was in a range of 2.0 – 29.2 days (adj. R2 \square 0.84) with an geometric mean of 13.6 days.	opamid follo ge of 2.0 – 29	wed partly {	SFO (7 tria j. R2 □ 0.8	ls) and pa '4) with ar	rtly FOMC	(3 trials) ki mean of 13	netics. Base 6.6 days.	d on best fit	(adjusted		2005a -g
Soil Photolysis SETAC (1995), OECD (2001), US-EPA: N: 161-3	In photolysis on the soil surface, mandipropamid degraded with an experimental half-life of $14.9 - 25.2$ days (n = 4), arithmetic mean = 19.5 days. This DT50 is equivalent to approx. $20 - 40$ midsummer days at 40 °N (arithmetic mean 30 days) No degradation rates for metabolites (all < 10 % of AR) could be obtained.	l surface, ma DT50 is equ etabolites (al	ndipropami ivalent to aj 1 < 10 % of	d degradec pprox. 20 - AR) could	l with an e - 40 midsu be obtain	xperimenta ımmer days ed.	l half-life o at 40 °N (a	f 14.9 – 25. trithmetic m	2 days (n = ² ean 30 days	t), arithmetic) No		Kuet, S. F., Dick, J., 2003

5.1.1 Stability

Aquatic hydrolysis

One hydrolysis study in sterile buffer solutions at pH values of 4, 5, 7 and 9 (50 °C over 7 days, 25 °C over 32 days) using ethyl labelled mandipropamid was carried out. No degradation of mandipropamid occurred, therefore, mandipropamid can be considered hydrolytic stable in a pH range of 4 to 9.

Aquatic photolysis

Aquatic photolysis of mandipropamid was investigated in sterile buffer solutions at pH 7 (n = 2) and in sterilized natural water (pH 7, n = 2) using Cl-phenyl and methoxy-phenyl labelled mandipropamid. All test systems were continuously irradiated with a xenon arc lamp (> 290 nm) to simulate natural light. Photo-degradation of mandipropamid was pronounced, the photolytic half-life under experimental conditions varied from 1.4 – 6.7 days in sterile buffer solutions and 0.9 – 1.0 days in sterilized natural water. Experimental half life times were calculated to correspond to 1.5 – 8.0 environmental midsummer days at a latitude of 40 °N. Under the influence of irradiation, mandipropamid was degraded to a large number of compounds, none of them exceeding 10 % AR individually. No distinct differences in degradation pattern were observed between sterilized buffer and natural water. Formation of CO2 (7.8 % of AR after 7 DAT)using Cl-phenyl mandipropamid was less pronounced indicating a higher stability of the Cl-phenyl moiety against photolysis. In contrast to the Cl-phenyl label, photo-degradation of methoxy-phenyl labelled mandipropamid resulted in extensive formation of multiple polar compounds, which were shown not to exceed 10 % AR individually.

In a separate study the **quantum yield of the direct photochemical degradation** of mandipropamid was investigated. This study was carried out in sterilized buffer solution at pH 7.4 and 25 °C. Mandipropamid was irradiated with a xenon arc lamp at 280, 300 and 330 nm over a test period of 12 hrs. Quantum yield was calculated to be $\Box = 0.492$ (at 280 nm) and $\Box = 0.370$ (at 300 nm). No adsorption owing to mandipropamid was considered at 330 nm (the study was complicated by impurities, which absorbed light at higher wave lengths). Using GC-SOLAR, the notifier calculated expected environmental half-lives of 39 - 71 days in summer and spring at 40 °N. The environmental half-life of mandipropamid, calculated on the basis of the quantum yield (GC-SOLAR), is distinct longer than directly measured in photolysis studies.

5.1.2 Biodegradation

5.1.2.1 Biodegradation estimation

No data available

5.1.2.2 Screening tests

Ready biodegradability of the active substance (OECD Annex IIA 7.7)

Reference: NOA446510 technical: Determination of 28 day ready biodegradability

Author(s), year: Wallace, S. J., 2002 Report/Doc. number: NOA 446510 / 0016 Guideline(s): OECD 301F (1992)

GLP: Yes
Deviations: No
Validity: Yes

Material and Methods:

Test substance: Mandipropamid technical, purity 97.1 %

Reference substance: Na-acetate

Inoculum: Activated sludge from local sewage treatment (30 mg L⁻¹ in each treatment)

Treatments: a) Blank control

b) Na-acetate (200 mg L⁻¹)

c) Mandipropamid techn. (100 mg L⁻¹)

d) Mandipropamid techn. (100 mg L⁻¹) + HgCl₂ (64 mg L⁻¹)

pH of all treatments approx. 7.5

Analysis: Chemical oxygen demand (COD)

Findings:

Table 33: Mean biodegradation of techn. mandipropamid and reference substances [% of added].

DAT	Na-acetate	Mandipropamid techn.	Mandipropamid techn. + HgCl ₂
5	68	< 5	< 5
9	73	< 5	< 5
15	78	< 5	< 5
20	78	< 5	< 5
28	77	< 5	< 5

The degradation of mandipropamid technical was < 5 % after 28 days.

Conclusion:

Mandipropamid is not considered readily biodegradable.

5.1.2.3 Simulation tests

Water/sediment studies

The following 2 studies were combined for discussion:

Reference: NOA 446510: Degradation in two aquatic sediment systems

(methoxyphenyl ring)

Author(s), year: Grosjean, J., Hurt, A. D., 2005

Report/Doc. number: NOA 446510 / 0388

Guideline(s): OECD 308 (2002), SETAC (1995), US-EPA: N (1982)

GLP: Yes
Deviations: None
Validity: Yes

Reference: NOA 446510: Degradation in two aquatic sediment systems

Author(s), year: Hurt, A. D., Bramley, Y. M., Grosjean, J., Davison, K., 2005

Report/Doc. number: NOA 446510 / 0395

Guideline(s): OECD 308 (2002), SETAC (1995), US-EPA: N (1982)

GLP: Yes
Deviations: None
Validity: Yes

Dark water/sediment studies (n = 8) were conducted under aerobic and anaerobic conditions with two contrasting natural systems, Calwich Abbey and Swiss Lake. Both sites were sampled individually for each label tested, therefore, main properties of water and sediment for Me-Ph and Cl-Ph labelled experiments were slightly different. However, these slight differences are not considered to significantly affect the comparability of the two labels used. The Calwich Abbey represents a loamy sediment rich in organic and microbial C (mean of both samplings: pH of sediment 7.1, silt loam, 5.7 % organic C, 845 µg microbial C g^{-1}), the sandy Swiss Lake sediment is considered as nutrient poor system (pH of sediment 4.8, sand, 0.8 % organic C, 124 µg microbial C g^{-1}). Experiments were conducted at 20 °C in the dark for a period of 120 days (Me-Ph label) and 365 days (Cl-Ph label).

Owing to the high organic C content and high microbial biomass in the Calwich Abbey system, overall degradation of mandipropamid was significantly faster in this system compared to the less active Swiss Lake system.

In general, mineralization of mandipropamid to ¹⁴CO₂ was significantly higher for Me-Ph labelled than for Cl-Ph labelled mandipropamid. One hundred days after onset of the experiment conducted under aerobic conditions, 30.5 - 35.5 % of AR were released as ¹⁴CO₂ from Me-Ph labelled mandipropamid, respective amounts for Cl-Ph labelled mandipropamid were only 3.9 – 4.3 % of AR. Under anaerobic conditions similar amounts of ¹⁴CO₂ were released from Me-Ph labelled mandipropamid (32.0 – 38.6 % of AR after 100 days), from Cl-Ph labelled mandipropamid only negligible amounts of ¹⁴CO₂ (0.4 – 2.5 % of AR) were released. These results clearly indicate that the Cl-Ph moiety of mandipropamid is much more persistent in comparison to the Me-Ph moiety. A distinct amount of radioactivity not trapped in the experimental set-up (indicated by incomplete mass balance), was attributed to the formation of methane (based on results from additional experimental work). The formation of methane (16.5 – 42.2 % of AR by 100 DAT, based on the difference of applied and recovered radioactivity) was much higher under anaerobic conditions and was almost exclusively attributed to the Me-Ph moiety of mandipropamid. Using Cl-Ph labelled mandipropamid the recovery of applied recovery was almost complete, formation of methane considered negligible.

In contrast to the release of $^{14}\text{CO}_2$, formation of NER was hardly depending on the label position used, under aerobic conditions maximum levels of 36.5 - 48.1 % of AR (62 to 100 days after onset) were observed, with a decreasing tendency thereafter. Anaerobic incubation led to significantly smaller formation of NER, maximum amounts of 16.2 - 30.9 % of AR (30 to 120 days after onset) were observed irrespective of the label used.

Dissipation of mandipropamid from the water layer was rapid (following SFO kinetics), dissipation half-life varied from 0.7-14.1 days (arithmetic mean 5.8 days) under aerobic and 1.0-20.2 days (arithmetic mean 9.1 days) under anaerobic conditions. Dissipation of mandipropamid was more pronounced in the Calwich Abbey system reach in organic C, indicating that dissipation from the water layer into the sediment was predominately driven by the high K_{FOC} of mandipropamid. In fact, degradation in the water layer was calculated to be negligible. The high stability of mandipropamid in the water under dark conditions could be expected from the hydrolysis studies (sterile conditions), but was also demonstrated in additional experiments using only the (non-sterilized) water layers of both water/sediment systems without sediment. Concluding, degradation in the water is considered to be negligible for PEC_{SW} and PEC_{SED} calculations using FOCUS surface water STEP 2 and 3.

Subsequent degradation of mandipropamid in the sediment phase was fast, $DegT_{50}$ varied from 4.4 – 7.7 days (arithmetic mean 5.7 days) under aerobic and 3.0-5.5 days (arithmetic mean 4.3 days) under anaerobic conditions. Degradation was slightly slower in the nutrient poor Swiss Lake system, different labels used did not differ from each other. Based on the aerobic water/sediment studies, a mean $DegT_{50}$ in the sediment of 5.7 days (n=4) was considered appropriate for PEC_{SW} and PEC_{SED} using FOCUS surface water STEP 2 and 3 calculations. As mentioned above, mandipropamid significantly dissipated into the sediment at a maximum level of 64.0 % of AR after 1 day of incubation.

Table 55: Summary of DT50 and DT90 [days] for the dissipation and degradation of mandipropamid in laboratory water/sediment systems (under aerobic and anaerobic conditions) and in one outdoor pond study (both labels, values shaded in grey were used for PECSW and PECSED calculations).

				Wa	ter		Sedin	nent	To	tal
Condi-	Label	System ^a	Degra	dation	Dissip	oation	Degrae	dation	Degra	dation
tions	Label	System	DegT ₅₀	DegT ₉₀	DT ₅₀	DT ₉₀	DegT ₅₀	DegT ₉₀	DegT ₅	DegT ₉₀
Aerobic	Me-Ph	Calwich Abbey	Stable	Stable	3.44	19.3	4.41	14.7	10.3	28.7
water/		Swiss Lake	234	777	4.93	30.4	5.72	19.0	14.1	41.3
sediment	Cl-Ph	Calwich Abbey	Stable	Stable	0.69	2.30	4.86	16.2	5.93	17.2
		Swiss Lake	Stable	Stable	14.1	46.8	7.69	25.6	25.9	61.9
Arithmetic	mean		Stable	Stable	5.79	24.7	5.67	18.9	14.1	37.3
Geometric n	nean		Stable	Stable	3.58	15.9	5.54	18.4	12.2	33.5
Anaerobi	Me- Ph	Calwich Abbey	Stable	Stable	0.96	3.18	3.04	10.1	4.55	11.7
с	rn	Swiss Lake	Stable	Stable	8.33	27.7	4.84	16.1	15.7	37.4
water/ sediment	Cl-Ph	Calwich Abbey	Stable	Stable	6.75	22.4	3.95	13.1	12.76	30.3
		Swiss Lake	34.3	114	20.2	72.5	5.54	18.4	23.7	76.0
Arithmetic	mean		Stable	Stable	9.06	31.4	4.34	14.4	14.2	38.9
Geometric n	nean		Stable	Stable	5.75	19.4	4.24	14.1	12.1	31.7
Outdoor	Cl-Ph		10.6	35.1	2.50	11.8	3.18	10.6	5.86	16.9
pond	Me-		14.4	47.9	3.19	12.0	1.94	6.45	5.53	15.0

				Wa	ter		Sedir	nent	To	tal
Condi-	Label	System ^a	Degra	dation	Dissip	oation	Degrae	lation	Degra	dation
tions	Label	System	DegT ₅₀	DegT ₉₀	DT ₅₀	DT ₉₀	DegT ₅₀	DegT ₉₀	DegT ₅	DegT ₉₀
	Ph									
Arithmetic	mean		12.5	41.5	2.85	11.9	2.56	8.53	5.70	16.0
Geometric r	nean		12.4	41.0	2.82	11.9	2.48	8.27	5.69	15.9
Overall ari	thmetic m	ean	-b	-	6.51	24.8	4.52	15.0	12.4	33.6
Overall geor	metric mea	an	_b	-	4.13	16.2	4.24	14.1	10.5	28.2

^a Properties of the water/sediment systems were not identical for the labels used (considered by RMS not to drastically affect risk assessment).

Under aerobic conditions, 3 metabolites were observed > 10 % of AR (major) in the total water/sediment system: SYN 504851 (38.5 % of AR after 100 days), SYN 521195 (17.7 % of AR, 14 days) and SYN 539678 (12.6 % of AR, 21 days). One further metabolites, SYN 500003, was close to 10 % of AR (9.4 % of AR) and is more toxic than the parent according to the acute oral toxicity in rats (Pooler, 2006). SYN 500003 was therefore considered relevant for further risk assessment. The minor metabolites SYN 536638 and SYN 539679 did not exceed 8.4 % of AR in sum (metabolites co-eluted in HPLC). In the water phase, only the polar metabolite SYN 504851 exceeded 10 % of AR, the polar metabolite SYN 500003 occurred close to 10 % of AR in the water phase. All other, less polar metabolites were mainly attributed to the sediment phase. Amounts of SYN 504851 steadily increased during incubation, indicating a high stability of this degradate in both, the water and sediment phase. Degradation half-lives of SYN 521195, SYN 539678 and SYN 500003 in the total system were calculated to be 10.0, 29.1 and 43.7 days, respectively. Formation pattern of metabolites were similar between both labels used (SYN 504851 and SYN 500003 can only be observed using Cl-Ph labelled mandipropamid).

^b Considered as stable under dark conditions but not under environmental conditions (irradiation).

Proposed pathway of mandipropamid in water/sediment.

Pattern of metabolites observed under anaerobic conditions were similar to patterns observed under aerobic conditions, however, almost all metabolites occurred at higher levels under anaerobic conditions (all > 10 % of AR). The major metabolite SYN 504851, representing the main sink of radioactivity, steadily increased to maximum levels of 73.5 % of AR 100 days after onset of the experiment. In one experiment, levels of SYN 504851 decreased to 45.5 % of AR after 365 days. The metabolites SYN 539678, SYN 500003, SYN 521195 and the sum of SYN 536638/SYN 539679 reached maximum levels of 29.3 % (30 days), 25.9 % (45 days), 15.4 % (14 days) and 11.7 % (10 days) of AR. None of these metabolites was persistent, DegT₅₀ for the total system was 19.2, 28.5, 11.3 and 14.9 days, respectively. Similar to the experiments conducted under aerobic conditions, only the polar metabolites SYN 504851 and SYN 500001 exceeded 10 % of AR in the water phase.

Table 56: Observed maximum occurrence [% of AR] of mandipropamid in the sediment and of metabolites in water/sediment systems (based on individual replicates, data stated in brackets give day of maximum occurrence, values shaded in grey were used for PECSW and PECSED calculations).

Compart ment	Water/sediment study	Mandi- propamid	SYN 504851	SYN 521195	SYN 539678	SYN 500003
	Aerobic	-	38.5 (100)	17.7 (14)	12.6 (21)	9.4 (45)
Total	Anaerobic	-	73.5 (100)	15.4 (14)	29.3 (30)	25.9 (45)
	Outdoor pond	•	11.1 (120)	10.8 (15)	6.9 (15)	6.4 (36)
	Aerobic	-	22.7 (100)	3.4 (21)	1.9 (42)	9.4 (45)
Water	Anaerobic	-	56.7 (179)	6.0 (30)	9.2 (30)	20.2 (45)
	Outdoor pond	T.	7.2 (120)	nd ^a	nd	5.4 (36)
	Aerobic	64.0 (1)	28.5 (100)	15.6 (14)	11.2 (30)	3.9 (10)
Sediment	Anaerobic	53.6 (1)	45.9 (100)	12.5 (21)	20.1 (30)	11.6 (30)
	Outdoor pond	23.7 (2)	4.5 (71)	10.8 (15)	6.9 (15)	1.7 (71)

a nd denotes not detected

Table 57: Summary of degradation half-lives [days] of metabolites in the total system of investigated aquatic systems (values shaded in grey were used for PECSW and PECSED calculations).

Aquatic s	ystem	Label	SYN 504851	SYN 521195	SYN 539678	SYN 500003
	Calwich	Me-Ph	_ ^a	9.7	19.7	-
Aerobic	Abbey	Cl-Ph	No decline	4.1	35.1	13.2
water/sediment	Swiss Lake	Me-Ph	-	10.1	36.9	-
	SWISS Lake	Cl-Ph	No decline	16.2	25.7	74.1
Arithmetic mean	1		No decline	10.0	29.4	43.7
Geometric mean			No decline	8.98	28.5	31.3
	Calwich	Me-Ph	-	10.6	19.7	-
Anaerobic	Abbey	Cl-Ph	441	6.3	23.0	34.2
water/sediment	Swiss Lake	Me-Ph	-	13.2	12.0	-
	SWISS Lake	Cl-Ph	No decline	15.4	23.0	23.0
Arithmetic mean	1		No decline	11.4	19.4	28.6
Geometric mean			No decline	10.8	18.8	28.1
Outdoor	nond	Me-Ph	-	28.4	73.3	-
Outubbi	ponu	Cl-Ph	No decline	21.2	71.0	No decline
Arithmetic mean	1		No decline	24.8	72.2	No decline
Geometric mean		_	No decline	24.5	72.1	No decline
Overall arithmet	tic mean		No decline	13.5	33.9	36.1
Overall geometric	mean		No decline	11.8	29.0	29.6

^a Not detectable using this label

The degradation behaviour of mandipropamid was additionally elucidated in one **outdoor pond study**, which was designed to account for the impact of natural light and water plants in order to simulate more realistic environmental conditions. As demonstrated in the aquatic photolysis studies, mandipropamid is rather photosensitive, therefore, a higher overall degradation rate of mandipropamid may be expected under outdoor conditions.

