

Recommendation from the Scientific Committee on Occupational Exposure Limits for epichlorohydrin

SCOEL/SUM/169 September 2011



European Commission

Table of content

1 (Docurrend	ce/use and occupational exposure	4				
2 ŀ	lealth sigi	nificance	4				
1	2.1 Toxico	kinetics	4				
	2.1.1	Human data	4				
	2.1.2	Animal data	4				
	2.1.3	Biological monitoring	4				
1	2.2 Acute	toxicity	5				
	2.2.1	Human data	5				
	2.2.2	Animal data	5				
1	2.3 Irritatic	on and corrosivity	5				
	2.3.1	Human data	5				
	2.3.2	Animal data	6				
1	2.4 Sensiti	sation	6				
	2.4.1	Human data	6				
	2.4.2	Animal data	6				
1	2.5 Repec	ated dose toxicity	7				
	2.5.1	Human data	7				
	2.5.2	Animal data	7				
1	2.6 Genot	toxicity	7				
	2.6.1	In vitro	8				
	2.6.2	In vivo - human data	8				
	2.6.3	In vivo – animal data	8				
1	2.7 Carcir	nogenicity	9				
	2.7.1	Human data	9				
	2.7.2	Animal data	0				
	2.7.3	Mode of action and cancer risk assessment	1				
2.8 Reproductive toxicity							
	2.8.1.	Human data	2				
	4.9.2	Animal data	2				
Re	commen	dation	3				
Re	References						

Recommendation from the Scientific Committee on

Occupational Exposure Limits for

Epichlorohydrin

	8-hour TWA: not a STEL (15 min): not a		ssigned (see "Recommendation" page 11) ssigned (see "Recommendation" page 11)				
	Additional classification:		skin notation (see "Recommendation" page 11)				
	BLV:		not assigned				
	SCOEL carcinogen (group:	A (non-threshold carcinogen)				
	Carcinogenic risk assessment: see "Recommendation" (page 11)						

<u>Substance identificati</u>	i <u>on</u> : 1-c	1-chloro-2,3-epoxypropane			
<u>Synonyms:</u>	ch	chloropropylene oxide, chloromethyl-oxirane			
Structural formula:		$H_2C - CH - CH_2CI$			
CAS no.: Molecular formula: Molecular weight: Melting point: Boiling point:	106-89-8 C₃H₅CIO 92.53 -48°C 116°C				
EU-Classification:					
Flam. Liq. 3 Carc. 1B Acute Tox. 3 * Acute Tox. 3 * Acute Tox. 3 * Skin Corr. 1B Skin Sens. 1		H226 H350 H331 H311 H301 H314 H317	Flammable liquid and vapour May cause cancer Toxic if inhaled Toxic in contact with skin Toxic if swallowed Causes severe skin burns and eye damage May cause an allergic skin reaction		

<u>Conversion factor:</u> 1 ppm = 3.84 mg/m³; 1 mg/m³ = 0.260 ppm (DFG 2009)

<u>Criteria documents used</u>: This summary is mainly based on the recent documentation of DFG (2009), which includes data reported earlier by BUA (1992), IARC (1999) and DFG (2003). This was supplemented by a recent literature search by SCOEL.

1 Occurrence/use and occupational exposure

Epichlorohydrin is a major raw chemical for the production of epoxy and phenoxy resins. It and is also used in the manufacture of glycerine, in curing propylene-based rubbers, as a solvent for cellulose esters and ethers, and in resins with high wet-strength for the paper industry (IARC 1999, NTP 2002).

2 Health significance

Epochlorohydrin is a directly acting alkylating agent. With nucleophiles it reacts preferably with the epoxide ring, but also with the chlorine function. The biological properties of the compound are related to this reactivity.

2.1 Toxicokinetics

2.1.1 Human data

Incubation of epichlorohydrin in the presence of human cells of lung and bronchial parenchyma led to a decrease of mutagenicity, which is likely to be ralated to rapid metabolic inactivation (De Flora *et al.* 1984).

2.1.2 Animal data

After both inhalation and oral administration, more than 90% epichlorohydrin was rapidly absorbed and distributed in the organism of rats within 2–4 hours (CMA 1979a; Gingell *et al.* 1985; Weigel *et al.* 1978). After application of radiolabeled epichlorohydrine, the highest systemic tissue concentrations were reached in the kidneys, intestine, liver, lacrimal glands, pancreas and spleen. The highest level of radioactivity was found in the stomach after oral administration and in the nasal mucosa after inhalation. Lower radioactivity was detected in the blood, lungs, brain and sex organs (Weigel *et al.* 1978). At different dose levels and with various types of administration of ¹⁴C-labelled epichlorohydrin, 90% of the activity was excreted within 72 hours, *i.e.* 46–54% in the urine, 25–40% via the lungs and maximally 4% via the faeces (CMA 1979a; Gingell *et al.* 1985).

After initial reaction with glutathione, epichlorohydrin is metabolized, leading to metabolites that are detected in the urine. These are the mercapturic acid derivatives N-acetyl-S-(3-chloro-2-hydroxypropyl)-L-cysteine, S-(2,3-dihydroxypropyl)-L-cysteine and N-acetyl-S-(2,3-dihydroxypropyl)-L-cysteine. In addition, a likewise excretable 3-chloro-1,2-propanediol is formed from hydrolysis. N-Acetyl-S-(3-chloro-2-hydroxypropyl)-L-cysteine (36% of the dose) and 3-chloro-1,2-propanediol (4%) are considered to be the main metabolites (Gingell *et al.* 1985). According to Fakhouri and Jones (1979), 3-chloro-1,2-propanediol *in vivo* leads to the formation of β -chlorolactic acid, which in turn yields oxalic acid. The latter metabolite was however not detected in other studies.

2.1.3 Biological monitoring

The analysis of haemoglobin adducts derived from epichlorihydrin has been proposed as a means of biological monitoring. As epichlorohydrin is a bifunctional alkylating agent (Romano et al. 2007), the reaction with the N-terminal valine of haemoglobin occurs either with its chlorine or with its epoxide moiety. N-(2,3-Dihydroxypropyl)valine was first described as haemoglobin adduct. This was also detected in rats after intraperitoneal administration of 40 mg/kg body weight. An increased amount of it was found in smokers (Hindsø Landin et al. 1996).