The study was carried out in open ponds (1.8 x 1.0 m, approx. 0.3 m of water depth) with two labels (Me-Ph and Cl-Ph, no replicates). Sediment depth was 0.1 m. The pond was partly planted with water plants. Mandipropamid was applied in July (150 g ai ha⁻¹), the duration of the experiment was 120 days. The sediment used was a sandy clay loam, pH 7.2, 1.9 % organic C and 85 μ g microbial C g⁻¹.

From total radioactivity recovered, it can be concluded that approx. 45 % (Cl-Ph label) and 70 % (Me-Ph label) of AR were released as CO_2 (and other volatiles, likely methane) after 120 days. However, CO_2 or other volatiles were not trapped in the test system, therefore, these numbers remain indicative. Highest amounts of NER could be found at study end with levels of 27.1 - 29.2 % of AR.

Degradation half-life of mandipropamid in the total outdoor pond system was 5.7 days following SFO kinetics (without normalization, $R^2 > 0.95$, no difference between labels). In contrast to the dark laboratory water/sediment studies, mandipropamid is also considered to show significant degradation in the water layer (calculated $DegT_{50} = 12.5$ days), likely owing to the influence of irradiation. In consequence, dissipation from the water phase was fast with an arithmetic mean half-life of 2.9 days. In the sediment layer mandipropamid degraded with a mean half-life of 2.6 days (maximum amounts occurring in the sediment were 23.7 % of AR after 2 days). The comparability of degradation rates between the laboratory water/sediment studies and the outdoor pond studies is clearly restricted due to the non-normalized conditions of the outdoor pond study. In contrast to the dark water/sediment studies, only the metabolites SYN 504851 and SYN 521195 exceeded 10 % of AR in the total outdoor pond system (11.1 % or AR at study end and 10.5 % of AR after 15 days, respectively). SYN 504851 was almost stable in the system (no decrease during study), $DegT_{50}$ of SYN 521195 was calculated to be 13.5 days.

Route of Degradation in soil

The route and rate of laboratory soil degradation of mandipropamid was extensively investigated in total 25 experiments using 5 soils with a wide range of soil properties (pH, organic C, texture, origin), under varying test conditions (temperature, ai concentration, soil moisture content) and varying incubation conditions (aerobic, anaerobic, sterile). For purposes of an environmental fate assessment only those studies conducted under more realistic conditions (aerobic conditions, 150 - 675 g ai ha⁻¹, moisture conditions close to pF2 and temperatures between 19 - 25 °C) were considered appropriate (which gives a final number of 15 relevant degradation experiments). For each soil type, on which multiple studies were conducted (e.g. dose rate experiments), arithmetic mean DT_{50} and DT_{90} values were used to avoid bias towards any particular soil type.

Degradation of mandipropamid in soil is considered to be mainly driven by soil microbial activity and by photolysis if located close to or on the soil surface. No degradation is observed under sterile conditions. The following degradation processes are considered to mainly attribute to the overall metabolism of mandipropamid in viable soils:

- Hydrolytic cleavage of the 2-propynyl moiety on the chloromandelic acid or methoxyphenyl ring (oxidative dealkylation)
- Hydrolytic methyl-ether cleavage (oxidative dealkylation)
- Reduction of a 2-propynyl moiety to a 2-propenyl moiety (reduction by acetylene hydratase)
- Hydrolytic cleavage of the central amid bridge
- Addition of water to a 2-propynyl group

Under aerobic conditions mineralization of mandipropamid to CO_2 accounts for 9.0-44.2% of AR after 120 days without significant differences between labels used (Cl-phenyl, methoxy-phenyl, ethyl). Formation of NER accounted for maximum 19.4-45.4% of AR by 120 days of incubation. Organic matter fractionation resulted in 5-10% of AR associated with fulvic acids, 5-13% with humic acids and 21-28% of AR bound to insoluble humins. Both ring systems were incorporated into the soil matrix.

Microbial degradation of mandipropamid in soil leads to formation of a large number of minor metabolites, all of them accounting for less than 5 % of AR with the exception of CGA 380778, which reached maximum levels of 6.3 % of AR in one experiment. Further identified minor metabolites (SYN 536638, NOA 458422, CGA 380775, SYN 500003, U7 and U8) were all observed at maximum amounts < 5 % of AR, none of them persistent. Several compounds or metabolite fractions close to 2 % of AR remained unidentified. Most identified minor soil metabolites (among them one unknown fraction observed in one sample at 5.1 % of AR) were included into groundwater risk assessment for reasons of precaution.

Anaerobic experiments were conducted using methoxy-phenyl and Cl-phenyl labels with one soil incubated aerobically for 30 days ('aging period') and waterlogged thereafter. After switching to anaerobic conditions, formation of CO_2 significantly slowed down (almost negligible after 120 days of anaerobic incubation), additional formation of NER during the anaerobic incubation phase accounted for 5.8 - 11.8% of AR. The degradation pattern of aerobic and anaerobic soil samples were similar, metabolites which were formed owing to reductive processes (e.g. SYN 536638) were also observed in aerobically incubated soils. This is thought to be due to micro-sites of soil aggregates, in which anaerobic conditions might occur. During the anaerobic phase of incubation no metabolite exceeded 5% of AR.

Soil photolysis of mandipropamid was investigated using methoxy-phenyl and Cl-phenyl labelled mandipropamid applied to dry and moist soil samples. On moist soil surfaces, mandipropamid degraded with a half-life of 27.5 days (Me-Ph label) and 40.2 days (Cl-Ph label) based on midsummer day equivalents at 40 °N. Mineralization of mandipropamid to $^{14}\text{CO}_2$ owing to irradiation was more pronounced for methoxy-phenyl labelled than for Cl-phenyl labelled mandipropamid, indicating that the methoxy-phenyl moiety is more sensitive to photo-degradation than the Cl-phenyl moiety. In dark control samples no degradation occurred. The metabolite pattern formed under soil photolysis were similar to the pattern formed in soils owing to microbial degradation. All metabolites observed were < 10 % of AR.

Rate of degradation (laboratory) in soil

(Note: The notifier based their risk assessment exclusively on the usage of arithmetic means (of DT_{50} values) instead of geometric means. This (non-recommended) approach gives more conservative mean DT_{50} values (endpoints) and is therefore accepted by the RMS.)

The laboratory soil degradation rate of mandipropamid could be best described applying first order multi compartment (FOMC) kinetics. This degradation behaviour likely results from weak enantiomer-selective degradation, which leads to a FOMC like degradation behaviour and which was definitively shown in one study investigating both enantiomers (R/S enantiomers) separately. However, enantiomer selectivity of the degradation in soil is weak observed with 1.2 to 1.7fold faster degradation of the R-enantiomer than the S-enantiomer. Therefore, enantiomer-selective degradation has no significant impact on the fate assessment of mandipropamid in soil and was taken into account further.

On the basis of simple first order (SFO) kinetics, mandipropamid degraded with an overall half-life time in a range of 12.6 - 93.1 days (n = 15, $R^2 > 0.95$), respective DT_{90} was in a range of 41.7 - 309 days. The degradation rate in the lab was strongly depending on the application rate, at higher dose rates degradation significantly slowed down. Based on averaged results for each soil type (n = 5), DT_{50} values were in a range of 30.6 - 85.7 days with an arithmetic mean of 53.0 days (respective DT_{90} values were 102 - 285 days, arithmetic mean 176 days). After transformation to standard conditions (20 °C and pF2), an arithmetic mean DT_{50} of 47.9 days was calculated based on the 5 soil types used.

Based on FOMC kinetics (best fit), arithmetic mean laboratory DT_{50} of the 5 soil types was calculated to be 47.7 days (range of 28.0 - 80.8 days), which is similar to the mean DT_{50} calculated using SFO kinetics (53.0 days). However, arithmetic mean DT_{90} was distinct longer with 346 days (in a range of 131 - 636 days). On the basis of FOMC kinetics it can be concluded, that mandipropamid might have the potential to accumulate in soil under unfavourable conditions. However, in field dissipation studies (n = 10) all DT_{90} values (according to best fit) were less than 365 days, indicating that the risk for accumulation of mandipropamid in soils can be considered low under environmental conditions.

Degradation half-life of the most pronounced soil metabolite, CGA 380778, maximum occurrence 6.3 % of AR, was obtained from parent studies (n = 15) and from degradation studies with the metabolite (n = 3). Overall DT₅₀ was in a range of 5.2 – 37.7 days. Averaged for each soil type (n = 5), an arithmetic mean DT₅₀ of 21.7 days (in a range of 8.5 – 36.7 days) was calculated (SFO kinetics). This value is equivalent to a normalized DT₅₀ of 20.0 days at 20 °C and pF2. Degradation rates of further minor soil metabolites were partly based on individual observations of these

metabolites in parent degradation studies or on separate metabolite degradation studies. All soil metabolites observed degraded faster than the parent.

Under anaerobic conditions degradation of mandipropamid was significantly slower, DT_{50} was in a range of 158 - 179 days (based on SFO and DFOP kinetics, n = 2), arithmetic mean DT_{50} was 169 days. Respective DT_{90} was 758 days. No degradation rates for anaerobic metabolites could be obtained owing to their low occurrence.

Under sterile conditions (n = 1) no degradation of mandipropamid was observed.

In soil photolysis experiments mandipropamid degraded with an experimental half-life of 14.9 - 25.2 days (n = 4), arithmetic mean 19.5 days, equivalent to approx. 20 - 40 midsummer days at 40 °N (arithmetic mean 30 days). No degradation rates for metabolites (all < 10 % of AR) could be obtained. Degradation rates of mandipropamid owing to soil photolysis were in a similar range compared to microbial degradation under aerobic conditions, indicating that soil photolysis does not significantly contribute to the overall dissipation of mandipropamid in soils.

Field dissipation studies

Ten representative bare ground field dissipation studies were conducted with mandipropamid (broadcast spray application) on a representative range of soil types from northern to southern Europe from 2002 - 2004. The trials included eight trials with a single application of 200 g ai ha⁻¹ and two trials in Switzerland that were carried out with a single application of 300 and 700 g ai ha⁻¹. The latter trials were conducted to evaluate dissipation under worst case conditions.

The dissipation of mandipropamid in these studies was consistent with degradation, since the decline of residues levels in the 0-10 cm increment was not associated with a significant increase in residue levels in the 10-20 and 20-30 cm depth layer (only trace levels could be found close to the LOQ) and volatilization from soil is considered to be negligible. Dissipation of mandipropamid followed partly SFO (7 trials) and partly FOMC (3 trials) kinetics. Based on best fit, DT_{50} was in a range of 2.0-29.2 days ($R^2>0.87$, adj. $R^2\geq0.84$) with an arithmetic mean of 17.0 days. Respective DT_{90} values were 42.1-240 days, arithmetic mean 92.8 days. The two longest DT_{90} values (199 and 240 days) were observed in field trials which were kept free of any vegetation (both located in Germany), the other field trials were covered by grass. Since DT_{90} of mandipropamid under field conditions is less than 1 year, field accumulation studies are not triggered. Residual amounts of mandipropamid in the ten field studies remaining in soil at trial termination (after 157 – 254 days) were in the range of 1.5-6.5% of nominal applied, indicating a low risk for accumulation. In contrast to the laboratory degradation studies, no dependence of degradation rate on the application rate was observed (one study in Switzerland).

Only low amounts of CGA 380778 were measured in the soil samples and these were within the top 0-10 cm soil layer (maximum 3 % of the nominal applied amount). For SYN 536638, residues could not be detected in the 0-10 cm layer at or above the LOQ. No residues of CGA 380778 or SYN 536638 could be detected in the 10-20 or 20-30 cm layer at all.

Field DT_{90} of mandipropamid applied once to the soil is less than 1 year and accumulation studies are not triggered therefore. Nevertheless, an accumulation study (Switzerland, conducted for 6 years) is currently being carried out to investigate the potential for accumulation of mandipropamid, when applied several times a year (up to 6 times as proposed for potatoes). Mandipropamid is applied as foliar spray each year at an application rate of 6 x 150 g ai ha with an interval of 6 – 7

days. The field trial is conducted and maintained according to GAP. Residue data are available for the first 3 years. Regarding to maximum levels observed no evidence of accumulation of mandipropamid can be deduced in the study, to date. However, residual amounts of mandipropamid detected in soil immediately before the 1st application of the 2nd and 3rd year indicated at least a limited potential for accumulation if applied several times in a year.

Summary: Degradation

Degradation in water:

Abiotic degradation:

Mandipropamid was hydrolytically rather stable at a pH range of 4 to 9 in the hydrolysis study.

Under the influence of irradiation, mandipropamid was rapidly photolytically degraded to a large number of compounds, none of them exceeding 10 % AR individually. Formation of CO2 was 7.8 % of AR after 7 DAT using Cl-phenyl mandipropamid. In contrast to the Cl-phenyl label, photodegradation of methoxy-phenyl labelled mandipropamid resulted in extensive formation of multiple polar compounds, which were shown not to exceed 10 % AR individually.

Degradation in water:

Biotic degradation

The results of a **readily biodegradability** study indicate, that mandipropamid is not readily biodegradable.

In **water/sediment** degradation of mandipropamid in the total system followed SFO kinetics was fairly rapid irrespective of the label used and of aerobic or anaerobic conditions. Degradation half-life varied from 5.9 - 25.9 days (n = 4, arithmetic mean 14.1 days) under aerobic and 4.6 - 23.7 days (n = 4, arithmetic mean 14.2 days) under anaerobic conditions.

The mineralization to CO2 was significantly higher for methoxy-phenyl labelled than for Cl-phenyl labelled mandipropamid. One hundred days after onset of the experiment conducted under aerobic conditions, 30.5 - 35.5 % of AR were released as 14CO2 from methoxy-phenyl labelled mandipropamid, respective amounts for Cl-phenyl labelled mandipropamid were only 3.9 - 4.3 % of AR.

Note: Aquatic toxicity studies for metabolites SYN 504851, SYN 536638 and SYN 536638 are available but are missing for SYN 521195 and for the major metabolites in the sediment SYN521195 and SYN 539678. Thus a reliable classification regarding the hazardous to aquatic environment for all degradation products is not possible.

Degradation in water:

In an **outdoor pond study** degradation half-life of mandipropamid in the total outdoor pond system was 5.7 days following SFO kinetics (without normalization, R2 > 0.95, no difference between labels). In contrast to the dark laboratory water/sediment studies, mandipropamid is also considered to show significant degradation in the water layer (calculated DegT50 = 12.5 days), likely owing to the influence of irradiation. In consequence, dissipation from the water phase was fast with an arithmetic mean half-life of 2.9 days. In the sediment layer mandipropamid degraded with a mean half-life of 2.6 days. From total radioactivity recovered, it can be concluded that approx. 45 % (Cl-Ph label) and 70 % (Me-Ph label) of AR were released as CO2 (and other volatiles, likely methane) after 120 days.

Note: Aquatic toxicity studies for metabolite SYN 504851 is available but is missing for SYN 521195. Thus a reliable classification regarding the hazardous to aquatic environment for all degradation products is not possible.

<u>Degradation of mandipropamid in soil</u> is considered to be mainly driven by soil microbial activity and by photolysis if located close to or on the soil surface.

Under aerobic conditions DT50 values were in a range of 30.6 - 85.7 days with an arithmetic mean of 53.0 days (n = 5). Mmineralization of mandipropamid to CO2 accounts for 9.0 - 44.2 % of AR after 120 days without significant differences between labels used (Cl-phenyl, methoxy-phenyl, ethyl). Formation of NER accounted for maximum 19.4 - 45.4 % of AR by 120 days of incubation. Under anaerobic conditions degradation of mandipropamid was significantly slower, DT50 was in a range of 158 - 179 days (based on SFO and DFOP kinetics, n = 2), mean DT50 = 169.

The dissipation of mandipropamid in soil followed partly SFO (7 trials) and partly FOMC (3 trials) kinetics. Based on best fit, DT50 was in a range of 2.0 - 29.2 days (R2 > 0.87, adj. R2 3 0.84) with an arithmetic mean of 13.6 days.

Conclusion:

Mandipropamid is not readily biodegradable under test conditions within 28 days. Ultimate degradation could not be shown in discussed abiotic and biotic degradation studies. Available aquatic degradation studies with the exeption of the Hydrolysis study indicate primary degradation, but due to missing data on aquatic toxicity of degradants, it is not possible to show that the metabolites are not classifiable, therefore a non rapid degradation is proposed.

5.2 Environmental distribution

5.2.1 Adsorption/Desorption

Reasonable adsorption/desorption coefficients (K_{FOC} , 1/n values) were determined for methoxyphenyl labelled mandipropamid in seven soil batch experiments using four EU and three US soils with a representative spectrum of soil properties. K_{FOC} values were in a range of $405-1294~L~kg^{-1}$, with an arithmetic mean of $847~L~kg^{-1}$. Respective 1/n values were in a range of 0.80-0.92 with an arithmetic mean of 0.85. Adsorption of mandipropamid was strongly correlated with the organic matter content of the soil. No dependency on either pH of soil or other soil characteristics was observed.

Valid batch experiments were also conducted on a large number of soil and water/sediment metabolites of mandipropamid using unlabelled and/or labelled test compounds (CGA 380778, CGA 380775, SYN 536638, SYN 539678, SYN 521195, SYN 500003 and SYN 504851). Arithmetic mean K_{FOC} values of less polar metabolites (all with the exception of SYN 500003 and SYN 504851) were in the range of 448 - 1677 L kg⁻¹, indicating a medium to low mobility of these metabolites in soil. The two polar metabolites SYN 500003 and SYN 504851 exhibited mean K_{FOC} values of 11 and 5 L kg⁻¹, indicating very high mobility in soils. Mean 1/n value (0.76 – 0.92) gave evidence for non-linear adsorption/desorption isotherms for almost all metabolites with the exception of SYN 539678 (1/n = 1.00).

5.2.2 Volatilisation

Mandipropamid has a low vapour pressure of $< 9.4 \times 10^{-7}$ at 25 °C and a Henry's Law constant of $< 9.2 \times 10^{-5}$ at 25 °C. Therefore volatilisation of mandipropamid would be considered negligible. The low potential for volatilisation from soil and leaf surfaces was also demonstrated in two laboratory studies conducted according to BBA guidelines. Based on a theoretical calculation of the potential for photo-oxidation of mandipropamid in the atmosphere a first order half-life of 1.36 hrs was estimated. Concluding, air is not a likely route of environmental contamination.

5.2.3 Distribution modelling

Not relevant to classification

Summary: Evironmental Distribution (not relevant for classification and labelling)

Evironmental Distribution (not relevant for classification and labelling)				_
	Test guideline	р	GLP	Reliabili
	/ design	Н	(y/n)	ty
Adsorption/Desorption KFOC values were in a range of 405 – 1294 L kg-1, with an arithmetic mean of 847 L kg-1. Respective 1/n values were in a range of 0.80 – 0.92 with an arithmetic mean of 0.85.				
Volatilisation Henry's constant of $< 9.2 \times 10$ -5 Pa m³/mol (25° C) Vapour pressure of $< 9.4 \times 10$ -7 Pa (25° C)				

5.3 Aquatic Bioaccumulation

Table 58: Summary of relevant information on aquatic bioaccumulation

Method	Results	Remarks	Reference
Partition co-efficient ‡	$\log P_{O/W} = 3.2$ in pure water (990	Effect of pH (4-	Das R., (2003c)
(state temperature, pH and purity)	g/kg) at 25 °C	10): not relevant	(NOA446510/0027)
		as the active	Das R., (2006c)
		substance shows	(Doc. 10115098)
		no pH	
		dependency	
Determination of the accumulation	Mandipropamid accumulated in		Roberts, G., Peurou,
and elimination of Mandipropamid	whole fish with BCF values of 35 and		F., 2003
OECD 305	48 (the overall mean lipid content was		BL7579/B
	11 % w/w and was used for		
	normalisation of the BCF)		

5.3.1 Aquatic bioaccumulation

5.3.1.1 Bioaccumulation estimation

No estimation available

5.3.1.2 Measured bioaccumulation data

Reference: NOA446510: Determination of the accumulation and elimination of

[14C]NOA446510 in fathead minnow (Pimephales promelas)

Author(s), year: Roberts, G., Peurou, F., 2003

Report/Doc. number: BL7579/B Guideline(s): OECD 305

GLP: Yes

Deviations: None of relevance

Validity: Acceptable

Test substance: [14C] Mandipropamid (NOA446510), radiochemical purity: 99.5 %, batch:

ILA-208.9 and unlabelled Mandipropamid (NOA446510), purity 99 %,

batch: AMS 1131/2

Material and methods:

Test species: Fathead minnow (Pimephales promelas)

Number of 88 fish per test concentration and solvent control

organisms:

Weight, length 3.9 (2.0 - 6.6) g, 51.5 (41.3 - 65.2) mm

Type of test, duration: Flow-through test

Applied

concentrations:

Nominal: 0 (solvent control), 3.2 and 32 μ g/L Measured (mean): -- (solvent control), 3.2 and 31 μ g/L

Solvent Dimethylformamide (DMF)

Test conditions:

Water quality: Dechlorinated tap water, hardness: 43 – 47 mg/L as CaCO₃

Temperature: 24.8 – 25 °C pH: 7.2 – 7.6

 O_2 content: > 60 %, $5.8 - 8.6 \text{ mg } O_2/L$

Light regime: 16 hours light / 8 hours darkness, 20 min transition period

Feeding
Test parameters:

Ecostart (proprietary fish food): 2 % of the total fish weight per day Concentration of [¹⁴C] Mandipropamid equivalents in fish tissues were determined by LSC-method at 3, 6, 12, 24, 48, 119, 167, 190 h (exposure) and 6, 25, 48, 97, 147, 195 h (depuration phase). Four fish for tissue analysis were removed from each test concentration and control at each sampling time. Additionally a lipid analysis (by chloroform/methanol extraction) was carried out on fish sampled at day 0 and day 7 (exposure) and at day 8 (depuration). TLC-analysis of fish tissues were performed at the end of exposure and depuration phase.

For chemical analysis (LSC) of NOA446510 in test solutions samples were taken at -48 h (pre-exposure phase), 0, 3, 6, 12, 24, 48, 119, 167, 190 h (exposure) and 6, 25, 48, 97, 147, 195 h (depuration phase)

Calculations/statistics:

BCF was calculated as ratio of [¹⁴C] Mandipropamid equivalents concentration in water and [¹⁴C] Mandipropamid equivalents

concentration of in fish tissues and as ratio of $k_{\text{d}}\,\text{and}\,k_{\text{u}}$ (rate constant k

was determined by KINETICS program)

Findings:

Analytical data – water:

The mean measured concentrations of [14 C]-NOA446510 equivalents were 100 % (low concentration) and 97 % (high concentration) of nominal. TLC analysis confirmed that NOA446510 was stable in the high test concentration (32 µg/L), 79 – 89 % was determined as active substance. In the low concentration (3.2 µg/L) some degradation was observed (maybe due to photolysis) and recoveries ranged from 42 to 82 %. It was attempted to minimize the degradation by covering some sensitive parts of the test apparatus with black plastic.

Analytical data – fish tissue (TLC):

Due to very low levels of [¹⁴C]-NOA446510 in fish tissues a characterisation and a quantification of active substance or degradation products were not possible.

Lipid content:

No differences between male and female fish were noted.

Mean lipid content on day 0, day 7 and at the end of depuration phase was 10, 11.8 and 9.8 % w/w, respectively. The overall mean lipid content was 11 % w/w and was used for normalisation of the BCF, assuming that the accumulation was wholly contained within the lipid.

Table 59: Uptake, bioconcentration and depuration of [14C]NOA446510 in the fathead minnow

			N	Iean concen	tration of	[14C]NOA	446510 (µg/	/kg)		
Hour	Vis	cera	Fl	lesh	Car	cass		Whole	body	
Hour	3.2	32	3.2	32	3.2	32	3.2 μ	ıg/L	32 µ	ıg/L
	μg/L	μg/L	μg/L	μg/L	μg/L	μg/L	μg/kg	BCF	μg/kg	BCF
				Uı	ptake phas	se				
3	100	1090	10	160	20	320	40	12.5	400	12.9
6	160	1850	10	190	20	370	50	15.6	610	19.6
12	330	3060	20	190	20	300	80	25	730	23.5
24	470	4270	40	250	50	640	130	40.6	1080	34.8
48	530	5330	20	260	60	560	100	31.3	1590	51.3
120	490	5750	40	340	50	790	110	34.4	1430	46.1
167	500	7600	20	340	50	650	90	28.1	1570	50.6
190	640	4860	20	300	50	690	130	40.6	1350	43.5
				Dep	uration ph	ase				·
6	320	3840	10	140	20	160	60	18.8	880	28.4
25	70	2160	ND	30	10	60	20	6.3	250	8.1

			N	Iean concen	tration of	[14C]NOA	446510 (μg/	kg)		
Hour	Vis	cera	Fl	lesh	Car	cass		Whole	body	
nour	3.2	32	3.2	32	3.2	32	3.2 μ	g/L	32 μ	ιg/L
	μg/L	μg/L	μg/L	μg/L	μg/L	μg/L	μg/kg	BCF	μg/kg	BCF
48	70	610	ND	20	10	50	20	6.3	130	4.2
97	20	320	ND	ND	ND	20	ND	< 3	60	1.9
147	20	130	ND	ND	ND	20	10	3	30	1.0
195	20	90	ND	ND	ND	20	ND	< 3	20	0.6

ND = Not detected: $< 20 \mu g/kg$ for $32 \mu g/L$ or $< 10 \mu g/kg$ for $3.2 \mu g/L$ test concentrations.

Table 60: Mean concentrations and measured BCF of Mandipropamid in fish during 190 hours exposure and percentage of elimination after 6 and 165 hours

			3.2 μg/I	.i			32 μg/L		
Parameter	Tissue	Mean	BCF	1	% nation	Mean	BCF	% Elin	nination
		(µg/kg)		6 h	165 h	(µg/kg)		6 h	165 h
	Viscera	526 ± 67.3	164	49	96	5885 ± 1200	184	31	98
	Flesh	28 ± 11	9	64	100	310 ± 38.3	10	53	100
Wet weight	Carcass	52 ± 4.47	16	64	100	673 ± 95.4	21	76	97
	Whole	112 ± 17.9	35*	46	100	1485 ± 115	48*	46	99
	body	112 = 17.5	33	10	100	1105 = 115	10	10	,,,
Lipid	Whole	1018 ± 163	318			13500 ± 1043	422		
content	body	1016 ± 103	310			13300 ± 1043	422		

^{*} The overall mean lipid content was 11 % w/w and was used for normalisation of the BCF

Conclusion:

Mandipropamid accumulated in whole fish with BCF values of 35 and 48. In viscera (non-edible) portions BCF values of 164 and 184 were determined. All BCF values are based on calculations with total 14 C-residues. Mandipropamid was stable under test conditions. The plateau concentration was reached after 48 hours. During the depuration period the 14C-residues were completely eliminated (99 – 100 % in whole fish) after 8 days. The depuration half-life (CT50) was < 1 days.

5.3.2 Summary and discussion of aquatic bioaccumulation

Based on determined BCF values (35 and 48) mandipropamid is considered to have a low bioaccumulation potential. Based on the LogKow (3.2)

CLH REPORT FOR MANDIPROPAMID

4 Aquatic toxicity

Table 61: Summary of relevant information on aquatic toxicity

		an armin his way							
Method	Results							Remarks	Reference
	Toet organism	Test condition	Time	Endpoint	Test	NOEC	$\mathrm{EC}_{50}/\mathrm{LC}_{50}$		
	rest organism				conc.	[mg/L]	[mg/L]		
OECD 203	Oncorhynchus mykiss Rainbow trout	static	96 hr	mortality	mm	≥ 2.9	> 2.9 ^{a)}		Volz 2002
OECD 203	Pimephales promelas Fathead minnow	static	96 hr	mortality	mm	≥ 6.04	> 6.04 ^{a)}		Peter 2002
OECD 203	Cyprinus carpio Common carp	flow through ^{b)}	96 hr	mortality	mm	> 2.0	> 2.0		Maynard & Woodyer 2004a
OECD 203	Cyprinus carpio Common carp	flow through	96 hr	mortality	mm	5.54	8.63		Matsuura 2005
US EPA OPPTS 850.1075	Cyprinodon variegatus Sheephead minnow	flow through	96 hr	mortality	mm	2.8	4.5		Palmer et al 2005a
EPA OPPTS 850.1400	Pinephales promelas Fathead minnow	flow-through	32 d	hatchability mortality growth	u	≥ 2.0 0.5 0.5	> 2.0 1.0 1.0		Maynard 2003/ yes
OECD 202	Daphnia magna Waterflea	static	48 hr	immobility	nom	5.0	7.1		Maynard & Woodyer 2004b
OECD 202, Part II	<i>Daplmia magna</i> Waterflea	semi static	21 d	mortality reproduction length	m	≥ 2.64 0.87 0.28	>2.64 2.64 0.87		Grade 2003/ yes
OECD 201	Selenastrum capricornutum Green alga	static	72 hr	biomass growth rate	mm	> 27.8	> 27.8 ^{a)}		Grade 2001a
OECD 201	Anabaena flos-aquae Blue alga	static	96 hr	biomass growth rate	mm	≥ 19.8	> 19.8 ^{a)}		Knauer 2002
OECD 221 (Draft October 2000)	<i>Lemna gibba</i> Duckweed	static	2 d	biomass growth rate	mm	3.0	> 4.4		Bätscher 2005
EPA OPPTS 850.1035	Americamysis bahia Saltwater mysid	flow through	96 hr	mortality	mm	0.58	1.7		Palmer et al 2005b
EPA OOPTS 850.1025	Crassostrea virginica Eastern oyster	f.	96 hr	shell deposition	mm	0.46	76.0		Palmer et al 2005c
Toct and passed activation to to to to T	מין שלייויים ליים מים מים שליים שליים מים מים מים מים מים מים שליים שלים	mon lonimon to (mm) bourseom	coitostacococo (mod	2					

Test conc.: test concentration based on mean measured (mm) or nominal (nom) concentration a highest tested concentration (maximal solubility of test the substance under test conditions)

5.4.1 Fish

5.4.1.1 Short-term toxicity to fish

Reference: Acute Toxicity Test of NOA446510 to Rainbow Trout (Oncorhynchus

mykiss) Under Static Conditions

Author(s), year: Volz, E., 2002

Report/Doc. number: 2023552 Guideline(s): OECD 203

GLP: Yes

Deviations: None of relevance

Validity: Acceptable

Test substance: Mandipropamid (NOA446510), purity: 96.5 %, batch: SEZ2BP007

Material and Methods:

Test species: Rainbow trout (*Oncorhynchus mykiss*)
Number of 7 fish per concentration and control

organisms:

Weight, length: 1.47 g (1.25 - 1.76 g), 5.6 cm (5.2 - 6.0 cm)

Type of test, Static test, 96 hours

duration: Applied

concentrations:

Nominal: 0 (control), 0 (solvent control), 0.36, 0.65, 1.2, 2.1, 3.8 mg/L

Measured (mean): -- (control), -- (solvent control), 0.32, 0.53, 0.96, 1.6, 2.9 mg/L; the highest

mean measured concentration represented the maximal solubility of

NOA446510 under test conditions

Solvent 0.1 mL/L Dimethylformamide (DMF)

Test conditions:

Water quality: Filtered and UV sterilized well water, hardness: 177 mg/L as CaCO₃

Temperature: 13.5 - 14 °C

pH: 7.9 – 8.0 (0 h), 8.3 (96 h)

 O_2 content: 91 - 100 %

Light regime: 16 hours light / 8 hours darkness

Test parameters: Mortality and sublethal effects were assessed after 2, 24, 48, 72 and

96 hours;

for chemical analysis (HPLC method) of NOA446510 in test solutions

samples were taken at 0 and 96 hours

Statistics: None

Findings:

Analytical data: The mean measured concentrations at the start and the end of the test were

80 - 83% and 69 - 78% of nominal, over study period overall mean

measured concentrations were 76 - 87 % of nominal.

Behavioural effects: None

Mortality: None in the control and at all treatment group after 2, 24, 48, 72 and

96 hours

Conclusion: LC_{50} (96 h) > 2.9 mg/L, NOEC \geq 2.9 mg/L based on mean measured conc.

Reference: Acute Toxicity Test of NOA446510 to Fathead Minnow (*Pimephales*

promelas) Under Static Conditions

Author(s), year: Peter, P., 2002

Report/Doc. number: 2023555
Guideline(s): OECD 203
GLP: Yes

Deviations: Temporarily the oxygen content fell below 60 % saturation after 72 hours

and an aeration of test media was required. However this had no adverse

influence on the results or quality of the study.

Validity: Acceptable

Test substance: Mandipropamid (NOA446510), purity: 96.5 %, batch: SEZ2BP007

Material and Methods:

Test species: Fathead Minnow (*Pimephales promelas*)
Number of 7 fish per concentration and control

organisms:

Weight, length: 0.23 g (0.18 - 0.32 g), 3.0 cm (2.6 - 3.3 cm)

Type of test, Static test, 96 hours

duration: Applied

concentrations:

Nominal: 0 (control), 0 (solvent control), 0.65, 1.2, 2.1, 3.8, 6.8 mg/L

Measured (mean): -- (control), -- (solvent control), 0.67, 1.16, 2.02, 3.53, 6.04 mg/L; the

highest mean measured concentration represented the maximal solubility of

NOA446510 under test conditions

Solvent 0.1 mL/L Dimethylformamide (DMF)

Test conditions:

Water quality: Filtered and UV sterilized well water, hardness: 181 mg/L as CaCO₃, from

72 hours on gently aerated

Temperature: 24.6 - 25.1 °C

pH: 7.7 – 7.4 (0 h), 7.5 – 7.6 (96 h)

 O_2 content: 69 – 101%, except at 72 hours when it dropped temporarily below 60 % for

the three highest test concentrations

Light regime: 16 hours light / 8 hours darkness

Test parameters: Mortality and sublethal effects were assessed after 2, 24, 48, 72 and

96 hours:

for chemical analysis (HPLC method) of NOA446510 in test solutions

samples were taken at 0 and 96 hours

Statistics: None

Findings:

Analytical data: Mean measured concentrations were 90 - 101 % (start) and 88 - 114 %

(end) of nominal, overall mean measured concentrations over the study

period were 89 – 107 % of nominal

Behavioural effects: None

Mortality: None in the control and at all treatment groups after 2, 24, 48, 72 and

96 hours

Conclusion: LC50 (96 h) > 6.04 mg/L, NOEC ≥ 6.04 mg/L based on mean measured

conc.

Reference: NOA446510: Acute toxicity to common carp (Cyprinus carpio) in a

flow-through test system

Author(s), year: Maynard, S. & Woodyer, J., 2004a

Report/Doc. number: BL7872/B Guideline(s): OECD 203

GLP: Yes

Deviations: None of relevance

Validity: Acceptable

Test substance: Mandipropamid (NOA446510), purity: 96.5 %, batch: SEZ2BP007

Material and Methods:

Test species: Common carp (*Cyprinus carpio*)
Number of 10 fish per concentration and control

organisms:

Weight, length: 0.9 g (0.68 - 1.17 g), 3.3 cm (3.0 - 3.5 cm)

Type of test, Flow through Limit-test, 96 hours

duration: Applied

concentrations:

Nominal: 0 (control), 0 (solvent control), 2.0 mg/L

Measured (mean): -- (control), -- (solvent control), 2.0 mg/L

Solvent 0.1 mL/L Dimethylformamide (DMF)

Test conditions:

Water quality: Dechlorinated tap water, hardness: 49.3 mg/L as CaCO₃

Temperature: 21.7 °C

pH: 7.49 – 7.61 (0 h), 7.34 – 7.62 (96 h)

 O_2 content: 74 - 101 %

Light regime: 16 hours light / 8 hours darkness

Test parameters: Mortality and sublethal effects were assessed after 0, 24, 48, 72 and

96 hours;

for chemical analysis (HPLC method) of NOA446510 in the test solution

samples were taken at 0, 48 and 96 hours

Statistics: None

Findings:

Analytical data: Mean measured concentration at 0, 48 and 96 hours was 100 % of nominal

at each sample time

Behavioural effects None

Mortality: None in the control and in all treatment levels after 0, 24, 48, 72 and

96 hours

Conclusion: LC50 (96 h) > 2.0 mg/L, NOEC $\geq 2.0 \text{ mg/L}$ based on mean measured conc.