Later, Bader *et al.* (2009) described a method of quantitation of the adduct N-(3-chloro-2hydroxypropyl)valine in human haemoglobin. This adduct was found in the blood of persons exposed to epichlorohydrin at a freight train accident. These methods could provide a basis for biomonitoring in the future. However, more detailed occupational field studies are lacking so far.

2.2 Acute toxicity

2.2.1 Human data

An airborne concentration of 20 ppm epichlorohydrin caused corrosion to the eyes and nasal mucosa after one hour. 40 ppm led to irritation to the eyes and throat, which lasted for 48 hours (no other details on exposure period). 100 ppm was intolerable even for the shortest period. Exposure of the eyes to the liquid substance led to opacity and necrosis of the cornea (no other details; Lefaux 1966).

Cases of severe skin burns after contact with epichlorohydrin were described. Erythema, oedema and papules were observed during some days after direct skin contact. The persons affected complained about agonizing itching. The symptoms had subsided in all persons only after about two weeks (Ippen and Mathies 1970).

Schultz (1964) reported a person who had developed chronic asthmatic bronchitis after inhalation exposure to epichlorohydrin. Severe fatty degeneration of the liver was diagnosed bioptically.

2.2.2 Animal data

After exposure periods between 2 and 8 hours, the LC₅₀ was 3000 to 250 ppm and the 4hour LC₅₀ for rats was 500 ppm (Hine *et al.* 1981; Pallade *et al.* 1967; Pozzani and Carpenter 1960; Srám *et al.* 1981). An oral LD₅₀ of about 220 mg/kg body weight was determined in rodents (Hine *et al.* 1981; Lawrence *et al.* 1972; Pozzani and Carpenter 1960), and the dermal LD₅₀ established in rabbits was 754 (Lawrence *et al.* 1972) and 1038 mg/kg body weight (Hine *et al.* 1981). CNS, respiratory tract or renal lesions were specified as causes of death depending on the type of administration (Laskin *et al.* 1980; Pallade *et al.* 1967; Weigel *et al.* 1978).

In an inhalation study, the respiratory rate of rats was clearly reduced within 15 minutes at a concentration of 363 ppm, and halved at 1342 ppm. Marked discharge from the nasal mucosa was observed at 1963 ppm (Gardner *et al.* 1985).

2.3 Irritation and corrosivity

2.3.1 Human data

A 46-year-old pharmaceutical company worker developed severe erythema and oedema on the face, neck, back and hands after having continuously been exposed to epichlorohydrin for 11 months. The symptoms subsided after the worker left the workplace for two weeks, but returned when he resumed work at his former workstation. They cannot be assigned to epichlorohydrin with certainty since there was co-exposure to other substances in the synthesis of propanolol and oxprenolol (Rebandel and Rudzki 1990).

Luo et al. (2004) investigated the influence of the glutathione S-transferase hGSTM1 and hGSTT1 genes on the toxic effect of epichlorohydrin. In hGSTM1 null genotype workers, there was a dose-response of lung function tests (FEV1, FEV1/FVC, MMEF) for epichlorohydrin exposure, but not in the hGSTM1 non-null genotype workers. The exposure was found to be significantly associated with a decreased FEV1 value (p = 0.09) and a decreased MMEF value (p = 0.053) after adjusting for other factors. The hGSTM1 null genotype was found to be significantly associated with a decreased FEV1 value (p = 0.09) and a decreased FEV1 value (p = 0.053), decreased FEV1 value (p = 0.056), and decreased MMEF value (p = 0.012) after adjusting for other factors. The study was interpreted to indicate that obstructive lung abnormalities and small airway lung damage are associated with epichlorohydrin exposure.

The irritancy of epichlorohydrine to the respiratory tract is in accordance with human experience after an accidental release of epichlorohydrine (Basting et al. 2006)

2.3.2 Animal data

Experimental exposures of rats and rabbits to 25 or 50 ppm epichlorohydrine (6h/d, 5d/wk for 10 weeks) caused severe irritating effects to the nasal turbinates (John et al. 1983a). In mice, the RD_{50} concentration was assessed to be 687 ppm. In addition to the nasal cavity, this concentration induced lesions in the lower respiratory tract, when applied 6h/d for 5 days (Buckley et al. 1984).

2.4 Sensitisation

2.4.1 Human data

In a Dutch plant for the production of epoxy resins, 26 cases of eczema were observed among 228 male workers. All workers were occupationally exposed to epichlorohydrin and bisphenol A, the starting materials of the manufacturing process. The authors reported that the workers had unintentional skin contact with bisphenol A epoxy resins, for example during maintenance operations, although closed systems were used for manufacturing the epoxy resins and protective measures such as wearing gloves had been taken. A patch test was carried out with 19 of the 26 workers. In addition to other substances, the two epoxy resins (molecular weight 385 and 980), which were the main production products, and bisphenol A, 1% epichlorohydrin in petrolatum and 1% in isopropanol, were tested. In the patch test, a positive reaction to 1% epichlorohydrin in petrolatum was obtained in 8 cases; it was isolated in 4 cases and occurred together with reactions to epoxy resins in 4 cases. The observed delayed-type sensitization to epichlorohydrin may have been caused by two routes of sensitization: first by direct skin contact and second aerogenically by increased airborne concentrations due to the volatility of epichlorohydrin (Prens *et al.* 1986).

The same research group reported another 6 cases (5 workers of a plant for the production of epoxy resins and a vehicle painter) of occupational contact dermatitis, which was considered to be caused by exposure to epichlorohydrin or bisphenol A epoxy resins. In addition to other substances, an epoxy resin with an average molecular weight of 385 as well as bisphenol A and 1% epichlorohydrin in solvent were patch-tested. All patients showed delayed-type reactions to epichlorohydrin, two of them to epichlorohydrin alone (van Joost 1988).