Reference: A 96-hour Acute Toxicity Test of NOA 446510 (Mandipropamid) with

Common Carp

Author(s), year: Matsuura, T., 2005

Report/Doc. number: 93451 Guideline(s): OECD 203

GLP: Yes

Deviations: None of relevance

Validity: Acceptable

Test substance: Mandipropamid (NOA446510), purity: 96.5 %, batch: SEZ2BP007

Material and Methods:

Test species: Common carp (*Cyprinus carpio*)
Number of 10 fish per concentration and control

organisms:

Weight, length: 1.4 ± 0.16 g, 5.1 ± 0.18 cm Type of test, Flow through test, 96 hours

duration: Applied

concentrations:

Nominal: 0 (control), 0 (solvent control), 4.55, 5.92, 7.67, 10, 13 mg/L Heasured (mean): -- (control), -- (solvent control), 4.92, 5.54, 7.16, 9.3, 12 mg/L

Solvent 0.1 mL/L Dimethylformamide (DMF)

Test conditions:

Water quality: Dechlorinated tap water, hardness: 49.3 mg/L as CaCO₃

Temperature: 22.5 – 22.8 °C pH: 7.8 – 8.7

 O_2 content: > 60 %

Light regime: 16 hours light / 8 hours darkness

Test parameters: Mortality and sublethal effects were assessed after 3, 24, 48, 72 and

96 hours;

for chemical analysis (HPLC method) of NOA446510 in test solutions

samples were taken at 0, 48 and 96 hours

Statistics: LC50: Probit analysis, NOEC: directly from the raw data

Findings:

Analytical data: Mean measured concentrations were 90.8 - 97.4% (0 h) and 89.1 - 94.9%

(96 h)

Behavioural effects At 7.16 mg/L and higher concentrations loss of equilibrium, reduced

activity, surface swimming and lethargy were observed.

Mortality: See table below

Table 62: Cumulative mortality of carps (C. carpio) exposed to Mandipropamid

Mean measured		Cı	ımulative mortality (%)	
concentration (mg/L)	3 hours	24 hours	48 hours	72 hours	96 hours
Blank control	0	0	0	0	0
Solvent control	0	0	0	0	0
4.92	0	0	0	0	0
5.54	0	0	0	0	0
7.16	0	0	20	20	40
9.30	0	0	30	50	50
12.0	10	70	70	80	90

Mean measured		Cı	ımulative mortality (9	%)	
concentration (mg/L)	3 hours	24 hours	48 hours	72 hours	96 hours
		NOEC = :	5.54 mg/L		
		LC50 = 8.63 mg/L (95)	% CL 7.5 – 10.1 mg/L))	

Conclusion: LC50 (96 h): 8.63 mg/L, NOEC: 5.54 mg/L based on mean measured conc.

Reference: NOA446510: A 96-Hour Flow-Through Acute Toxicity Test with the

Sheephead Minnow (Cyprinodon variegatus)

Author(s), year: Palmer. S.J.; Kendall, T.Z. & Krueger, H.O., 2005a

Report/Doc. number: 528A-138

Guideline(s): US EPA OPPTS 850.1075

GLP: Yes

Deviations: None of relevance Validity: Acceptable.

Test substance: Mandipropamid (NOA446510), purity: 96.1 %, batch: SEZ3BP004

Material and Methods:

Test species: Sheephead minnow (*Cyprinodon variegatus*)

Number of 7 fish per concentration and control

organisms:

Weight, length: 0.23 g (0.11 - 0.45 g), 2.5 cm (2.1 - 3.0 cm)

Type of test, Flow-through test, 96 hours

duration: Applied

concentrations:

Nominal: 0 (control), 0 (solvent control), 1.3, 2.2, 3.6, 6.0, 10 mg/L Measured (mean): -- (control), -- (solvent control), 1.1, 1,8, 2.8, 4.3, 6.1 mg/L

solvent 0.1 mL/L Dimethylformamide (DMF)

Test conditions:

Water quality: Filtered natural seawater diluted with well water, salinity: 20 %

Temperature: 21.8 - 21.9 °C

pH: 8.2 - 8.3 (0 h), 8.3 (96 h)O₂ content: $> 60 \%, 7.2 - 8.0 \text{ mg O}_2/L$ Light regime: 16 hours light / 8 hours darkness

Test parameters: Mortality and sublethal effects were assessed after 3, 24, 48, 72 and

96 hours;

for chemical analysis (HPLC method) of NOA446510 in test solutions

samples were taken at 0, 24 and 96 hours

Statistics: LC₅₀: Probit analysis, NOEC: directly from raw data

Findings:

Analytical data: Overall mean measured concentrations were in the range of 61 - 85 % of

nominal concentrations.

Behavioural effects At 4.3 mg/L and next higher concentration sublethal effects like erratic

swimming, surfacing and lying on the bottom of the aquarium were

observed. Thus the NOEC is 2.8 mg/L

Mortality: See table below

Table 63: Cumulative mortality of sheephead minnow exposed to Mandipropamid

Measured		Cı	ımulative mortality (%)	
concentration (mg/L)	3 hours	24 hours	48 hours	72 hours	96 hours
Blank control	0	0	0	0	0
Solvent control	0	0	0	0	0
1.1	0	0	0	0	0
1.8	0	0	0	0	0
2.8	0	0	0	0	0
4.3	0	0	40	40	40
6.1	10	30	100	100	100
		NOLC =	2.8 mg/L		
		$LC_{50} = 4.5 \text{ mg/L } (95)$	%CL 2.8 – 6.1 mg/L)		

Conclusion: LC₅₀ (96 h) 4.5 mg/L, NOEC 2.8mg/L based on mean measured conc.

5.4.1.2 Long-term toxicity to fish

Reference: NOA446510 tech: Early-life stage toxicity test to the fathead minnow

(Pimephales promelas)

Author(s), year: Maynard, S., 2003

Report/Doc. number: BL7577/B

Guideline(s): EPA OPPTS 850.1400

GLP: Yes

Deviations: None of relevance

Validity: Acceptable

Test substance: Mandipropamid (NOA446510), purity: 96.5 %, batch: SEZ2BP007

Material and Methods:

Test species: Fathead minnow (*Pimephales promelas*)

Number of 4 replicates x 20 eggs per test concentration and control

organisms:

Age: Eggs < 24 hours

Type of test, Flow-through test, 28 days post hatch

duration: Applied

concentrations:

Nominal: 0 (control), 0 (solvent control), 0.13, 0.25, 0.5, 1.0, 2.0 mg/L Heasured (mean): -- (control), -- (solvent control), 0.13, 0.23, 0.48, 0.87, 1.8 mg/L

Solvent Dimethylformamide (DMF)

Test conditions:

Water quality: Dechlorinated tap water, hardness: 38 – 58 mg/L as CaCO₃

Temperature: 24 - 25.5 °C pH: 7.5 - 7.9

 O_2 content: > 60 %, 5.6 - 8.4 mg O_2/L Light regime: 16 hours light / 8 hours darkness

Feeding Larvae were fed with rotifers from hatch until day 7 of post hatch, on day 7

(post hatch) additionally brine shrimp eggs were offered and from day 8 (post hatch) on only brine shrimp were fed, feeding was performed 3 x daily from Monday – Friday and 2 x daily on weekend. From day 16 (post

hatch) on high protein pelleted fish food was fed ad libidum.

Test parameters: Mortality and abnormal appearance or behaviour were assessed daily, the

length and the weight were determined at test termination (endpoints:

hatchability, survival and growth of larvae).

For chemical analysis (HPLC method) of NOA446510 in test solutions

samples were taken at 0, 4, 10, 18, 24 and 32 days

Statistics: Hatchability and survival data: Steel's Many-One Rank Test (non-

parametric method), larval length and weight: ANOVA followed by Dunnett's test or Wilcoxon Rank Sum Test (in the case of heterogeneous

variance data)

Findings:

Analytical data: Overall mean measured concentrations were 87 - 100 % of nominal.

Biological Time to hatch: Hatching was completed at day 4.

observation: During hatching and until 8 day post hatch hatched fry were observed to be

smaller, less active and took longer to "swim up" at 1.0 and 2.0 mg/L. At days 14, 21 and 27 (post hatch) fry at the highest test concentration (2 mg/L) were slightly smaller and less active. At 0.13, 0.25 and 0.5 mg/L concentration levels and at controls all surviving fish appeared normal and

healthy during the test.

Effects: See table 64 and table 65

Table 64: Hatchability and survival

NOA446510 nominal in mg/L	Mean number of eggs at start	Mean number of larvae hatched	Hatch in %	Mean number of larvae surviving at 28 d post hatch	Survival in %
control	21.25	19.5	92	18.75	96
solvent control	19.75	19.25	97	19.25	100
0.13	21	20.25	96	20	99
0.25	17.5	16.75	96	16.5	99
0.5	20.25	19	94	18.	95
1.0	20.5	19	93	16.5	87*
2.0	20.5	18.5	90	12.25	66*

^{*} significant difference when compared to pooled control (p=0.05)

Table 65: Length and Weight

NOA446510 nominal in mg/L	Number of fish	Mean length (± SD) in mm	Relative SD in %	Mean weight (± SD) in mg	Relative SD in %
control	75	20.0 ± 1.92	10	138.0 ± 37.6	27
solvent control	77	19.4 ± 2.02	10	129.5 ± 39.0	30
0.13	80	19.7 ± 1.46	7	129.8 ± 30.0	23
0.25	66	20.2 ± 1.51	7	140.4 ± 34.3	24
0.5	72	19.3 ± 1.69	9	125.2 ± 34.0	27
1.0	66	19.0 ± 2.03*	11	119.8 ± 40.1*	33
2.0	49	18.2 ± 2.15*	12	106.8 ± 39.1*	37

^{*} significant difference when compared to pooled control (p=0.05)

Conclusion: Mortality: NOEC: 0.5 mg/L, LOEC: 1.0 mg/L;

Hatchability: NOEC \geq 2 mg/L, LOEC > 2 mg/l;

Growth (weight and length): NOEC: 0.5 mg/L, LOEC: 1.0 mg/L;

based on nominal concentrations

5.4.2 Aquatic invertebrates

5.4.2.1 Short-term toxicity to aquatic invertebrates

Reference: NOA446510: Acute toxicity to Daphnia magna

Author(s), year: Maynard, S. & Woodyer, J., 2004b

Report/Doc. number: BL7871/B Guideline(s): OECD 202 GLP: Yes

Deviations: None of relevance

Validity: Acceptable

Test substance: NOA446510, purity: 96.5 %, batch: SEZ2BP007

Material and Methods:

Test species: Waterflea (Daphnia magna)

Number of 4 replicates each with 5 daphnids per treatment

organisms:

Age: First instar < 24 hours old

Type of test, Static test, 48 h

duration: Applied

concentrations:

Nominal: 0 (control), 0 (solvent control), 0.63, 1.3, 2.5, 5.0, 10 mg/L Measured (mean): -- (control), -- (solvent control), 0.65, 1.3, 2.5, 4.9, 11 mg/L

Solvent 0.01 mL/L Dimethylformamide (DMF)

Test conditions:

Water quality: Elendt's M4 medium, hardness: 226 mg/L as CaCO₃

Temperature 20.0 - 20.2 °C

pH 7.93 – 8.07 (0 h), 7.98 – 8.01 (48 h)

 O_2 content: 99 %, 9.0 mg O_2/L

Light regime: 16 hours light / 8 hours darkness

Test parameters: Immobility was assessed after 24 and 48 hours. For chemical analysis

(HPLC method) of NOA446510 in the test media samples were taken at

test initiation (0 h) and termination (48 h).

Statistics: EC50: Binominal method, NOEC: directly from the raw data

Findings:

Analytical data: The mean measured concentrations at the start and end of the test were in

the range of 96 – 110 %, overall mean measured concentration ranged from

98 - 110 % of nominal.

Effects: After 48 hours no immobility was observed in the control, solvent control

and in test concentrations up to 5 mg/L, at 10 mg/L all daphnids (100 %) were immobile. Thus the NOEC was determined to be 5 mg/L and the

EC50 was calculated to be 7.1 mg/L (95% CL: 5 - 10 mg/L).

Conclusion: EC50 (48 h): 7.1 mg/L, NOEC: 5.0 mg/L based on nominal conc.

5.4.2.2 Long-term toxicity to aquatic invertebrates

Reference: Daphnia magna Reproduction Test: Effects of NOA 446510 on the

Reproduction of the Cladoceran Daphnia magna STRAUS in a Semi-

Static Laboratory Test

Author(s), year: Grade, R. 2003

Report/Doc. number: 2013604

Guideline(s): OECD 202, Part II

GLP: Yes

Deviations: None of relevance

Validity: Acceptable

<u>Test substance:</u> NOA446510, purity: $99 \pm 2\%$, batch: Kl-6380/1

Material and Methods:

Test species: Waterflea (*Daphnia magna*)

Number of 12 replicates each with one daphnid per treatment and control

organisms:

Age: First instar < 24 hours old

Type of test, Semi static test, 21 d, renewals of test solutions: 0, 2, 5, 7, 9, 12, 14, 16, 19

duration: days

Applied

concentrations:

Nominal: 0 (control), 0 (solvent control), 0.033, 0.10, 0.30, 0.90, 2.7 mg/L -- (control), -- (solvent control), 0.023, 0.085, 0.28, 0.87, 2.64 mg/L

Solvent 0.1 mL/L Dimethylformamide (DMF)

Test conditions:

Water quality: Elendt's M4 medium, hardness: 264 mg/L as CaCO₃

Temperature 20.1 - 22 °C

pH 7.8 - 8.3 (fresh solution), 8.2 - 8.6 (old solution)

 O_2 content: 90 - 99 % saturation

Light regime: 16 hours light / 8 hours darkness, 30 min transition period

Feeding Daily (except Sunday) with *Scenedesmus obliquus* suspension (1 x 10⁸

cells/L)

Test parameters: Survival, time to first brood, production of young and other sublethal

effects were controlled and recorded daily, the length of daphnids was

measured at the end of the exposure;

for chemical analysis (HPLC method) of NOA446510 in the test solution samples were taken from fresh (on days 7, 14, 19) and old solutions (on

days 2, 9, 16, 21) from each test concentration.

Statistics: EC₅₀ (mortality): Probit model, NOEC: Dunnett's Test (mortality),

Bartlett's Test (reproduction)

Findings:

Analytical data: The mean measured concentrations ranged from 69.7 - 97.8 %

Effects: See table 66

Table 66: Summary of effects of long-term exposure of NOA446510 on Daphnia magna

Treatment (mean measured concentrations in mg a.s./L)	Adult mortality at day 21 (%)	Mean time to first brood (days)	Mean number of live offspring per female over 21 days	Mean length of parent (μm)
Blank control	0	8.5	113	4086
Solvent control	0	8.4	100	4001
0.023	0	8.1	100	3952
0.085	0	8.3	107	3981
0.28	0	8.3	96	3945
0.87	0	8.3	91	3862*
2.64	0	11.0*	16*	3302*
NOEC	≥ 2.64 mg/L	0.87 mg/L	0.87 mg/L	0.28 mg/L
EC ₅₀			1.64 mg/l	

^{*} Statistically significantly different from control (p < 0.05)

Conclusion: Mortality adult: $NOEC \ge 2.64 \text{ mg/L}$, LOEC > 2.64 mg/L

Reproduction: NOEC: 0.87 mg/L, LOEC: 2.64, Length: NOEC 0.28 mg/L, LOEC 0.87 mg/L EC₅₀ (number of live offspring): 1.64 mg/L based on mean measured concentrations

5.4.3 Algae and aquatic plants

Reference: Growth inhibition test of NOA 446510 to green algae (Selenastrum

capricornutum) under static conditions

Author(s), year: Grade, R., 2001a

Report/Doc. number: 2013586 Guideline(s): OECD 201

GLP: Yes

Deviations: None of relevance

Validity: Acceptable

Test substance: Mandipropamid (NOA446510), purity: $99 \pm 2\%$, batch: KI-6380/1

Material and Methods:

Test species: Green alga (Selenastrum capricornutum)

Number of 1.03×10^4 cells/mL; 3 replicates of each concentration, 6 replicates of the

organisms: medium control Type of test, Static test, 72 h

duration: Applied

concentrations:

Nominal: 0 (medium control), 0 (solvent control), 1.25, 2.5, 5.0, 10, 20, 40 mg/L Measured (mean): -- (medium control), -- (solvent control), 1.15, 2.1, 4.2, 8.45, 17.9, 27.8

mg/L

Solvent 0.01 mL/L Dimethylformamide (DMF)

Test conditions:

Water quality: Alga culture medium (according to OECD guideline), hardness 28 mg/L as

CaCO₃

Temperature 22 ± 1 °C

pH Treatments: 7.9 – 8.1 (0 h), 8.5 – 9.0 (72 h); control: 7.9 (0 h), 9.0 (72 h) Incubation: Continuous illumination with cool white fluorescent light (approx. 8000

lux), orbital shaking at 150 rpm

Test parameters: Cell densities were measured using CytoFluor II (Fluorescence Multi-Well

Plate Reader), growth rate and biomass were determined after 24, 48 and 72 hours; for chemical analysis (HPLC with UV-detection) of test the substance, samples of test solution were taken from freshly prepared test

solution (0 h) and at the end of the exposure (72 h)

Statistics: EC50 Logit analysis, NOEC: Dunnett's test ($\alpha = 0.05$)

Findings:

Analytical data: Mean measured concentrations test ranged from 60.5 - 98 % (start) and

from 78.5 to 89.5 % (end) of nominal concentrations.