Another publication by this research group focussed on 5 workers of a plant for the production of epoxy resins who had developed contact dermatitis at their workplace. Here, too, the authors saw a connection with the delayed-type reaction to 1% epichlorohydrin in petrolatum, which they detected in the patch test in all 5 patients. A reaction to epichlorohydrin alone was found in 2 patients (van Joost *et al.* 1990).

Moreover, other research groups described case reports of contact sensitization to epichlorohydrin that had not been acquired in epoxy resin production (for a detailed description, see DFG 2009).

2.4.2 Animal data

Skin sensitization was examined in a Guinea pig maximization test in 15 Guinea pigs (20 animals in the control group). Intra-dermal and dermal inductions were carried out with 5% epichlorohydrin in ethanol, and 1% epichlorohydrin in ethanol was used for dermal challenge. A positive reaction was observed in 9 of 15 animals (Thorgeirsson and Fregert 1977).

No sensitization was found in another, insufficiently documented maximization test in 5 Guinea pigs at a test concentration of 0.01% epichlorohydrin in vegetable oil (Lawrence *et al.* 1972). However, this study has only limited applicability for assessment since the number of animals was too small, the test substance concentration might have been too low and insufficient information was provided on range finding. Another study in which none of 18 Guinea pigs reacted after 8 intracutaneous injections during challenge (Weil *et al.* 1963)

cannot be assessed either because of inadequate documentation (e.g. test substance concentrations not specified).

In a modified test carried out according to Landsteiner in 10 guinea pigs dermally treated four times with epichlorohydrin for induction (as well as a single intradermal treatment with Freund's adjuvant), all 10 animals reacted positively in the dermal challenge (no other details for example on the test substance concentration used; Rao *et al.* 1981).

2.5 Repeated dose toxicity

2.5.1 Human data

No published data are available.

2.5.2 Animal data

In a 90-day study (CMA 1979b; Quast *et al.* 1979), 20 female and 20 male B6C3F1 mice, Fischer 344 rats and Sprague-Dawley rats were exposed to epichlorohydrin for 6 hours on 5 days per week at concentrations of 5, 25 and 50 ppm. Whereas no effects were recorded at 5 ppm, hyperplasia, metaplasia and inflammatory infiltrates were found in the nose, the most sensitive organ, at the higher concentrations. Damage to the kidneys, liver and adrenals of varying severity was observed in the different animal strains.

In a study described by BUA (1992), continuous inhalation of 5 ppm by rats over 98 days led to weight loss and morphological changes to the liver, heart, kidneys and CNS, to increased urinary coproporphyrin and an increase of leukocytes in the peripheral blood. After inhalation of 0.05 and 0.5 ppm, these effects were not found or were only slight as compared with the controls. The validity of this study is unclear.

Four applications of 600 mg/kg body weight were fatal for 10/10 rats and three applications of 1200 mg/kg body weight were fatal for 4/10 rats (Freuder and Leake 1941).

After intraperitoneal injection of 11, 22 and 56 mg/kg body weight three times a week over a period of 12 weeks, there was a dose-dependent, significant decrease of haemoglobin in the blood, an increase of eosinophils in all treated animals and a reduction of lymphocytes in the two groups treated with the highest doses. The weights of heart, liver and kidneys increased in the animals treated with 56 mg/kg body weight (Lawrence *et al.* 1972).

Adult male and female Sprague-Dawley rats received epichlorohydrin via gavage in distilled water for 10 consecutive days at dose levels of 3, 7, 19, and 46 mg/kg per day, and for 90 days at dose levels of 1, 5, and 25 mg/kg per day. Epichlorohydrin did not adversely effect mortality, but toxicity, at the higher doses, was evident by losses in body weight gain and organ weights, reductions in food and water consumption, and in the hematological and microscopic examinations in both study periods. Significant decreases in erythrocyte count, hemoglobin, and hematocrit levels were found in the high dose level in males after 10 and 90 days. Dose-related increases in kidney and liver weights were observed in both sexes at 25 mg/kg per day in the 90-day study and in various organs for both 19 and 46 mg/kg per day in the 10-day study. Histopathological examination identified the forestomach as the primary target organ for both sexes and in both studies with significant dose-related increases in mucosal hyperplasia (acanthosis) and hyperkeratosis. Based on the data presented, a lowest observable adverse effect level (LOAEL) for oral exposure of Sprague-Dawley rats to epichlorohydrin was 3 mg/kg per day for 10 days and 1 mg/kg per day was suggested as the no observed adverse effect level (NOAEL) for a 90 day oral exposure (Daniel et al. 1996).

2.6 Genotoxicity

Detailed reviews on the genotoxic effects of epichlorohydrin are available by Giri (1997), IARC (1999) and Kolman et al. (2002). The main results are summarized below.

2.6.1 In vitro

In most of the *in vitro* test systems used, epichlorohydrin induced genotoxic effects that were almost always observed even in the absence of an added metabolic activation system. Studies in bacterial test systems showed that epichlorohydrin led to DNA lesions in *E. coli* and *B. subtilis* and induced gene mutations in *S. typhimurium* and *E. coli* strains and in *Klebsiella pneumoniae*. It caused DNA lesions, recombinations, gene mutations and aneuploidies in yeasts. Epichlorohydrin induced DNA single strand breaks and SCE in mammalian cells. Induction of gene mutations at different loci and of chromosome mutations was detected in numerous studies (IARC 1999).

The main adduct obtained after reaction of epichlorohydrin with 2'-deoxynucleosides in vitro is 7-(3-chloro-2-hydroxypropyl)guanine, resulting from reaction of the epoxide ring of epichlorohydrine (Singh *et al.* 1996). Another aspect of DNA interaction is the formation of interstrand DNA-cdrosslinks (Romano *et al.* 2007).

Holzer et al. (2008) incubated rat and human nasal mucosa cells with epichlorohydrin and used the COMET assay as an endpoint of genotoxicity. In contrast to the cells derived from rats, pronounced interindividual differences in susceptibility were found with the human samples.

2.6.2 In vivo - human data

The DNA adduct 7-(3-chloro-2-hydroxypropyl)guanine (vs.) was detected at a concentration of 0.8–7.1 adducts/10⁹ nucleotides in the lymphocytes of workers who were classified as exposed to epichlorohydrin on account of the fact that they worked in an epichlorohydrin-processing plant. No details are available about the level of exposure. This adduct was not found in non-exposed control persons (Plna *et al.* 2000).

Significantly increased sister chromadid exchange (SCE) frequencies were detected in the lymphocytes of 21 workers with high exposure to epichlorohydrin (4.5 years; 1.1–3.9 ppm) compared with 29 non-exposed control persons adjusted for age. Smoking was excluded as the only cause of the increase. The SCE frequency in 35 workers with low exposure (4.2–7.0 years; 0.1–0.2 ppm) was not significantly increased compared with the control persons (Cheng *et al.* 1999).

The lymphocytes of workers who were exposed to 0.4–0.86 mg/m³ (0.11–0.23 ppm) during a 12-hour shift showed no increased frequency of hprt mutations, a slight increase of micronuclei and a significant increase of SCE and high frequency cells (> 10 SCE per cell) (Hindsø Landin *et al.* 1997).

Kucerová et al. (1977) found significantly increased frequencies of structural chromosomal aberrations (chromatid and chromosome breaks) in the lymphocytes of workers occupationally exposed to epichlorohydrin concentrations of 0.125 to 1.25 ppm. The workers were examined before the beginning of exposure (1.37 aberrations/cell), one year (1.91 aberrations/cell) and two years (2.69 aberrations/cell; p < 0.001) after the beginning of exposure. Šrám et al. (1980) re-examined 23 of these workers after 4-year exposure (3.02 aberrations/cell) and compared them with an adjusted control group (for age, smoking and drinking habits; n = 34; 2.06 aberrations/cell; p < 0.01) and with the general population (n = 21; 1.33 aberrations/cell; p < 0.01).

In another study of Picciano (1979), 93 exposed persons (no concentrations specified; presumably 5 ppm TWA; age 35.8 years) and 75 control persons (age 25.2 years) were examined. The frequencies of cells with chromatid breaks, chromosome breaks and marker chromosomes (rings, dicentric chromosomes and translocations), of severely damaged cells and of the total number of damaged cells were significantly higher (p < 0.005) in the exposed group than in the control group.

2.6.3 In vivo – animal data

In a study on the ability of epichlorohydrin to bind covalently to macromolecules, the [2-1⁴C]-labelled substance (6.35 micromol/kg body weight) was intraperitoneally injected into mice and rats. In mice, an association of radioactivity with the purified DNA from liver, lung, kidney and stomach, which was quantitatively similar for all organs, was observed 22 hours after administration. A covalent binding index (CBI) of 23 was determined for rat liver DNA. The corresponding value for benzene was 7 and that for 1,2-dibromoethane was 515 (Prodi *et al.* 1986).

In another study in rats, a quantitatively similar binding to the DNA of various organs was detected 6 and 24 hours after intraperitoneal injection of [2-³H]epichlorohydrin (0.97 µmol/kg body weight), and N7-(3-chloro-2-hydroxypropyl)guanine was identified as the main DNA adduct. A CBI value of 0.6 was determined. A CBI value of 2 was found by the same working group for the chemically less reactive propylene oxide. This discrepancy was attributed to a relatively more rapid elimination of epichlorohydrin (Hindsø Landin *et al.* 1999).

Epichlorohydrin induced X chromosomal recessive lethal mutations in the fruit fly Drosophila melanogaster (Knaap et al. 1982; Vogel et al. 1981). A third test had a negative result (Würgler and Graf 1981).

Epichlorohydrin (50 and 100 mg/kg body weight) was administered intramuscularly or subcutaneously in a host-mediated assay with NMRI mice and the *Salmonella* strains G46, TA100, TA1950, TA1951 and TA1952 (Srám *et al.* 1976). An increased rate of revertants was found for the strains G46, TA100 and TA1950. In another assay with *Schizosaccharomyces pombe* (after intraperitoneal administration of the yeast suspension and intraperitoneal treatment or intrasanguinous administration of the yeast suspension and intravenous treatment) and two different mouse strains [CD1 and (CD1xC57BL)F1], negative results were reported by Rossi *et al.* (1983b) for doses between 2 and 100 mg/kg body weight.

A third assay with NMRI mice and *Escherichia coli* strains K-12 uvrB/recA (mainly cell death of the repair-deficient bacterial strains) yielded negative results after orally administered 240 mg/kg body weight or intraperitoneally injected 140 mg/kg body weight (Hellmér and Bolcsfoldi 1992).

Increased sperm head anomalies were described in a study in mice (11 days after a single oral administration of 50 mg/kg body weight; Cassidy *et al.* 1983). This finding was however not confirmed in a second study with intraperitoneal injection of 0.025–0.2 ml/kg body weight and day for 5 days (Topham 1980). Since morphological sperm anomalies are generally not interpreted as mutations, these results are not relevant.

Epichlorohydrin induced chromosomal aberrations in the bone marrow of ICR mice in a dose range of 1–50 mg/kg body weight (in DMSO) after single or several (on 5 consecutive days) intraperitoneal and oral administrations (Srám *et al.* 1976).

No significant increase in the incidence of chromosomal aberrations was found in the bone marrow of CD1 mice in another study after oral administration of 50 or 200 mg/kg body weight (Rossi *et al.* 1983a). The authors attribute the negative result to the fact that epichlorohydrin was no longer detectable in the blood as early as 20 minutes after oral (in DMSO) or intraperitoneal (in water) administration.

Several authors obtained uniformly negative results in micronucleus tests with mice (Asita *et al.* 1992; Kirkhart 1981; Salamone *et al.* 1981; Tsuchimoto and Matter 1981).

Nor did epichlorohydrin induce any dominant lethal mutations (Epstein *et al.* 1972). The originally negative result was confirmed in detailed investigations (once intraperitoneally 5, 10 and 20 mg/kg body weight; once orally 20 and 40 mg/kg body weight; five times intraperitoneally 1 and 4 mg/kg body weight; five times orally 4 and 16 mg/kg body weight) (Srám *et al.* 1976).