Effects:

Morphological None

effects:

Biomass & growth See table 67

rate:

Table 67: Effects of Mandipropamid (NOA446510) on the green alga S. capricornutum

NOA446510 [mg/L] nominal	NOA446510 [mg/L] mean measured	Biomass (AUC) % inhibition in 72 h	Growth rate % inhibition in 72 h
1.25	1.15	2	1
2.5	2.1	4	0
5.0	4.2	14	1
10	8.45	39	7
20	17.9	25	4
40	27.8	18	2
EC ₅₀ ((0-72 h)	> 27.8 mg/L	> 27.8 mg/L
NOEC	(0-72 h)	≥ 27.8 mg/L	≥ 27.8 mg/L

Conclusion: E_bC_{50} (0-72 h): > 27.8 mg/L, NOEC (0-72 h): \geq 27.8 mg/L,

 E_rC_{50} (0-72 h): > 27.8 mg/L, NOEC (0-72 h): \geq 27.8 mg/L

based on mean measured concentrations

Reference: Growth inhibition test of NOA446510 tech. to Blue Algae (Anabaena

flos-aquae) under static conditions

Author(s), year: Knauer, K., 2002

Report/Doc. number: 2023553 Guideline(s): OECD 201

GLP: Yes

Deviations: None of relevance

Validity: Acceptable

<u>Test substance:</u> Mandipropamid (NOA446510), purity: 96.5 %, batch: SEZ2BP007

Material and Methods:

Test species: Blue alga (Anabaena flos-aquae)

Number of 2.04×10^4 cells/mL; 3 replicates per each concentration, 6 replicates for the

organisms: medium control Type of test, Static test, 96 h

duration:

Applied

concentrations:

Nominal: 0 (medium control), 0 (solvent control), 1.25, 2.5, 5.0, 10, 20 mg/L

Measured (mean): -- (medium control), -- (solvent control), 1.43, 2.8, 6.03, 13.7, 19.8 mg/L

Solvent 0.01 mL/L Dimethylformamide (DMF)

Test conditions:

Water quality: Alga culture medium (according to OECD guideline), hardness 20 mg/L as

CaCO₃

Temperature 23 ± 2 °C

pH 7.6 – 7.8 (0 h), 7.4 – 8.1 (72 h); control: 7.6 (0 h), 8.2 (96 h)

Incubation: Continuous illumination with cool white fluorescent light (approx. 2000)

lux), shaking rate at 100 rpm

Test parameters: Cell densities were measured using CytoFluor II (Fluorescence Multi-Well

Plate Reader), growth rate and biomass were determined after 24, 48, 72 and 96 hours; for chemical analysis (HPLC with UV-detection) of the test substance, samples of test solution were taken from freshly prepared test

solution (0 h) and at the end of exposure (96 h)

Statistics: EC₅₀ Logit analysis, NOEC: Dunnett's test ($\alpha = 0.05$)

Findings:

Analytical data: Mean measured concentrations ranged from 101 to 157 % (start) and from

97 to 122 % (end) of nominal concentrations.

Effects:

Morphological None

effects:

Biomass & growth See table below

rate:

Table 68: Effects of Mandipropamid (NOA446510) on the blue alga A. flos-aquae

NOA446510 [mg/L] nominal	NOA446510 [mg/L] mean measured	Biomass (AUC) % inhibition in 96 h	Growth rate % inhibition in 96 h
1.25	1.43	0.97	0
2.5	2.8	0	0
5.0	6.03	33	5.3
10	13.7	35	9.2
20	19.8	29	1.9
EC ₅₀ (0-96 h)	> 19.8 mg/L	> 19.8 mg/L
NOEC	(0-96 h)	≥ 19.8 mg/L	≥ 19.8 mg/L

Conclusion: E_bC_{50} (0-96 h): > 19.8 mg/L, NOEC (0-96 h): \geq 19.8 mg/L,

 E_rC_{50} (0-96 h): > 19.8 mg/L, NOEC (0-96 h): \geq 19.8 mg/L

based on mean measured concentrations

Reference: NOA-446510 - Toxicity to the Aquatic Higher Plant Lemna gibba in a

7-Day Static Growth Inhibition Test

Author(s), year: Bätscher, R., 2005

Report/Doc. number: 857050

Guideline(s): OECD 221 (Draft October 2000)

GLP: Yes

Deviations: None of relevance

Validity: Acceptable

Test substance: Mandipropamid (NOA446510), purity: 96.5 %, batch: SEZ2BP007

Material and

Methods:

Test species: Duckweed (Lemna gibba)

Number of 12 fronds (3 colonies with 4 fronds) per vessel; 3 replicates for each

organisms: concentration, control and solvent control

Type of test, Static test, 7 d

duration: Applied

concentrations:

Nominal: 0 (medium control), 0 (solvent control), 0.1, 0.26, 0.64, 1.6, 4.0, 10 mg/L -- (medium control), -- (solvent control), 0.1, 0.25, 0.62, 1.6, 3.0, 4.2 mg/L

Solvent 0.01 mL/L Dimethylformamide (DMF)

Test conditions:

Water quality: 20XAP growth medium (according to OECD guideline), hardness 300

mg/L as CaCO₃

Temperature 23 ± 2 °C

pH 7.6 – 7.8 (0 h), 7.4 – 8.1 (72 h); control: 7.6 (0 h), 8.2 (96 h)

Incubation: Continuous illumination with cool white fluorescent light (mean 7540 Lux)

in a temperature controlled water bath

Test parameters: Frond numbers were counted at 0, 3, 5 and 7 days to determine growth rate

and biomass (AUC), final biomass was determined on the basis of dry weight; for chemical analysis (HPLC analysis with UV/VIS) of test the substance, samples of test solution were taken from freshly prepared test

solutions (0 h) and at the end of exposure (7 d)

Statistics: NOEC: Dunnett's test, one tailed ($\alpha = 0.05$)

Findings:

Analytical data: Overall mean measured concentrations ranged from 42 % (10 mg/L) to

100 % (0.1 mg/L) of nominal.

Effects:

Morphological None

effects:

Biomass & growth Doubling time T_d of the control = 2.0 d (fulfilled the validity criterion),

rate: see also table below

Table 69: Effects of Mandipropamid (NOA446510) on duckweed Lemna gibba

NOA446510 [mg/L] nominal	NOA446510 [mg/L] mean measured	Biomass (AUC) % inhibition in 7 d	Growth rate % inhibition in 7 d
solvent control		0.0	0.0
control		-1.1	-0.6
0.1	0.1	1.0	0.7
0.26	0.25	-2.6	-0.9

NOA446510 [mg/L] nominal	NOA446510 [mg/L] mean measured	Biomass (AUC) % inhibition in 7 d	Growth rate % inhibition in 7 d
0.64	0.62	4.6	3.2
1.6	1.6	1.4	2.2
4.0	3.0	1.0	0.1
10	4.2	9.3*	6.8*
EC ₅₀ ((0-7 d)	> 4.2 mg/L	> 4.2 mg/L
NOEC	(0-7 d)	3.0 mg/L	3.0 mg/L

^{*} Significant difference (α =0.05) from the control

Conclusion: E_bC_{50} (7 d): > 4.2 mg/L, NOEC (7d): 3.0 mg/L,

 E_rC_{50} (7 d): > 4.2 mg/L, NOEC (7d): 3.0 mg/L,

based on mean measured concentrations

<u>Comment:</u> Mandipropamid is a fungicide and therefore a study on higher aquatic

plants is not required according to directive 91/414/EEC. However, in general an exposition of *Lemna gibba* is possible and therefore the study is

considered in the aquatic risk assessment.

5.4.4 Other aquatic organisms (including sediment)

Reference: NOA-446510 - A 96-Hour Flow-Through Acute Toxicity Test with the

Saltwater Mysid (Americamysis bahia)

Author(s), year: Palmer, S., Kendall, T. & Krueger, H., 2005b

Report/Doc. number: WIL 528A-137

Guideline(s): EPA OPPTS 850.1035

GLP: Yes

Deviations: None of relevance

Validity: Acceptable.

Test substance: Mandipropamid (NOA446510), purity: 96.1 %, batch: SEZ3BP007

Material and Methods:

Test species: Saltwater mysid (Americamysis bahia)

Number of 2 replicates each with 10 mysids per treatment and control

organisms:

Age: Juveniles < 24 hours old Type of test, Flow-through test, 96 h

duration: Applied

concentrations:

Nominal: 0 (control), 0 (solvent control) 0.65, 1.1, 1.8, 3.0, 5.0 mg/L Measured (mean): -- (control), -- (solvent control), 0.58, 0.94, 1.5, 2.3, 3.9 mg/L

solvent 0.1 mL/L Dimethylformamide (DMF)

Test conditions:

Water quality: Filtered natural seawater diluted with well water, salinity: 20 %

Temperature 23.9 - 24.8 °C

pH 8.1 – 8.3 (0 h), 7.98 – 8.01 (48 h)

 O_2 content: > 60 %, $6.3 - 7.4 \text{ mg } O_2/L$

Light regime: 16 hours light / 8 hours darkness

Test parameters: Mortality and sublethal effects were assessed after 7, 24, 48, 72 and

96 hours. For chemical analysis (HPLC method) of NOA446510 in the test

solution samples were taken at 0, 24 and 96 hours.

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Statistics: EC₅₀: Probit analysis, NOEC: directly from the raw data

Findings:

Analytical data: Overall mean measured concentrations were in the range of 77 - 89 % of

nominal concentrations.

Behavioural effects At 1.5 mg/L and higher concentrations sublethal effects like erratic

swimming, lethargy and loss of equilibrium were observed.

Mortality:

Analytical data: Overall mean measured concentration ranged from 77 – 89 % of nominal.

Effects: See table below

Table 70: Cumulative mortality of saltwater mysids exposed to Mandipropamid

Mean measured	Cumulative mortality (%)				
concentration (mg/L)	7 hours	24 hours	48 hours	72 hours	96 hours
Control	0	0	0	0	0
Solvent control	0	0	0	0	0
0.58	0	0	0	0	0
0.94	0	5	10	10	10
1.5	0	5	15	20	30
2.3	0	40	45	55	85
3.9	0	20	95	100	100
NOEC = 0.58 mg/L					
	L	C_{50} (96 h) = 1.7 mg/L	(95%CL 1.5 – 2.0 mg/	L)	

Conclusion: EC₅₀ (48 h): 1.7 mg/L, NOEC: 0.58 mg/L based on mean measured conc.

Reference: NOA-446510 - A 96-Hour Flow-Through Shell Deposition Test with

the Eastern Oyster (Crassostrea virginica)

Author(s), year: Palmer, S., Kendall, T. & Krueger, H., 2005c

Report/Doc. number: WIL 528A-139

Guideline(s): EPA OOPTS 850.1025

GLP: Yes

Deviations: None of relevance

Validity: Acceptable

Test substance: Mandipropamid (NOA446510), purity: 96.5 %, batch: SEZ2BP007

Material and Methods:

Test species: Eastern Oyster (*Crassostrea virginica*)
Number of 20 oysters per treatment and control

organisms:

Length: $41.7 \pm 2.6 \text{ mm}$

Type of test, Flow-through test, 96 h

duration:

Applied

concentrations:

Nominal: 0 (control), 0 (solvent control), 0.13, 0.25, 0.50, 1.0, 2.0 mg/L Heasured (mean): -- (control), -- (solvent control), 0.13, 0.25, 0.46, 0.8, 1.6 mg/L

solvent 0.1 mL/L Dimethylformamide (DMF)

Test conditions:

Water quality: Filtered natural seawater diluted with well water, salinity: 20 %

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Temperature 19.1 - 19.7 °C

pH 8.3 (0 h), 8.01 (96 h)

 O_2 content: > 60 %, 6.4 – 7.7 mg O_2/L

Light regime: 16 hours light / 8 hours darkness

Test parameters: Observations of mortalities were made at 1, 24, 48, 72 and 96 hours, the

shell growth was determined at test termination. For chemical analysis (HPLC method) of NOA446510 in the test solution samples were taken at

0, 48 and 96 hours.

Statistics: EC₅₀, NOEC: Kruskal-Wallis (ANOVA) and Dunn's multiple comparison

Findings:

Analytical data: Overall mean measured concentrations were in the range of 80 - 100 % of

nominal concentrations.

Mortality: None

Shell Deposition: See table below

Table 71: Mean shell deposition of eastern oysters (C. virginica) exposed to Mandipropamid

Mean measured concentration (mg/L)	Mean shell deposition (mm ± SD)	Inhibition of shell growth (%)			
Pooled control a)	4.5 ± 1.1				
0.13	4.0 ± 0.73	11			
0.25	4.1 ± 1.2	8.9			
0.46	4.3 ± 0.97	4.4			
0.80	$2.9 \pm 1.3*$	36			
1.6	0.08 ± 0.36	68			
NOEC = 0.46 mg/L					
EC	C_{50} (96 h) = 0.97 mg/L (95%CL 0.78 – 1.1 m	g/L)			

a) no statistically significant difference between dilution control and solvent control

Conclusion: EC₅₀ (48 h): 0.97 mg/L, NOEC: 0.46 mg/L based on mean measured conc.

^{*} statistically significant difference from the pooled control ($p \le 0.05$)

Summary and discussion: Acute (short-term) aquatic toxicity:

Data element: Acute (short-term) aq	uatic toxicity of the ac	ctive substance Mandipropamie	d	
Generally expressed in terms of LC5	50 or EC50 (mg/L)			
	L(E)C50 [mg/L]	Test guideline / design	GLP (y/n)	Reliability
Fish (96 hr LC50):				
Oncorhynchus mykiss Rainbow trout	> 2.9 ^{a)}	OECD 203	у	у
Pimephales promelas Fathead minnow	> 6.04 a)	OECD 203	у	У
Cyprinus carpio Common carp	> 2.0	OECD 203	у	У
Cyprinus carpio Common carp	8.63	OECD 203	у	у
Cyprinodon variegatus Sheephead minnow	4.5	US EPA OPPTS 850.1075	у	у
Crustacea (48 hr EC50):			•	•
Daphnia magna	7.1	OECD 202	у	у
Algae and water plants: (ErC50)			-	•
Selenastrum capricornutum Green alga	> 27.8 a) growth rate	OECD 201	y	у
Anabaena flos-aquae Blue alga	> 19.8 a) growth rate	OECD 201	у	у
Lemna gibba	> 4.4 (14d) growth rate	OECD 221 (Draft October 2000)	у	у
Other aquatic organisms (96 hr LC5	0):			
Americamysis bahia Saltwater mysid	1.7	EPA OPPTS 850.1035	у	у
Crassostrea virginica Eastern oyster	0.97	EPA OOPTS 850.1025	у	у

Conclusion: Mandipropamid is toxic to standard test species of fish, aquatic invertebrates, algae and higher plants. The most sensitive species is the saltwater oyster *Crassostrea virginica* with an EC50 of 0.97 mg/L.

^{a)} highest tested concentration (maximal solubility of test the substance under test conditions)

Summary and discussion: Chronic (long-term) aquatic toxicity

Data element: Chronic (long-term)	aquatic toxicity of the	e active substance Mandipropan	nid	
Generally expressed in terms of NO	DEC (mg/L)			
	NOEC [mg/L]	Test guideline / design	GLP (y/n)	Reliability
Fish (NOEC):				
Pimephales promelas	0.5 mortality growth	EPA OPPTS 850.1400	у	у
Crustacea (21 d NOEC,):				
Daphnia magna	0.28 length	OECD 202, Part II 4	У	у
Algae and water plants: (NOEC)				
Selenastrum capricornutum Green alga	≥ 27.8	OECD 201	У	у
Anabaena flos-aquae Blue alga	≥ 19.8	OECD 201	У	у
<i>Lemna gibba</i> Duckweed	3.0	OECD 221 (Draft October 2000)	у	у

Conclusion: Mandipropamid is chronic toxic to fish and daphnids (*Daphnia magna*). The most sensitive species is *Daphnia magna* with a NOEC= 0.28 mg/L.

Note: Aquatic toxicity studies for metabolites SYN 504851 and SYN 536638 are available but are missing for SYN 521195 and for the major metabolites in the sediment SYN521195 and SYN 539678. Thus a reliable classification regarding the hazardous to aquatic environment for all degradation products is not possible.