2.7 Carcinogenicity

2.7.1 Human data

In a nested case-control study by Bond *et al.* (1986) among 19608 workers of a chemical production facility who were examined for possible health damage caused by carbon tetrachloride, a lowering of lung cancer mortality was found for the very small subcohort of persons ever exposed to epichlorohydrin (odds ratio 0.3; 95% CI 0.1–0.9; 5 cases).

Barbone et al. (1992, 1994) studied the frequency of lung cancer and CNS cancer in a nested case-control study based on a cohort previously examined by Delzell et al. (1989).

A positive association was found between potential exposure to epichlorohydrin and lung cancer after adjustment for smoking habits (odds ratio 1.7; 95% CI 0.7-4.1; 51 cases), but not in the calculation with regard to exposure period and cumulative dose. An association with potential exposure to epichlorohydrin was detected in 11 cases with CNS cancer (7 with brain tumours, 2 with meningiomas and 2 with benign tumours) and 44 controls matched for age (odds ratio 4.2; 95% CI 0.7-26). Associations with the exposure period (p = 0.11) and cumulative exposure (p = 0.08) were also observed. These results are not statistically significant. The small number of cases must be considered.

Delzell et al. (1989) reported an excess of lung cancer in a cohort study among 2642 male workers with at least six-month exposure to epichlorohydrin. At an expected level of 0.91 (p < 0.03), 4 of 44 persons exposed in the production of the substance developed lung cancer.

Tsai et al. (1996) reported a cohort of 863 workers who had previously been examined by Enterline (1982) and Enterline et al. (1990). The rate of workers affected by lung cancer did not increase (SMR 0.7; 95% CI 0.5-1.1; 23 cases). Increased incidence rates of prostate cancer (SMR 2.3; 95% CI 1.0-4.5; 8 cases) and malignant melanomas (SMR 3.2; 95% CI 0.7-9.4; 3 cases) were found among workers whose initial exposure had taken place at least 20 years before. No relation between the frequency of developing cancer and the estimated exposure level was obtained in this study.

Olsen et al. (1994) described results of a retrospective cohort study on cancer mortality among 1064 male workers in the production areas for epoxy resins, glycerol and allyl chloride/epichlorohydrin. Exposures to epichlorohydrin in glycerol production (highest exposures) were between 1 ppm and 5 ppm before 1970 and below 1 ppm after 1970. A total of 66 cohort members died up to 1989, 10 of them from cancer (SMR 0.5; 95% CI 0.2-0.9).

2.7.2 Animal data

In a lung adenoma test with intraperitoneal administration of 20, 50 and 100 mg/kg body weight to an A/J strain of mice three times a week for 8 weeks, significantly more lung tumours were induced only in the males that had received the highest dose (Stoner et al. 1986).

In an initiation-promotion study, 2 mg epichlorohydrin in 0.1 ml acetone was applied once to the skin of 30 ICR/Ha Swiss mice. After 2 weeks, 2.5 µg phorbol myristate acetate in 0.1 ml acetone was applied three times a week for a period of 385 days. From the 92nd day, skin papillomas developed in 9 animals and a skin carcinoma in one animal. A skin papilloma was observed in 3 of the mice treated with phorbol myristate acetate alone after about 224 days, whereas the control group treated with acetone alone developed no tumours (Van Duuren et al. 1974). Another study carried out with epichlorohydrin as an initiator in 20 mice was negative (Van Duuren et al. 1972).

In a whole-body inhalation study carried out by Laskin et al. (1980), groups of 100 male Sprague-Dawley rats were subjected to lifetime exposure to 0, 10 and 30 ml epichlorohydrin/m³ (purity 99%) for 5 hours daily on 5 days per week. Two further groups of 100 and 40 male rats were exposed to 100 ppm 6 hours daily for 30 days and then observed during their entire lifespan. One group of 100 male controls was sham-exposed and another group of 50 control animals remained untreated. No tumours developed after exposure to 10 ppm. Exposure to 30 ppm yielded a nasal papilloma in one rat after 40 days and a squamous cell carcinoma of the nasal cavity in a second rat after 752 days. Among the 140 rats that had been exposed to 100 ppm for 30 days, 15 rats developed squamous cell carcinomas and 2 rats developed nasal papillomas between days 330 and 933 of the study. One bronchial papilloma was observed on day 583 after the beginning of the study. Four of the exposed rats developed pituitary adenomas; a squamous cell carcinoma in the forestomach and further tumours were found in one animal. A total of 5 tumours occurred in the 150 control animals: 3 subcutaneous fibromas, 1 forestomach papilloma and 1 malignant lymphoma. The authors regarded the respiratory tract tumours as being related to exposure, unlike the other tumours.

Konishi et al. (1980) administered 0, 375, 750 and 1500 mg epichlorohydrin/l (purity not specified) to 6-week-old Wistar rats with the drinking water over a period of 81 weeks. The

10

animals were then sacrificed and the tissues examined histopathologically. Hyperplasias and forestomach tumours were found in the treated rats in relation to the dose in the order of the specified doses: hyperplasias: 0/10, 7/9, 9/10, 12/12; papillomas: 0/10, 0/9, 1/10, 7/12; carcinomas: 0/10, 0/9, 1/10, 2/12. No tumours were detected in other tissues.

Wester *et al.* (1985) administered daily doses of 0, 2 and 10 mg epichlorohydrin/kg body weight with a purity of 99.5% by gavage to groups of 50 newly weaned female and male Wistar rats daily on 5 days per week over a period of 2 years. Subsequently the animals was sacrificed. A dose-dependent increase of hyperplasias, papillomas and forestomach carcinomas was observed. In the order of the specified doses, the males showed 5/50, 24/49 and 6/49 hyperplasias, 1/50, 6/49 and 4/49 papillomas and 0/50, 6/49 and 35/49 carcinomas. The females revealed 3/47, 12/44 and 7/39 hyperplasias, 2/47, 3/44 and 0/39 papillomas and 0/47, 2/44 and 24/39 carcinomas.

Fifty mice that had been treated epicutaneously with epichlorohydrin (2 mg in 0.1 ml acetone, three times a week for 580 days) developed no tumours (Van Duuren *et al.* 1974). This observation is consistent with the findings of Weil *et al.* (1963), who observed no tumour formation after lifetime application of one brush filling each of undiluted epichlorohydrin to the shaved dorsal skin of 90-day-old CH3 mice three times a week.

After subcutaneous administration of 1 mg epichlorohydrin in 0.05 ml tricaprilin once a week for 580 days, 6/50 female IVR/HA Swiss mice developed local sarcomas and one developed an adenocarcinoma. The control incidence for local sarcomas was 1/50 (Van Duuren *et al.* 1974). Another, similar study yielded sarcomas in 2/50 mice (Van Duuren *et al.* 1972).

The intraperitoneal injection of 1 mg epichlorohydrin in 0.05 ml tricaprilin, once a week for 450 days, led to lung papillomas in 11 of 30 ICR/HA Swiss mice, whereas lung papillomas were observed in 10 of 30 control animals treated with tricaprilin and a local sarcoma was observed in one mouse (Van Duuren *et al.* 1974).

2.7.3 Mode of action and cancer risk assessment

Epichlorohydrin is reasonably anticipated to be a human carcinogen based on the experimental data (IARC 1999, NTP 2002). The primary experimental tumours are local. When administered by gavage, it induced forestomach tumours of rats. By inhalation (minimal effective concentration: 30 ppm), it induced tumours of the nasal cavity in rats. Subcutaneous injection produced local sarcomas in mice.

The local effect of epichlorohydrin on the rat nasal tissue has a parallel in the effect of the non-chlorinated compound, propylene oxide (for documentation, see SCOEL/SUM 161). Similar to propylene oxide, the induction of local cell proliferation appears to be a decisive factor (Girolamo et al. 2006). A particular impact of peak concentrations of epichlorohydrine for the local cancer risk is in-line with this view (Ginsberg *et al.* 1996).

In vitro, the genotoxicity (cell transformation tests and DNA strand break induction) of both chemically related compounds has been compared, and epichlorohydrine turned out to be about 10-times more genotoxic than propylene oxide (Kolman and Dusinská 1995, Kolman *et al.* 1997).

Similar to propylene oxide, the protective effect of a rapid metabolic detoxification of epichlorohydrin is of relevance (Hindsø Landin *et al.* 1999).

By contrast to propylene oxide, epichlorohydrin is a bifunctional alkylating agent and induces DNA interstrand cross-links (Romao *et al.* 2007). Therefore, although there are similarities between epichlorohydrin and propylene oxide, differences in the modes of action ought to be considered.

The validity of the present database for the derivation of a quantitative assessment of human cancer risk has been a matter of debate. On the one hand, some authors consider the quantitative experimental data as not sufficient for such an assessment (Ginsberg *et al.* 1996, Kolman *et al.* 2002). On the other hand, the Dutch Expert Committee on Occupational Standards (DECOS) has used a linear extrapolation from the experimental data as a default method and estimated the additional lifetime cancer risk for epichlorohydrin to be 4×10^{-5} for 40 years of human occupational exposure to 0.19 mg/m³, and accordingly 4×10^{-3} for 40 years of occupational exposure to 19 mg/m³ (DECOS 2000).

2.8 Reproductive toxicity

2.8.1. Human data

Two studies of possible fertility disorders after exposure to epichlorohydrin, and in some cases simultaneous exposure to allyl chloride and 1,3-dichloropropene, were negative (Milby and Whorton 1980; Venable *et al.* 1980).

4.9.2 Animal data

John et al. (1983a) exposed 30 male Sprague-Dawley rats and 10 New Zealand rabbits to concentrations of 0, 0.5, 25 and 50 ml epichlorohydrin/m³ over 10 weeks for 6 hours daily on 5 days per week. The male rats (25) were mated with non-exposed female rats during and up to 10 weeks after exposure. The rate of fertilized females was significantly reduced only in the rats exposed to 50 ppm during the exposure phase (tested after 2, 4, 7 and 10 weeks), but not in the matings after the end of exposure (tested after 2, 5 and 10 weeks). The number of implantations was however significantly reduced during the exposure phase at 25 ppm. Histopathology or the weight of the reproductive organs revealed no changes compared with the control either during exposure or after the exposure period. The exposed rabbits were only mated in the tenth week of exposure and showed no reduced fertility.

Daily oral administration of 15 mg epichlorohydrin/kg body weight for 12 days led to sterility in male SD rats after one week. The animals were fertile again one week later (Hahn 1970). The histopathological examination of the testes, epididymides, prostate and seminal vesicles on day 12 of treatment revealed no differences from the control animals. This statement is based on an abstract without data and is therefore only of limited validity.

Cooper *et al.* (1974) observed sterility in male Sprague-Dawley rats lasting up to 10 weeks after five oral administrations of 50 mg/kg body weight daily and reduced fertility for the same period after a single administration of 100 mg/kg body weight (5 males per dose). The histopathological examination of the complex of testes, epididymides and *ductus deferens* revealed no changes up to 8 weeks after the single treatment. The validity of the study is restricted since the number of animals used was small and there was no control group.

In a study carried out by Cassidy *et al.* (1983) in Wistar rats, a significant increase in morphologically abnormal sperm head counts in sonicated testicular homogenates was observed in the group with higher exposure 11 days after the single oral administration of 25 and 50 mg/kg body weight. Total sperm counts were clearly reduced only in the group with lower exposure. The testis weight was unchanged in both dose groups. The examination of testicular sperm head anomalies 11 days after exposure is not an evaluated method.

Toth et al. (1989) treated male Long-Evans rats orally with 0, 12.5, 25 and 50 mg epichlorohydrin/kg body weight daily for 21 days. Following the last exposure, the males were mated with ovarectomized, hormone-treated females (1:1) for 3 hours to observe the mating behaviour and to obtain sperm samples for analysis. Two days later, the male rats treated with the highest dose were daily mated with one female in the pro-oestrus until all males had successfully copulated once within 5 days. After 48 hours, the male rats were sacrificed for histopathological examinations. Mating behaviour, the sperm count in ejaculates, the percentage of motile sperm or sperm morphology were not affected by the treatment with epichlorohydrin. Although all males treated with 50 mg/kg body weight and day had copulated (confirmed by the formation of a vaginal plug), none of the females was pregnant as opposed to 90% of the control group animals (examination of the implantations in the uteri 15 days after observation of the vaginal plug). The histopathological examination only showed a significant reduction of the sperm count in the caudae epididymides at the highest dose. Various motility parameters were however changed in relation to the dose (vigour and swimming pattern). The authors discussed this change as the cause of the lack of fertilization of ova in the highest dose group. It may have been due to damage to the spermatozoa energy metabolism in the epididymis induced by the metabolite 3-chloro-1,2-propanediol.