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5.5 Comparison with criteria for environmental hazards (sections 5.1 – 5.4)

conditions within 28 days. biotic aquatic degradation studies. hydrolysis studies indicate primary of some degradants it is not possible tous to the aquatic environment. Log Kow is <3 Aandipropamid Log Kow = 3.2 BCF < 100 propamid BCF is in the range of 35 and 48 = 0.97 mg/L	Endpoint	Classification	_	Evidence for
mid Mandipropamid is not readily biodegradable under test conditions within 28 days. Ultimate degradation studies with the exception of the hydrolysis studies indicate primary degradation, but due to missing data on aquatic toxicity of some degradants it is not possible to show that the metabolites are not classified as hazardous to the aquatic environment. Therefore a non rapid degradation is proposed. Log Kow is < 4 Mandipropamid Log Kow = 3.2 Mandipropamid Log Kow = 3.2 Mandipropamid BCF is in the range of 35 and 48 LC/EC ₅₀ < 1 mg/L Crassostrea virginica EC50 = 0.97 mg/L For non rapidly degradable substances: 0.1 < NOEC(21d) = 0.028 mg/L Daphmia magna NOEC(21d) = 0.028 mg/L		(criteria	in bold)	Mandipropamid
nd Mandipropamid is not readily biodegradable under test conditions within 28 days. Ultimate degradation studies with the exception of the hydrolysis studies indicate primary degradation, but due to missing data on aquatic toxicity of some degradants it is not possible to show that the metabolites are not classified as hazardous to the aquatic environment. Therefore a non rapid degradation is proposed. Log K _{ow} is < 4 Log K _{ow} is < 4 Mandipropamid Log K _{ow} = 3.2 BCF < 500 Mandipropamid BCF is in the range of 35 and 48 LC/EC ₅₀ ≤ 1 mg/L Crassostrea virginica EC50 = 0.97 mg/L For non rapidly degradable substances: 0.1 < NOEC(21d) = Daphnia magna NOEC(21d) = Daphnia magna		CLP (2 nd ATP)	DSD	
Mandipropamid is not readily biodegradable under test conditions within 28 days. Ultimate degradation could not be shown in abiotic and biotic aquatic degradation studies. Available degradation studies with the exception of the hydrolysis studies indicate primary degradation, but due to missing data on aquatic toxicity of some degradants it is not possible to show that the metabolites are not classified as hazardous to the aquatic environment. Therefore a non rapid degradation is proposed. Log K _{ow} is < 4 Mandipropamid Log K _{ow} = 3.2 BCF < 500 Mandipropamid BCF is in the range of 35 and 48 LC/EC ₅₀ ≤ 1 mg/L LC/EC ₅₀ ≤ 1 mg/L For non rapidly degradable substances: O1 < NOEC(21d) = Daphnia magna NOEC(21d) = Daphnia magna NOEC(21d) = Daphnia magna NOEC(21d) = NOEC(21d) = Daphnia magna NOEC(21d) = NOEC(21d	Degradation			
	Mandipropamid	Mandipropamid is not readily biodegradable u Ultimate degradation could not be shown in al Available degradation studies with the excepti degradation, but due to missing data on aquati to show that the metabolites are not classified Therefore a non rapid degradation is proposed	nder test conditions within 28 days. viotic and biotic aquatic degradation studies. on of the hydrolysis studies indicate primary c toxicity of some degradants it is not possible as hazardous to the aquatic environment.	The classification as R53 according to Directive 67/548/EEC is based on the fact that the active substance is not considered as ready biodegradable/rapid degradable.
cowLog K_{ow} is < 4 Mandipropamid Log K_{ow} = 3.2 Mandipropamid Log K_{ow} = 3.2 	Bioaccumulation			
Mandipropamid BCF is in the range of 35 and 48Mandipropamid BCF is in the range of 35 and 48Mandipropamid BCF is in the range of 35 and 48LC/EC50 $\le 1 \text{ mg/L}$ LC/EC50 $\le 1 \text{ mg/L}$ Crassostrea virginica EC50 = 0.97 mg/LGrassostrea virginica EC50 = 0.97 mg/LFor non rapidly degradable substances: 0.1 <noec <math="">\le 1 \text{ mg/L}NOEC(21d) = 0.28 mg/L</noec>	Criteria LogKow	$\begin{array}{c} \textbf{Log K}_{ow} \ \textbf{is} < \textbf{4} \\ \text{Mandipropamid Log K}_{ow} = 3.2 \end{array}$	$\begin{array}{c} \text{Log } \mathbf{K_{ow} \ is} < 3 \\ \text{Mandipropamid Log } \mathbf{K_{ow}} = 3.2 \end{array}$	The measured BCF after normalization to 11 % lipid content is in the range of 35 and 48 and is below the two classification
$LC/EC_{50} \le 1 \text{ mg/L}$ $Crassostrea virginica EC50 = 0.97 \text{ mg/L}$ $0.1 < NOEC \le 1 \text{ mg/l}$ 0.28 mg/L	Criteria BCF	BCF < 500 Mandipropamid BCF is in the range of 35 and 48	BCF < 100 Mandipropamid BCF is in the range of 35 and 48	Mandipropamid is considered to have a low bioaccumulation potential.
LC/EC50 \leq 1 mg/Ln rapidly degradable substances: $0.1 < NOEC \leq 1 mg/L$ nia magna $NOEC(21d) = 0.28mg/L$	Acute aquatic to	xicity		
n rapidly degradable substances: 0.1 <noec 0.97="" l="" mg="" noec(21d)="</td" ≤1=""><td>Criteria</td><td>LC/EC₅₀</td><td>≤ 1 mg/L</td><td>Mandipropamid is of high acute toxicity to saltwater oyster Crassostrea virginica with an EC50 of 0.97 mg/L and fulfills</td></noec>	Criteria	LC/EC ₅₀	≤ 1 mg/L	Mandipropamid is of high acute toxicity to saltwater oyster Crassostrea virginica with an EC50 of 0.97 mg/L and fulfills
n rapidly degradable substances: 0.1 <noec l="" mg="" noec(21d)="0.28mg/L</td" ≤1=""><td></td><td>Crassostrea virginic</td><td>a EC50 = 0.97 mg/L</td><td>the criteria for the proposed classification as R50 according to Directive 67/548/EEC and the criteria for the proposed classification as H400 according to Regulation EC 1272/2008. A M-factor of 1 is applicable based on 0.1 <l(e)c<sub>50 ≤1 mg/l.</l(e)c<sub></td></noec>		Crassostrea virginic	a EC50 = 0.97 mg/L	the criteria for the proposed classification as R50 according to Directive 67/548/EEC and the criteria for the proposed classification as H400 according to Regulation EC 1272/2008. A M-factor of 1 is applicable based on 0.1 <l(e)c<sub>50 ≤1 mg/l.</l(e)c<sub>
For non rapidly degradable substances: 0.1 <noec <math="">\le1 mg/l NOEC(21d) = 0.28mg/L</noec>	Chronic aquatic	toxicity		
NOEC(21d) = 0.28mg/L	Criteria	For non rapidly degradable substances: 0.1 <noec l<="" mg="" td="" ≤1=""><td></td><td>Mandipropamid is chronic toxic to daphnids (<i>Daphnia magna</i>) with a NOEC= 0.28 mg/L. Therefore Mandipropamid fulfills the criterio for the proposed closeification of H411 geometric</td></noec>		Mandipropamid is chronic toxic to daphnids (<i>Daphnia magna</i>) with a NOEC= 0.28 mg/L. Therefore Mandipropamid fulfills the criterio for the proposed closeification of H411 geometric
				to Regulation EC 1272/2008.

5.6 Conclusions on classification and labelling for environmental hazards (sections 5.1 – 5.4)

Conclusion of environmental classification according to Directive 67/548/EEC

Mandipropamid should be classified Dangerous for the Environment with the following risk and safety phrases:

N Dangerous for the Environment

R50 Very toxic to aquatic organisms

R53 May cause long term effects in the environment

Conclusion of environmental classification according to Regulation EC 286/2011 (2nd ATP to EC 1272/2008)

Based on the CLP Regulation, mandipropamid should be classified as:

Classification categories aquatic environmental hazard **acute category 1**aquatic environmental hazard **chronic category 2**

GHS Pictogram



Signal Word

Warning

H400 'Very toxic to aquatic life',

H411 'Toxic to aquatic life with long lasting effects'

M-factor (acute)

1

6 OTHER INFORMATION

Environmental fate properties and environmental hazard assessments of this CLH report are based on studies and summaries of the Draft Assessment Report and its addenda

7 REFERENCES

7.1 Physico-chemical properties

Author(s)	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N-R/NR	Owner
Das, R.	2002a	Melting point / melting range of NOA 446510 Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 109673 GLP Not Published Syngenta File N° NOA446510/0024	Y	SYN
Das, R.	2003a	Boiling point / boiling range of NOA 446510 Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 109674 GLP Not Published Syngenta File N° NOA446510/0038	Y	SYN
Vehling, H.	2005	Thermal stability / stability in air Syngenta Crop Protection AG, Basel, Switzerland Syngenta - Process Hazards Section, Huddersfield, United Kingdom, Report No HT05/284 GLP Not Published Syngenta File N° NOA446510/0401	Y	SYN
Füldner, H.	2003	Density of solids of NOA 446510 Syngenta Crop Protection AG, Basel, Switzerland Solvias AG, Basel, Switzerland, Report No L02- 009577 GLP Not Published Syngenta File N° NOA446510/0031	Y	SYN
Geoffroy, A.	2003	Vapour pressure curve of NOA 446510 Syngenta Crop Protection AG, Basel, Switzerland Solvias AG, Basel, Switzerland, Report No L03- 002392 GLP Not Published Syngenta File N° NOA446510/0064	Y	SYN
Das, R.	2006d	NOA 446510 - Statement on Method used for vapour pressure Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 10115200 not GLP	Y	SYN

Author(s)	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N-R/NR	Owner
		Not Published		
Baker, S.D.	2005	Henry's law constant Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection, Guildford, United Kingdom Syngenta File N° NOA446510/0445	Y	SYN
Das, R.	2002b	General physico-chemical properties of NOA 446510 Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 109670 GLP Not Published Syngenta File N° NOA446510/0025	Y	SYN
Das, R.	2005a	NOA 446510 tech Color, physical state and odor Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 114667 GLP Not Published Syngenta File N° NOA446510/0376	Y	SYN
Oggenfuss, P.	2004	NOA 446510 - Spectra Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 111640 GLP Not Published Syngenta File N° NOA446510/0082	Y	SYN
Das, R.	2003b	Water solubility of NOA 446510 Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 109671 GLP Not Published Syngenta File N° NOA446510/0026	Y	SYN
Das, R.	2006a	NOA 446510 - Statement on Method used for water solubility Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 10115199 not GLP Not Published	Y	SYN
Das, R.	2004a	CGA 380775 - Water solubility Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 113111	Y	SYN

Author(s) Year		Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N-R/NR	Owner
		GLP Not Published Syngenta File N° CGA380775/0001		
Das, R.	2004b	CGA 380778 - Water solubility Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 112389 GLP Not Published Syngenta File N° CGA380778/0001	Y	SYN
Das, R.	2004c	SYN 500003 - Water solubility Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 113440 GLP Not Published Syngenta File N° SYN500003/0001	Y	SYN
Das, R.	2005b	SYN 504851 - Water solubility Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 113684 GLP Not Published Syngenta File N° SYN504851/0005	Y	SYN
Das, R.	2004d	SYN 535839 - Water solubility Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 113301 GLP Not Published Syngenta File N° SYN535839/0001	Y	SYN
Das, R.	2005c	SYN 536638 - Water solubillity Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 113638 GLP Not Published Syngenta File N° SYN536638/0001	Y	SYN
Das, R.	2005d	NOA 458422 - Water solubility Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 113718 GLP Not Published Syngenta File N° CA4011/0005	Y	SYN
Das, R.	2005e	NOA 446510 tech Solubility in organic solvents	Y	SYN

Author(s) Year		Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N-R/NR	Owner
		Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 114668 GLP Not Published Syngenta File N° NOA446510/0375		
Das, R.	2006ь	NOA 446510 - Statement on Method used for solubility in organic solvents Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 10115182 not GLP Not Published	Y	SYN
Das, R.	2003c	Octanol / water partition coefficient of NOA 446510 Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 109672 GLP Not Published Syngenta File N° NOA446510/0027	Y	SYN
Das, R.	2006c	NOA 446510 - Statement on Method used for Octanol/Water Partition Coefficient Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 10115098 not GLP Not Published	Y	SYN
Das, R.	2005f	CGA 380775 - Octanol / water partition coefficient Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 113112 GLP Not Published Syngenta File N° CGA380775/0003	Y	SYN
Das, R.	2004e	Octanol / water partition coefficient Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 112390 GLP Not Published Syngenta File N° CGA380778/0002	Y	SYN
Das, R.	2005g	SYN 500003 - Octanol / water partition coefficient Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 113441 GLP	Y	SYN

Author(s) Year		Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N-R/NR	Owner
		Not Published Syngenta File N° SYN500003/0006		
Das, R.	2005h	Octanol/water parition coefficient Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 113685 GLP Not Published Syngenta File N° SYN504851/0008	Y	SYN
Das, R.	2004f	Octanol/water partition coefficient Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 113302 GLP Not Published Syngenta File N° SYN535839/0002	Y	SYN
Das, R.	2005i	SYN 536638 - Octanol / water partition coefficient Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 113639 GLP Not Published Syngenta File N° SYN536638/0002	Y	SYN
Das, R.	2005j	NOA 458422 - Octanol/water partition coefficient Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection Münchwilen AG, Münchwilen, Switzerland, Report No 113720 GLP Not Published Syngenta File N° CA4011/0007	Y	SYN
Buckel, T	2002	Hydrolysis of [Ethyl-1- 14C]-labelled NOA446510 under Laboratory Conditions Syngenta Crop Protection AG, Basel, Switzerland, Report No 02TB01 GLP Not Published Syngenta File N° NOA446510/0018	Y	SYN
Hand, L H	2003	Aqueous Photolysis of 14C-Methoxyphenyl-NOA446510 under Laboratory Conditions Syngenta - Jealott's Hill International, Bracknell, Berkshire, United Kingdom, Report No RJ3395B GLP Not Published Syngenta File N° NOA446510/0041	Y	SYN

Author(s) Year Schmidt, E. 2004		Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N-R/NR	Owner	
		Quantum yield of the direct photochemical degradation of NOA 446510 in aqueous solution Syngenta Crop Protection AG, Basel, Switzerland Solvias AG, Basel, Switzerland, Report No L02-009816 GLP Not Published Syngenta File N° NOA446510/0118	Y	SYN	
Martin, N.	2003	Dissociation constant of NOA 446510 in water Syngenta Crop Protection AG, Basel, Switzerland Solvias AG, Basel, Switzerland, Report No L02- 009578 GLP Not Published Syngenta File N° NOA446510/0029	Y	SYN	
Martin, N.	2004a	Dissociation constant of CGA 380775 in water Syngenta Crop Protection AG, Basel, Switzerland Solvias AG, Basel, Switzerland, Report No L04- 006774 GLP Not Published Syngenta File N° CGA380775/0002	Y	SYN	
Martin, N.	2004b	Dissociation constant of CGA 380778 in water Syngenta Crop Protection AG, Basel, Switzerland Solvias AG, Basel, Switzerland, Report No L04- 006773 GLP Not Published Syngenta File N° CGA380778/0004	Y	SYN	
Martin, N.	Martin, N. 2004c Dissociation constant of SYN 500003 in water Syngenta Crop Protection AG, Basel, Switzerland Solvias AG, Basel, Switzerland, Report No L04- 007084 GLP Not Published Syngenta File N° SYN500003/0002		Y	SYN	
Richner, D.			Y	SYN	

Author(s) Year		Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N-R/NR	Owner	
Martin, N.	2004d	Dissociation constant of SYN 535839 in water Syngenta Crop Protection AG, Basel, Switzerland Solvias AG, Basel, Switzerland, Report No L04- 006772 GLP Not Published Syngenta File N° SYN535839/0003	Y	SYN	
Martin, N.	2005	Dissociation constant of SYN 536638 in water Syngenta Crop Protection AG, Basel, Switzerland Solvias AG, Basel, Switzerland, Report No L04- 007792 GLP Not Published Syngenta File N° SYN536638/0003	Y	SYN	
Richner, D.	2005ь	Dissociation constant of NOA 458422 in water Syngenta Crop Protection AG, Basel, Switzerland Solvias AG, Basel, Switzerland, Report No L05- 000123 GLP Not Published Syngenta File N° CA4011/0006	Y	SYN	
Widmer, H	2003	Atmospheric Oxidation of NOA446510 by Hydroxyl Radicals; Rate Estimation Syngenta Crop Protection AG, Basel, Switzerland, Report No 95A2003006WI Not GLP Not Published Syngenta File N° NOA446510/0035	Y	SYN	
Jackson, W.	2005a	Plammability (solids) Syngenta Crop Protection AG, Basel, Switzerland Syngenta - Process Hazards Section, Huddersfield, United Kingdom, Report No HT05/280 GLP Not Published Syngenta File N° NOA446510/0405		SYN	
Jackson, W.	2005b	Relative self-ignition temperature for solids Syngenta Crop Protection AG, Basel, Switzerland Syngenta - Process Hazards Section, Huddersfield, United Kingdom, Report No HT05/282 GLP Not Published Syngenta File N° NOA446510/0403	Y	SYN	
Jackson, W.	2005c	Explosive properties Syngenta Crop Protection AG, Basel, Switzerland	Y	SYN	

Author(s)	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N-R/NR	Owner
		Syngenta - Process Hazards Section, Huddersfield, United Kingdom, Report No HT05/281 GLP Not Published Syngenta File N° NOA446510/0404		
Richner, D.	2005c	Surface tension of NOA 446510 tech. Syngenta Crop Protection AG, Basel, Switzerland Solvias AG, Basel, Switzerland, Report No L05- 002772 GLP Not Published Syngenta File N° NOA446510/0426	Y	SYN
Jackson, W.	2005d	Oxidising properties Syngenta Crop Protection AG, Basel, Switzerland Syngenta - Process Hazards Section, Huddersfield, United Kingdom, Report No HT05/283 GLP Not Published Syngenta File N° NOA446510/0402	Y	SYN

7.2 Human health hazard assessment

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant), Published or not	Data Protection Claimed Y/N-R/NR	Owner
Annex II Data	and Inform	ation			
Barlow, S.	KIIA 5.8.1	2005	Threshold of Toxicological Concern (TTC) A tool for assessing substances of unknown toxicity present at low levels in the diet	N	
Brammer, A.	KIIA 5.3.4/01	2005 b	NOA 446510: 1 Year Oral Toxicity Study in Dogs Syngenta Crop Protection AG, Basel, Switzerland Central Toxicology Laboratory (CTL), Cheshire, United Kingdom, Report No PD1273-REG GLP Not Published Syngenta File N° NOA446510/0521	Y	SYN
Callander, R.	KIIA 5.4.1/01	2005	NOA446510: Bacterial Mutation Assay In S Typhimurium And E.Coli Syngenta Crop Protection AG, Basel, Switzerland Central Toxicology Laboratory (CTL), Cheshire, United Kingdom, Report No YV6190/REV-001 GLP Not Published Syngenta File N° NOA446510/0487	Y	SYN
Callander, R. D.	KIIA 5.8.1/02	2006	SYN 500003: Bacterial Mutation Assay in S. Typhimurium and E. Coli Syngenta Central Toxicology Laboratory, Alderley Park, Cheshire Report No YV7195-REG GLP Not Published Syngenta File N° T003954-05	Y	SYN
Callander, R. D.	KIIA 5.8.2/01	2006	SYN 545038: Bacterial Mutation Assay in S. Typhimurium and E. Coli Syngenta Central Toxicology Laboratory, Alderley Park, Cheshire Report No YV7241-REG GLP Not Published Syngenta File N° T013670-05	Y	SYN

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant), Published or not	Data Protection Claimed Y/N-R/NR	Owner
Clay, P.	KIIA 5.4.3/01	2002	NOA446510: L5178Y TK+/- mouse lymphoma mutation assay Syngenta Limited, Cheshire, United Kingdom Central Toxicology Laboratory (CTL), Cheshire, United Kingdom, Report No CTL/VV0286/REG/REPT/REV-001 GLP Not Published Syngenta File N° NOA446510/0014	Y	SYN
Clay, P.	KIIA 5.4.5/01	2005	NOA446510: In Vivo Rat Liver Unscheduled DNA Synthesis Assay Syngenta Crop Protection AG, Basel, Switzerland Central Toxicology Laboratory (CTL), Cheshire, United Kingdom, Report No SR1193 GLP Not Published Syngenta File N° NOA446510/0438	Y	SYN
Fox, V.	KIIA 5.4.2/01	2002	NOA446510: In vitro cytogenetic assay in human lymphocytes Syngenta Limited, Cheshire, United Kingdom Central Toxicology Laboratory (CTL), Cheshire, United Kingdom, Report No CTL/SV1144/REG/REPT GLP Not Published Syngenta File N° NOA446510/0015	Y	SYN
Fox, V.	KIIA 5.4.4/01	2005	NOA446510: Mouse Bone Marrow Micronucleus Test Syngenta Crop Protection AG, Basel, Switzerland Central Toxicology Laboratory (CTL), Cheshire, United Kingdom, Report No SR1176 GLP Not Published Syngenta File N° NOA446510/0486	Y	SYN