After intraperitoneal treatment of rats with 3 (n = 3) and 6 (n = 7) mg epichlorohydrin/kg body weight and day for 4 days, sperm were obtained from the proximal region of the *caudae epididymides* and introduced into the uterus of stimulated female rats on day 5. On day 9, *corpora lutea* on the ovaries and implantations in the uteri were counted. These fertility parameters were reduced at both dose levels (Klinefelter *et al.* 1997).

Two examinations with the metabolite 3-chloro-1,2-propanediol provide information about the cause of the antifertile effect of epichlorohydrin.

Slott et al. (1997) treated groups of 9 male Syrian hamsters with 0, 33, 49, 66 and 83 mg 3chloro-1,2-propanediol/kg body weight and day for 4 consecutive days, mated them on day 5 and counted the foetuses in the uteri of the fertilized females on the day before parturition. There was a dose-dependent decrease in the pregnancy rate of the spermpositive females (100%, 78%, 67%, 22% and 0%). Epididymal sperm from the same males showed unaffected percentages of motile sperm, but sperm motility was reduced in relation to the dose. The sperm from treated males were also less likely to support *in vitro* fertilization (IVF). The authors concluded that 3-chloro-1,2-propanediol impairs sperm function.

A single oral administration of 5, 10, 25, 50 and 75 mg 3-chloro-1,2-propanediol/kg body weight reduced the following fertility parameters in SD rats: sperm ATP levels (3 hours and 5 days after treatment with 10 mg/kg body weight and higher), sperm motility (3 hours after treatment with 25 mg/kg body weight) and binding and penetration rates of zona *pelucida*-free oocytes *in vitro* from 10 mg/kg body weight without further increases at higher doses (Jelks *et al.* 2001). The authors concluded that altered ATP levels induced by 3-chloro-1,2-propanediol impair the fertilizing ability of sperm and thus confirm the assumptions of Toth *et al.* (1989).

No prenatal toxicity was found in an inhalation study with pregnant rats and rabbits at concentrations of 2.5 and 25 ppm although food consumption and weight gain of the rats were reduced at the high concentration (CMA 1979c; John *et al.* 1983b).

No teratogenic effects were observed in studies with rats and mice after oral administration of up to 160 mg/kg body weight, not even at maternally toxic doses and doses that led to reduced foetal weights (Marks *et al.* 1982).

Recommendation

Epichlorohydrin is a directly acting genotoxic carcinogen in animal studies with a mainly local effect, with the target in the upper respiratory tract tissues after inhalation. Pituitary tumours were also induced experimentally. An epidemiological study showed possible associations between exposure to epichlorohydrin and the occurrence of CNS tumours. However, these data are not sufficient to derive a conclusive evaluation of the carcinogenicity for humans, nor can a safe concentration be specified for humans at present (2.7.3).

On the basis of the data on the genotoxicity of epichlorohydrin *in vivo*, particularly cytogenetic findings and findings on the development of epichlorohydrin-specific DNA adducts among persons exposed to epichlorohydrin, epichlorohydrin has been classified in germ cell mutagen.

In consequence of the clearcut direct genotoxicity, epichlorohydrin is categorised into the SCOEL carcinogen group A as a non-threshold carcinogen (Bolt and Huici-Montagud 2008). Accordingly, the derivation of a health-based OEL is not possible. An assessment of human cancer risk based on the available experimental data is accompanied with great uncertainties (2.7.3). The Dutch Expert Committee on Occupational Standards (DECOS) has applied a linear extrapolation from the experimental data as a default method and estimated the additional lifetime cancer risk for epichlorohydrin to be 4 x 10⁻⁵ for 40 years of human occupational exposure to 0.19 mg/m³, and accordingly 4 x 10⁻³ for 40 years of occupational exposure to 19 mg/m³ (DECOS 2000).

Although analytical methods have been described, which may serve as a basis for biological monitoring (2.3.1), occupational field studies are lacking, so that a recommendation for a biological monitoring guidance value cannot be given.

Several, although not always adequately documented, clinical findings on the sensitizing effect of epichlorohydrin on the skin are available. Animal studies provided evidence of skin sensitization.

Epichlorohydrin not only has local effects, but also shows systemic toxicity and is lethal after repeated epicutaneous application. However, here the corrosive effect may have destroyed the skin barrier. Absorption of diluted, no longer irritant solutions via intact skin cannot be ruled out. Therefore, a skin notation is recommended.

In view of the above, SCOEL strongly recommends avoiding occupational exposure to epochlorohydrin.

References

Asita AO, Hayashi M, Kodama Y, Matsuoka A, Suzuki T, Sofuni T (1992) Micronucleated

- reticulocyte induction by ethylating agents in mice. Mutat Res 271: 29-37 Bader M, Rosenberger W, Gutzki FM, Tsikas D (2009) Quantification of N-(3-chloro-2hydroxypropyl)valine in human haemoglobin as a biomarker of epichlorohydrin exposure by gas chromatography-tandem mass spectrometry with stable-isotope dilution. J Chromatogr B Analyt Technol Biomed Life Sci 877: 1402-1415
- Barbone F, Delzell E, Austin H, Cole P (1992) A case-control study of lung cancer at a dye and resin manufacturing plant. Am J Ind Med 22: 835-849
- Barbone F, Delzell E, Austin H, Cole P (1994) Exposure to epichlorohydrin and central nervous system neoplasms at a resin and dye manufacturing plant. Arch Environ Health 49: 355–358