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant), Published or not	Data Protection Claimed Y/N-R/NR	Owner
Johnson, I.	KIIA 5.2.4/01	2004a	MANDIPROPAMID: Skin Irritation Study in the Rabbit Syngenta Crop Protection AG, Basel, Switzerland Central Toxicology Laboratory (CTL), Alderley Park, Macclesfield, Cheshire, United Kingdom Report No CTL/EB4953 GLP Not Published Syngenta File N° NOA446510/0106	Y	SYN
Johnson, I.	KIIA 5.2.5/01	2004 b	MANDIPROPAMID: Eye Irritation Study in the Rabbit Syngenta Crop Protection AG, Basel, Switzerland Central Toxicology Laboratory (CTL), Alderley Park, Macclesfield, Cheshire, United Kingdom Report No CTL/FB5931 GLP Not Published Syngenta File N° NOA446510/0113	Y	SYN
Johnson, I.	KIIA 5.2.6/01	2004c		Y	SYN
Kilgour J., Lister N.	KIIA 5.8.1	2006	Case for non-relevance using a Threshold of Toxicological Concern (TTC) approach	N	
Kilgour, J.	KIIA 5.2.3/01	2003	MANDIPROPAMID: 4-Hour acute inhalation toxicity study in rats (EPA and OECD) Syngenta Crop Protection AG, Basel, Switzerland Central Toxicology Laboratory (CTL), Alderley Park, Macclesfield, Cheshire, United Kingdom Report No CTL/HR2410/REG/REPT GLP Not Published Syngenta File N° NOA446510/0030	Y	SYN
Kilgour, J. D. et al	KIIA 5.8.2/02	2006	Statement: Mandipropamid (NOA446510) Impurities: Relevance of Potentially Significant Impurities in Relation to the Technical Specification	Y	SYN

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant), Published or not	Data Protection Claimed Y/N-R/NR	Owner
Kroes, R.	KIIA 5.8.1	2003	Structure-based thresholds of toxicological concern (TTC): guidance for application to substances present at low levels in the diet	N	
Kuhn, J.	KIIA 5.2.2/01	2005	NOA-446510 Technical (Batch SEZ2BP007) - Acute Dermal Toxicity Study in Rats Syngenta Crop Protection AG, Basel, Switzerland Stillmeadow Inc., Houston, United States Report No 9169-05 T003767-05 GLP Not Published Syngenta File N° NOA446510/0504	Y	SYN
Lees, D.	KIIA 5.3.7/01	2005	NOA 446510: 21/28 Day Dermal Toxicity Study In Rats Syngenta Crop Protection AG, Basel, Switzerland Central Toxicology Laboratory (CTL), Cheshire, United Kingdom, Report No LR0596-REG GLP Not Published Syngenta File N° NOA446510/0568	Y	SYN
Milburn, G.	KIIA 5.5.3/01	2005a		Y	SYN
Milburn, G.	KIIA 5.6.1/01	2005 b	NOA 446510:Two Generation Reproduction Toxicity Study In Rats Syngenta Crop Protection AG, Basel, Switzerland Central Toxicology Laboratory (CTL), Cheshire, United Kingdom, Report No RR0990/REG GLP Not Published Syngenta File N° NOA446510/0565	Y	SYN

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant), Published or not	Data Protection Claimed Y/N-R/NR	Owner
Milburn, G.	KIIA 5.7.1/01	2005c	NOA 446510: Acute Neurotoxicity Study in Rats Syngenta Crop Protection AG, Basel, Switzerland Central Toxicology Laboratory (CTL), Cheshire, United Kingdom, Report No AR7352-REG GLP Not Published Syngenta File N° NOA446510/0513	Y	SYN
Moore, G.	KIIA 5.2.1/01	2004	Acute Oral Toxicity Up and Down Procedure in Rats Syngenta Crop Protection AG, Basel, Switzerland Product Safety Labs, East Brunswick, United States Report No 14702 GLP Not Published Syngenta File N° NOA446510/0092	Y	SYN
Moxon, M.	KIIA 5.6.10/01	2005a	NOA 446510:Prenatal Developmental Toxicity Study in Rats Syngenta Crop Protection AG, Basel, Switzerland Central Toxicology Laboratory (CTL), Cheshire, United Kingdom, Report No RR0963-REG GLP Not Published Syngenta File N° NOA446510/0531	Y	SYN
Moxon, M.	KIIA 5.6.11/01	2005 b	NOA 446510:Prenatal Developmental Toxicity Study In The Rabbit Syngenta Crop Protection AG, Basel, Switzerland Central Toxicology Laboratory (CTL), Cheshire, United Kingdom, Report No RB0962/REG GLP Not Published Syngenta File N° NOA446510/0543	Y	SYN

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant), Published or not	Data Protection Claimed Y/N-R/NR	Owner
Pinto, P.	KIIA 5.3.2/01	2005a	MANDIPROPAMID:90 Day Dietary Toxicity Study In Rats Syngenta Crop Protection AG, Basel, Switzerland Central Toxicology Laboratory (CTL), Alderley Park, Macclesfield, Cheshire, United Kingdom Report No CTL/PR1263/REG/RE GLP Not Published Syngenta File N° NOA446510/0535	Y	SYN
Pinto, P.	KIIA 5.5.2/01	2005 b	NOA 446510:Two Year Chronic Toxicity And Carcinogenicity Study In Rats Syngenta Crop Protection AG, Basel, Switzerland Syngenta Limited, Cheshire, United Kingdom, Report No PR1274-Reg GLP Not Published Syngenta File N° NOA446510/0560	Y	SYN
Pinto, P.	KIIA 5.7.4/01	2005c		Y	SYN
Pooles, A.	KIIA 5.8.1/01	2006	SYN 500003: Acute Oral Toxicity in the Rat – Up and Down Procedure Syngenta Central Toxicology Laboratory, Alderley Park, Cheshire Safepharm Laboratories Limited, Shardlow Business Park, Derbyshire, SPL Project No 0006/0737 GLP Not Published	Y	SYN
Renwick A.G.	KIIA 5.8.1	2005	Structure-based thresholds of toxicological concern: guidance for application to substances present at low levels in the diet	N	

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant), Published or not	Data Protection Claimed Y/N-R/NR	Owner
Roberts, K.	KIIA 5.1/02	2005a	NOA 446510: Tissue depletion following a single oral dose (3 mg/kg and 300 mg/kg) in the rat Central Toxicology Laboratory (CTL), Alderley Park, Macclesfield, Cheshire, United Kingdom Report No CTL/UR0761/REG/REPT GLP Not published Syngenta File N° NOA446510/0533	Y	SYN
Roberts, K.	KIIA 5.1/03	2005 b	NOA 446510: Tissue accumulation and depletion following multiple oral dosing (3 mg/kg) in the rat Central Toxicology Laboratory (CTL), Alderley Park, Macclesfield, Cheshire, United Kingdom Report No CTL/UR0786/REG/REPT GLP Not published Syngenta File N° NOA446510/0527	Y	SYN
Silcock, R. and Duerden, A.	KIIA 5.1/01	2005	NOA 446510: Absorption, distribution and excretion in the rat Central Toxicology Laboratory (CTL), Alderley Park, Macclesfield, Cheshire, United Kingdom Report No CTL/UR0719/REG/REPT GLP Not published Syngenta File N° NOA446510/0563	Y	SYN
Wake, A.	KIIA 5.1/04	2005	NOA 446510: Biotransformation in the rat Central Toxicology Laboratory (CTL), Alderley Park, Macclesfield, Cheshire, United Kingdom Report No CTL/UR0758/REG/REPT GLP Not published Syngenta File N° NOA446510/0561	Y	SYN

7.3 Environmental hazard assessment

7.3.1 Fate and Behaviour in the environment

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N- R/NR	Owner
Adam, D.	KIIA 7.4.2/01	2004	Adsorption/desorption of 14C-CGA 380778 on Various Soils Syngenta Crop Protection AG, Basel, Switzerland RCC Ltd., Itingen, Switzerland, Report No 855381 GLP Not Published Syngenta File N° CGA380778/0005	Y	SYN
Adam, D.	KIIA 7.4.2/08	2005	Adsorption / Desorption of SYN539679 on Soils Syngenta Crop Protection AG, Basel, Switzerland RCC Ltd., Itingen, Switzerland, Report No A04037 GLP Not Published Syngenta File N° SYN539679/0001	Y	SYN
Berdat, T., Nicollier, G.	KIIA 7.2.3/01	2005 a	Rate of Degradation of [14C]CGA 380778 (Metabolite of NOA 446510) in Various Soils under Aerobic Laboratory Conditions at 20oC Syngenta Crop Protection AG, Basel, Switzerland, Report No T004943-04 GLP Not Published Syngenta File N° CGA380778/0010	Y	SYN
Berdat, T., Nicollier, G.	KIIA 7.2.3/05	2005 b	Rate of Degradation of [Chlorophenyl-U-14C]-labelled SYN 521195 (Metabolite of NOA446510) in Various Soils under Aerobic Laboratory Conditions at 20°C Syngenta Crop Protection AG, Basel, Switzerland, Report No T013941-04 GLP Not Published Syngenta File N° SYN521195/0003	Y	SYN
Berdat, T., Nicollier, G.	KIIA 7.4.2/06	2004	Adsorption / Desorption of NOA 458422 in Various Soils Syngenta Crop Protection AG, Basel, Switzerland, Report No T013298-04	Y	SYN

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N- R/NR	Owner
			GLP Not Published Syngenta File N° CA4011/0004		
Bramley, Y., Oliver, S.	KIIA 7.1.3/02	2005	Soil Photolysis of 14C-Chlorophenyl Ring Labelled NOA446510 under Laboratory Conditions Syngenta Crop Protection AG, Basel, Switzerland Syngenta, Jealott's Hill, United Kingdom, Report No RJ3535B GLP Not Published Syngenta File N° NOA446510/0236	Y	SYN
Buckel, T	KIIA 7.5/01	2002	Hydrolysis of [Ethyl-1- 14C]-labelled NOA446510 under Laboratory Conditions Syngenta Crop Protection AG, Basel, Switzerland, Report No 02TB01 GLP Not Published Syngenta File N° NOA446510/0018	Y	SYN
Clark, A.	KIIA 7.1.1/03 KIIA 7.2.1/03	2004	Metabolism of [14C-Chlorophenyl]-NOA-446510 in Viable Soil under Aerobic Conditions Syngenta Crop Protection AG, Basel, Switzerland Syngenta Crop Protection, Inc., Greensboro, United States, Report No 680-02 GLP Not Published Syngenta File N° NOA446510/0152	Y	SYN
Dorn, R.	KIIA 7.2.3/06	2005 a	Re-evaluation of times degradation times of NOA446510 and some selected metabolites in various soil under laboratory conditions Syngenta Crop Protection AG, Basel, Switzerland, Report No Ass05RD03 Syngenta File N° NOA446510/0416	Y	SYN
Dorn, R.	KIIA 7.3/01	2005 b	Summary of half-lives of NOA446510 in soil in various field trials across Europe Syngenta Crop Protection AG, Basel, Switzerland, Report No Ass05RD05 Syngenta File N° NOA446510/0418	Y	SYN

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N- R/NR	Owner
Dorn, R.	KIIA 7.8.3/03	2005 c	Evaluation of half-lives of selected metabolites of NOA446510 in aquatic systems Syngenta Crop Protection AG, Basel, Switzerland, Report No Ass05RD04 Syngenta File N° NOA446510/0417	Y	SYN
Evans, P.	KIIA 7.3.1/01	2003 a	Dissipation Study with NOA446510 in or on Soil in Switzerland Syngenta Crop Protection AG, Basel, Switzerland Syngenta - Jealott's Hill International, Bracknell, Berkshire, United Kingdom, Report No RJ3451B GLP Not Published Syngenta File N° NOA446510/0055	Y	SYN
Evans, P.	KIIA 7.3.1/02	2003 b	Dissipation Study with NOA446510 in or on Soil in Switzerland Syngenta Crop Protection AG, Basel, Switzerland Syngenta - Jealott's Hill International, Bracknell, Berkshire, United Kingdom, Report No RJ3450B GLP Not Published Syngenta File N° NOA446510/0054	Y	SYN
Evans, P.	KIIA 7.3.1/03	2005 a	Mandipropamid 250 g/L SC formulation (A12946C): Dissipation in or on Soil in Spain (2003) Syngenta Crop Protection AG, Basel, Switzerland Syngenta, Jealott's Hill, United Kingdom, Report No RJ3662B GLP Not Published Syngenta File N° NOA446510/0372	Y	SYN
Evans, P.	KIIA 7.3.1/04	2005 b	Mandipropamid 250 g/L SC formulation (A12946C): Dissipation in or on Soil in France (North) 2003 Syngenta Crop Protection AG, Basel, Switzerland Syngenta, Jealott's Hill, United Kingdom, Report No RJ3663B GLP Not Published	Y	SYN

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N- R/NR	Owner
Evans, P.	KIIA 7.3.1/05	2005 c	Syngenta File N° NOA446510/0392 Mandipropamid 250 g/L SC formulation (A12946C): Dissipation in or on Soil in France (South) 2003 Syngenta Crop Protection AG, Basel, Switzerland Syngenta, Jealott's Hill, United Kingdom, Report No RJ3664B GLP Not Published Syngenta File N° NOA446510/0391	Y	SYN
Evans, P.	KIIA 7.3.1/07	2005 d	Mandipropamid 250 g/L SC formulation (A12946B): Dissipation in or on Soil in France (South) 2004 Syngenta Crop Protection AG, Basel, Switzerland Syngenta, Jealott's Hill, United Kingdom, Report No RJ3667B GLP Not Published Syngenta File N° NOA446510/0394	Y	SYN
Evans, P.	KIIA 7.3.1/08	2005 e	Mandipropamid 250 g/L SC formulation (A12946B): Dissipation in or on Soil in France (North) 2004 Syngenta Crop Protection AG, Basel, Switzerland Syngenta, Jealott's Hill, United Kingdom, Report No RJ3666B GLP Not Published Syngenta File N° NOA446510/0393	Y	SYN
Evans, P.	KIIA 7.3.1/09	2005 f	Mandipropamid 250 g/L SC formulation (A12946B): Dissipation in or on Soil in Spain (2004) Syngenta Crop Protection AG, Basel, Switzerland Syngenta, Jealott's Hill, United Kingdom, Report No RJ3665B GLP Not Published Syngenta File N° NOA446510/0373	Y	SYN
Evans, P.	KIIA 7.3.3/01 KIIIA	2005 g	Mandipropamid 250 g/L SC formulation: 6-Years Long Term Residue Study in or on Soil in Switzerland – Interim Report		

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N- R/NR	Owner
	9.2.3/01		Syngenta Crop Protection AG, Basel, Switzerland, Report No RJ3708B GLP Not Published Syngenta File N° NOA446510/0511		
Gibbings, E., Ricketts, D.	KIIA 7.2.3/02	2005	Rate of Degradation of Water Sediment Metabolite SYN504851 in Three Soils under Laboratory Conditions Syngenta Crop Protection AG, Basel, Switzerland Syngenta - Jealott's Hill International, Bracknell, Berkshire, United Kingdom, Report No RJ3652B GLP Not Published Syngenta File N° NOA446510/0396	Y	SYN
Grosjean, J., Hurt, A.	KIIA 7.8.3/01	2005	NOA446510: Degradation in Two Aquatic Sediment Systems (Methoxyphenyl Ring) Syngenta Crop Protection AG, Basel, Switzerland Syngenta, Jealott's Hill, United Kingdom, Report No RJ3580B GLP Not Published Syngenta File N° NOA446510/0388	Y	SYN
Hand, L H	KIIA 7.6/01	2003	Aqueous Photolysis of 14C- Methoxyphenyl-NOA446510 under Laboratory Conditions Syngenta - Jealott's Hill International, Bracknell, Berkshire, United Kingdom, Report No RJ3395B GLP Not Published Syngenta File N° NOA446510/0041	Y	SYN
Hand, L., Fleming, E.	KIIA 7.4.2/07	2005	Adsorption/Desorption Properties of a Metabolite (SYN539678) in Three Soils Syngenta Crop Protection AG, Basel, Switzerland Syngenta, Jealott's Hill, United Kingdom, Report No RJ3674B GLP Not Published Syngenta File N° SYN539678/0001	Y	SYN
Hand, L., Fleming, E.	KIIA 7.4.9/01	2004 a	Volatilisation from Soil Surfaces Syngenta Crop Protection AG, Basel, Switzerland	Y	SYN

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N- R/NR	Owner
			Syngenta, Jealott's Hill, United Kingdom, Report No RJ3464B GLP Not Published Syngenta File N° NOA446510/0079		
Hand, L., Fleming, E.	KIIA 7.4.9/02	2004 b	Volatilisation from Leaf Surfaces Syngenta Crop Protection AG, Basel, Switzerland Syngenta, Jealott's Hill, United Kingdom, Report No RJ3468B GLP Not Published Syngenta File N° NOA446510/0081	Y	SYN
Hand, L., Howdle, M.	KIIA 7.2.1/04	2004	NOA 446510: Rate of Degradation in One Soil Under Various Laboratory Conditions Syngenta Crop Protection AG, Basel, Switzerland Syngenta, Jealott's Hill, United Kingdom, Report No RJ3487B GLP Not Published Syngenta File N° NOA446510/0117	Y	SYN
Harrison, C.	KIIA 7.4.2/13	2005	NOA446510: Adsorption/Desorption Properties of a Water Sediment Metabolite SYN504851 in Three Soils Syngenta Crop Protection AG, Basel, Switzerland Syngenta, Jealott's Hill, United Kingdom, Report No RJ3633B GLP Not Published Syngenta File N° SYN504851/0011	Y	SYN
Harrison, C.	KIIA 7.6/03	2004	Methoxyphenyl Labelled Photolysis in Sterile Natural Water under Laboratory Conditions Syngenta Crop Protection AG, Basel, Switzerland Syngenta, Jealott's Hill, United Kingdom, Report No RJ3481B GLP Not Published Syngenta File N° NOA446510/0197	Y	SYN
Hurt, A., Bramley, Y., Grosjean, J., et., al.	KIIA 7.8.3/02	2005	NOA446510: Degradation in Two Aquatic Sediment Systems Syngenta Crop Protection AG, Basel,	Y	SYN

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N- R/NR	Owner
			Switzerland Syngenta - Jealott's Hill International, Bracknell, Berkshire, United Kingdom, Report No RJ3669B GLP Not Published		
Indergand, P, Nicollier, G.	KIIA 7.4.2/11	2005 c	Syngenta File N° NOA446510/0395 Adsorption / Desorption of [Phenyl-U-14C] SYN 500003 in Various Soils Syngenta Crop Protection AG, Basel,	Y	SYN
			Switzerland, Report No T006595-04 GLP Not Published Syngenta File N° SYN500003/0017		
Indergand, P., Nicollier, G.	KIIA 7.2.3/04	2005 a	Rate of Degradation of [Phenyl-U-14C]-labelled SYN500003 (Metabolite of NOA446510) in Various Soils under Aerobic Laboratory Conditions at 20°C Syngenta Crop Protection AG, Basel, Switzerland, Report No T006596-04	Y	SYN
			GLP Not Published Syngenta File N° SYN500003/0018		
Indergand, P., Nicollier, G.	KIIA 7.4.2/05	2005 b	Adsorption / Desorption of [Chlorophenyl-U-14C]-labelled SYN 521195 in Various Soils Syngenta Crop Protection AG, Basel, Switzerland, Report No T013384-04 GLP Not Published Syngenta File N° SYN521195/0004	Y	SYN
Kuet, S., Dick, J.	KIIA 7.1.3/01	2003	Soil Photolysis of 14C-Methoxyphenyl Ring labelled NOA446510 under Laboratory Conditions Syngenta Crop Protection AG, Basel, Switzerland Syngenta - Jealott's Hill International, Bracknell, Berkshire, United Kingdom, Report No RJ3400B GLP Not Published Syngenta File N° NOA446510/0056	Y	SYN
Kuet, S., Dick, J., Stapleton, C.	KIIA 7.1.1/01 KIIA	2004	Metabolism and Rate of Degradation of ¹⁴ C-Chlorophenyl Ring Labelled NOA446510 under Aerobic Laboratory	Y	SYN

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N- R/NR	Owner
	7.2.1/06		Conditions, in Three Soils at 20°C Syngenta Crop Protection AG, Basel, Switzerland Syngenta, Jealott's Hill, United Kingdom, Report No RJ3469B GLP Not Published Syngenta File N° NOA446510/0119		
Kuet, S., Stapleton, C.	KIIA 7.6/04	2004	Photolysis of 14C-Chlorophenyl Ring Labelled NOA446510 in Sterile Natural Water under Laboratory Conditions Syngenta Crop Protection AG, Basel, Switzerland Syngenta, Jealott's Hill, United Kingdom, Report No RJ3510B GLP Not Published Syngenta File N° NOA446510/0166	Y	SYN
Mamouni, A.	KIIA 7.2.3/03	2005 a	SYN536638: Degradation in three soils incubated under aerobic conditions Syngenta Crop Protection AG, Basel, Switzerland RCC Ltd., Itingen, Switzerland, Report No 859080 GLP Not Published Syngenta File N° SYN536638/0009	Y	SYN
Mamouni, A.	KIIA 7.4.2/02	2005 b	Adsorption / Desorption of SYN 536638 on Soils Syngenta Crop Protection AG, Basel, Switzerland RCC Ltd., Itingen, Switzerland, Report No RCC 856844 GLP Not Published Syngenta File N° SYN536638/0005	Y	SYN
Mamouni, A.	KIIA 7.4.2/03	2005 c	Adsorption of SYN 535839 on Soils Syngenta Crop Protection AG, Basel, Switzerland RCC Ltd., Itingen, Switzerland, Report No 855163 GLP Not Published Syngenta File N° SYN535839/0004	Y	SYN
Mamouni, A.	KIIA	2005	Adsorption of SYN500003 on Soils	Y	SYN

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N- R/NR	Owner
	7.4.2/10	d	Syngenta Crop Protection AG, Basel, Switzerland RCC Ltd., Itingen, Switzerland, Report No 855434 GLP Not Published Syngenta File N° SYN500003/0016		
Mamouni, A.	KIIA 7.4.2/12	2005 e	Adsorption of SYN504851 on Soils Syngenta Crop Protection AG, Basel, Switzerland RCC Ltd., Itingen, Switzerland, Report No 855435 GLP Not Published Syngenta File N° SYN504851/0009	Y	SYN
Nicollier, G	KIIA 7.4.1/01	2003 c	Adsorption/Desorption of [Methoxyphenyl-U-14C]-labelled NOA446510 in Various Soils Syngenta Crop Protection AG, Basel, Switzerland, Report No 02TB04 GLP Not Published Syngenta File N° NOA446510/0048	Y	SYN
Nicollier, G, Glänzel, A	KIIA 7.2.1/05	2002	Rate of Degradation of [Ethyl 1-14C] labelled NOA446510 in 'Gartenacker' and 'Borstel' Soil at Different Dose Levels under Aerobic Laboratory Conditions at 20 °C Syngenta Crop Protection AG, Basel, Switzerland, Report No 02GN01 GLP Not Published Syngenta File N° NOA446510/0017	Y	SYN
Nicollier, G.	KIIA 7.1.1/02 KIIA 7.1.2/01 KIIA 7.2.1/01 KIIA 7.2.4/01	2003 a	Metabolism of [Chlorophenyl-U-14C]- labelled NOA446510 under Aerobic and Aerobic/Anaerobic Laboratory Conditions in one Soil at 20 °C Syngenta Crop Protection AG, Basel, Switzerland, Report No 02TB03 GLP Not Published Syngenta File N° NOA446510/0050	Y	SYN
Nicollier, G.	KIIA 7.1.1/04	2003 b	Metabolism of [Methoxyphenyl-U-14C] Labelled NOA446510 under Aerobic,	Y	SYN

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N- R/NR	Owner
	KIIA 7.1.2/02 KIIA 7.2.1/02 KIIA 7.2.4/02		Aerobic / Anaerobic and Sterile Aerobic Laboratory conditions in One Soil at 20 °C Syngenta Crop Protection AG, Basel, Switzerland, Report No 02RF02 GLP Not Published Syngenta File N° NOA446510/0065		
Nicollier, G.	KIIA 7.6/02	2003 d	Aqueous Photolysis of [Chlorophenyl-U-14C]-labelled NOA446510 under Laboratory Conditions Syngenta Crop Protection AG, Basel, Switzerland, Report No 02GN07 GLP Not Published Syngenta File N° NOA446510/0051	Y	SYN
Nicollier, G., Berdat, T.	KIIA 7.4.1/02	2004	NOA 446510: Adsorption / Desorption of [Methoxyphenyl-U-14C]-labelled NOA 446510 in Various US Field Soils Syngenta Crop Protection AG, Basel, Switzerland, Report No 04TB01 GLP Not Published Syngenta File N° NOA446510/0214	Y	SYN
Oliver, R., Webb, J., Edwards, P.	KIIA 7.8.1/01	2005	NOA446510: Degradation in an Outdoor Aquatic Sediment System Syngenta Crop Protection AG, Basel, Switzerland Syngenta - Jealott's Hill International, Bracknell, Berkshire, United Kingdom, Report No RJ3569B GLP Not Published Syngenta File N° NOA446510/0400	Y	SYN
Schmidt, E.	KIIA 7.6/05	2004	Quantum yield of the direct photochemical degradation of NOA 446510 in aqueous solution Syngenta Crop Protection AG, Basel, Switzerland Solvias AG, Basel, Switzerland, Report No L02-009816 GLP Not Published Syngenta File N° NOA446510/0118	Y	SYN
Simon, P.	KIIA 7.3.1/06	2005 a	Mandipropamid 250 g/L SC: Residues in/on soil, Germany 2003	Y	SYN

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N- R/NR	Owner
			Syngenta Crop Protection AG, Basel, Switzerland Syngenta Agro GmbH, Maintal, Germany, Report No gbg714003 GLP Not Published Syngenta File N° NOA446510/0364		
Simon, P.	KIIA 7.3.1/10	2005 b	Mandipropamid 250 g/L SC: Residues in/on soil, Germany 2004 Syngenta Crop Protection AG, Basel, Switzerland Syngenta Agro GmbH, Maintal, Germany, Report No gbg711004 GLP Not Published Syngenta File N° NOA446510/0358	Y	SYN
Tummon, O.	KIIA 7.3.1/12	2005	Mandipropamid: Residue Stability Study for Mandipropamid (NOA466510) and CGA380778 in Soil under Freezer Storage Conditions Syngenta Crop Protection AG, Basel, Switzerland Syngenta - Jealott's Hill International, Bracknell, Berkshire, United Kingdom, Report No RJ3728B GLP Not Published Syngenta File N° NOA446510/0545	Y	SYN
Volkel, W.	KIIA 7.4.2/04	2005 a	Adsorption / Desorption of SYN 521195 on Soils Syngenta Crop Protection AG, Basel, Switzerland RCC Ltd., Itingen, Switzerland, Report No 855433 GLP Not Published Syngenta File N° SYN521195/0001	Y	SYN
Volkel, W.	KIIA 7.4.2/09	2005 b	Adsorption/Desorption of CGA380775 on Soils Syngenta Crop Protection AG, Basel, Switzerland RCC Ltd., Itingen, Switzerland, Report No 855162 GLP Not Published	Y	SYN

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N- R/NR	Owner
			Syngenta File N° CGA380775/0004		
Völlmin, S	KIIA 7.1.1/05 KIIA 7.3.1/11	2002	Field Dissipation of NOA446510 after Bareground Application of [1-14C] NOA446510 Syngenta Crop Protection AG, Basel, Switzerland, Report No 01SV09 GLP Not Published Syngenta File N° NOA446510/0013	Y	SYN
Wallace, S.	KIIA 7.7/01	2002	NOA446510 technical: Determination of 28 day ready biodegradability Syngenta Crop Protection AG, Basel, Switzerland Brixham Environmental Laboratory, Brixham, United Kingdom, Report No BL7358/B GLP Not Published Syngenta File N° NOA446510/0016	Y	SYN
Widmer, H	KIIA 7.10/01	2003	Atmospheric Oxidation of NOA446510 by Hydroxyl Radicals; Rate Estimation Syngenta Crop Protection AG, Basel, Switzerland, Report No 95A2003006WI Not GLP Not Published Syngenta File N° NOA446510/0035	Y	SYN

7.3.2 Aquatic Toxicity

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N- R/NR	Owner
Grade, R.	IIA 8.2.5	2003	Daphnia magna Reproduction Test: Effects of NOA 446510 on the Reproduction of the Cladoceran Daphnia magna STRAUS in a Semi-Static Laboratory Test. Syngenta Crop Protection AG, Basel,	Y	SYN

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N- R/NR	Owner
			Switzerland Report No 2013604 GLP Not Published Syngenta File N° NOA446510/0033		
Grade, R.	IIA 8.2.6	2001 a	Growth inhibition test of NOA 446510 to green algae (<i>Selenastrum capricornutum</i>) under static conditions Syngenta Crop Protection AG, Basel, Switzerland Report No 2013586 GLP Not Published Syngenta File N° NOA446510/0002	Y	SYN
Knauer, K.	IIA 8.2.6	2002	Growth inhibition test of NOA446510 tech. to Blue Algae (<i>Anabaena flos-aquae</i>) under static conditions Syngenta Crop Protection AG, Basel, Switzerland Syngenta AG, Basel, Switzerland Report No 2023553 GLP Not Published Syngenta File N° NOA446510/0023	Y	SYN
Matsuura, T.	IIA 8.2.1	2005	A 96-hour Acute Toxicity Test of NOA 446510 (Mandipropamid) with Common Carp Syngenta Crop Protection AG, Basel, Switzerland Kurume Laboratory, Chemical Biotesting Centre, Fukuoka Prefecture, Japan Report No 93451 GLP Not Published Syngenta File N° NOA446510/0254	Y	SYN
Maynard, S.	IIA 8.2.1	2005 b	NOA446510 metabolite (SYN504851): Acute toxicity to rainbow trout (Oncorhynchus mykiss) Syngenta Crop Protection AG, Basel, Switzerland Brixham Environmental Laboratory, Brixham, United Kingdom Report No BL7967/B GLP Not Published Syngenta File N° SYN504851/0001	Y	SYN
Maynard, S.	IIA 8.2.2.2	2003	NOA446510 tech: Early-life stage toxicity test to the fathead minnow (<i>Pimephales promelas</i>) Syngenta Crop Protection AG, Basel, Switzerland	Y	SYN

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N- R/NR	Owner
			Brixham Environmental Laboratory, Brixham, United Kingdom Report No BL7577/B GLP Not Published		
Maynard, S.	IIA 8.2.4	2005 c	Syngenta File N° NOA446510/0063 NOA446510 metabolite (SYN504851): Acute toxicity to Daphnia magna in a 48-hour immobilization test Syngenta Crop Protection AG, Basel, Switzerland Brixham Environmental Laboratory, Brixham, United Kingdom Report No BL7968/B GLP Not Published Syngenta File N° SYN504851/0002	Y	SYN
Maynard, S.	IIA 8.2.6	2005 d	NOA446510 metabolite (SYN504851): Acute toxicity to the green alga Pseudokirchneriella subcapitata (formerly Selenastrum capricornutum) Syngenta Crop Protection AG, Basel, Switzerland Brixham Environmental Laboratory, Brixham, United Kingdom Report No BL7969/B GLP Not Published Syngenta File N° SYN504851/0003	Y	SYN
Maynard, S., Woodyer, J.	IIA 8.2.1	2004 a	NOA446510: Acute toxicity to common carp (<i>Cyprinus carpio</i>) in a flow-through test system Syngenta Crop Protection AG, Basel, Switzerland Brixham Environmental Laboratory, Brixham, United Kingdom Report No BL7872/B GLP Not Published Syngenta File N° NOA446510/0111	Y	SYN
Maynard, S., Woodyer, J.	IIA 8.2.4	2004 b	NOA446510: Acute toxicity to <i>Daphnia magna</i> Syngenta Crop Protection AG, Basel, Switzerland Brixham Environmental Laboratory, Brixham, United Kingdom Report No BL7871/B GLP Not Published Syngenta File N° NOA446510/0112	Y	SYN
Maynard, S.,	IIA 8.2.1	2005	NOA446510 metabolite (SYN500003):	Y	SYN

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N- R/NR	Owner
Young, B.		a	Acute toxicity to rainbow trout (Oncorhynchus mykiss) Syngenta Crop Protection AG, Basel, Switzerland Brixham Environmental Laboratory, Brixham, United Kingdom Report No BL7964/B GLP Not Published Syngenta File N° SYN500003/0003		
Maynard, S., Young, B.	IIA 8.2.4	2005 b	NOA446510 metabolite (SYN500003): Acute toxicity to <i>Daphnia magna</i> in a 48-hour immobilization test Syngenta Crop Protection AG, Basel, Switzerland Brixham Environmental Laboratory, Brixham, United Kingdom Report No BL7965/B GLP Not Published Syngenta File N° SYN500003/0004	Y	SYN
Maynard, S., Young, B.	IIA 8.2.6	2005 c	NOA446510 metabolite (SYN500003): Acute toxicity to the green alga Pseudokirchneriella subcapitata (formerly Selenastrum capricornutum) Syngenta Crop Protection AG, Basel, Switzerland Brixham Environmental Laboratory, Brixham, United Kingdom Report No BL7966/B GLP Not Published Syngenta File N° SYN500003/0005	Y	SYN
Palmer, S., Kendall, T., Krueger, H.	IIA 8.2.1	2005 a	NOA-446510 - A 96-Hour Flow-Through Acute Toxicity Test with the Sheepshead Minnow (Cyprinodon variegatus) Syngenta Crop Protection AG, Basel, Switzerland Wildlife International Ltd., Easton, MD, United States Report No WIL 528A-138 GLP Not Published Syngenta File N° NOA446510/0287	Y	SYN
Palmer, S., Kendall, T., Krueger, H.	IIA 8.2.4	2005 b	NOA-446510 - A 96-Hour Flow-Through Acute Toxicity Test with the Saltwater Mysid (<i>Americamysis bahia</i>) Syngenta Crop Protection AG, Basel, Switzerland Wildlife International Ltd., Easton, MD, United States	Y	SYN

Author(s)	Annex point/ reference number	Year	Title Source (where different from company) Company, Report No GLP or GEP status (where relevant) Published or not	Data Protection Claimed Y/N- R/NR	Owner
			Report No WIL 528A-137 GLP Not Published Syngenta File N° NOA446510/0290		
Palmer, S., Kendall, T., Krueger, H.	IIA 8.2.4	2005 c	NOA-446510 - A 96-Hour Flow-Through Shell Deposition Test with the Eastern Oyster (<i>Crassostrea virginica</i>) Syngenta Crop Protection AG, Basel, Switzerland Wildlife International Ltd., Easton, MD, United States Report No WIL 528A-139 GLP Not Published Syngenta File N° NOA446510/0288	Y	SYN
Peter, P.	IIA 8.2.1	2002	Acute Toxicity Test of NOA446510 to Fathead Minnow (<i>Pimephales promelas</i>) Under Static Conditions Syngenta Crop Protection AG, Basel, Switzerland Report No 2023555 GLP Not Published Syngenta File N° NOA446510/0011	Y	SYN
Roberts, G., Peurou, F.	IIA 8.2.3	2003	NOA446510: Determination of the accumulation and elimination of [14C]NOA446510 in fathead minnow (<i>Pimephales promelas</i>) Syngenta Crop Protection AG, Basel, Switzerland Brixham Environmental Laboratory, Brixham, United Kingdom Report No BL7579/B GLP Not Published Syngenta File N° NOA446510/0062	Y	SYN
Volz, E.	IIA 8.2.1	2002	Acute Toxicity Test of NOA446510 to Rainbow Trout (<i>Oncorhynchus mykiss</i>) Under Static Conditions Syngenta Crop Protection AG, Basel, Switzerland Report No 2023552 GLP Not Published Syngenta File N° NOA446510/0012	Y	SYN

8 ANNEXES