Basting I, Hoopmann M, Ehrenstein V, Suchenwirth R, Tödt H, Reichert J, Dressel H, Rosenberger A, Schmid M, Nowak D, Radon K (2006) Acute effects on the health of children after accidental exposure to epichlorohydrine. Gesundheitswesen 68: 309-315

- Beck MH, King CM (1983) Allergic contact dermatitis to epichlorhydrin in a solvent cement. Contact Dermatitis 9: 315
- Bolt HM, Huici-Montagud A (2008) Strategy of the Scientific Committee on Occupational Exposure Limits (SCOEL) in the derivation of occupational exposure limits for carcinogens and mutagens. Arch Toxicol 82: 61-64
- Bond GG, Flores GH, Shellenberger RJ, Cartmill JB, Fishbeck WA, Cook RR (1986) Nested case-control study of lung cancer among chemical workers. Am J Epidemiol 124: 53–66
- BUA (Beratergremium für Altstoffe Advisory Committee on Existing Chemicals of Environmental Relevance) (1992) Epichlorhydrin, BUA Stoffbericht 90, Hirzel Wissenschaftliche Verlagsgémeinschaft, Stuttgart
- Buckley LA, Jiang XZ, James RA, Morgan KT, Barrow CS (1984) Respiratory tract lesions induced by sensory irritants at the RD₅₀ concentration. Toxicol Appl Pharmacol 74: 417-429
- Cassidy SL, Dix KM, Jenkins T (1983) Evaluation of a testicular sperm head counting technique using rats exposed to dimethoxyethyl phthalate (DMEP), glycerol alpha-monochlorohydrin (GMCH), epichlorohydrin (ECH), formaldehyde (FA), or methyl methanesulfonate (MMS). Arch Toxicol 53:71–78
- Cheng T-J, Hwang S-J, Kuo'H-W, Luo J-C, Chang MJW (1999) Exposure to epichlorohydrin and dimethylformamide, glutathione S-transferases and sister chromatid exchange frequencies in peripheral lymphocytes. Arch Toxicol 73: 282–287
- CMA (Chemical Manufacturers Association) (1979a) Pharmacokinetics of epichlorohydrin (EPI) administered to rats by gavage or inhalation. Toxicology Research Laboratory Health and Environmental Sciences, Dow Chemical USA, Midland, MI
- CMA (1979b) Epichlorohydrin subchronic studies. I. A 90-day inhalation study in laboratory rodents. Toxicology Research Laboratory Health and Environmental Sciences, Dow Chemical USA, Midland, MI
- CMA (1979c) Epichlorohydrin subchronic studies. IV. The effects of maternally inhaled epichlorohydrin on rat and rabbit embryonal and fetal development. Toxicology Research Laboratory Health and Environmental Sciences, Dow Chemical USA, Midland, MI

Cooper ERA, Jones AR, Jackson H (1974) Effects of α -chlorohydrin and related compounds on the reproductive organs and fertility of the male rat. J Reprod Fertil 38: 379–386

- Daniel FB, Robinson M, Olson GR, Page NP (1996) Toxicity studies of epichlorohydrin in Sprague-Dawlwy rats. Drug Chem Toxicol 19: 41-58
- DECOS [Health Council of the Netherlands: Dutch Expert Committee on Occupational (2000)Epichlorohydrin (1-chloro-2,3-epoxypropane); Standards] health-based calculated occupational cancer risk values. The Hague: Health Council of the Netherlands. Publication no. 2000/10 OSH
- De Flora S, Bennicelli C, Zanacchi P, Camoirano A, Petruzzelli S, Giuntini C (1984) Metabolic activation and deactivation of mutagens by preparations of human lung parenchyma and bronchial tree. Mutat Res 139: 9-14
- Delzell E, Macaluso M, Cole P (1989) A follow-up study of workers at a dye and resin manufacturing plant. J Occup Med 31: 273–278
 DFG [Deutsche Forschungsgemeinschaft] (2003) 1-Chlor-2,3-epoxypropan (Epichlorhydrin).
- In: Toxikologisch-arbeitsmedizinische Begründungen von MAK-Werten, ed. Greim H, 36. Lieferung, pp.1-21, Wiley-VCH, Weinheim
- DFG [Deutsche Forschungsgemeinschaft] (2009) Epichlorohydrine. In: The MAK-Collection, Part 1, Wiley-VCH, Weinheim, in press
- Enterline PE (1982) Importance of sequential exposure in the production of epichlorohydrin and isopropanol. Ann NY Acad Sci 381: 344-349
- Enterline PE, Henderson V, Marsh G (1990) Mortality of workers potentially exposed to epichlorohydrin. Br J Ind Med 47: 269–276

Epstein E (1974) Allergy to epichlorohydrin masquerading as trichloroethylene allergy. Contact Dermatitis Newslett 16: 475

Epstein SS, Arnold E, Andrea J, Bass W, Bishop Y (1972) Detection of chemical mutagens by the dominant lethal assay in the mouse. Toxicol Appl Pharmacol 23: 288–325

Fakhouri G, Jones AR (1979) Epichlorohydrin: metabolism and toxicity in the rat. Aust J Pharm Sci 8: 11–14

Fregert S, Gruvberger P (1970) Sensitization to epichlorohydrin and cross-sensitization to propane oxide. Contact Dermatitis Newsletter 8: 172

Freuder E, Leake CD (1941) The toxicity of epichlorhydrin. Univ Calif Berkeley Publ Pharmacol 2: 69–78

Gardner RJ, Burgess BA, Kennedy Jr GL (1985) Sensory irritation potential of selected nasal tumorigens in the rat. Food Chem Toxicol 23: 87–92 Gingell R, Mitschke HR, Dzidic I, Beatty PW, Sawin VL, Page AC (1985) Disposition and

Gingell R, Mitschke HR, Dzidic I, Beatty PW, Sawin VL, Page AC (1985) Disposition and metabolism of [2-14C]epichlorohydrin after oral administration to rats. Drug Metab Dispos 13: 333–341

Ginsberg GL, Pepelko WE, Goble RL, Hattis DB (1996) Comparison of contact site cancer potency across dose routes: case study with epichlorohydrin. Risk Anal 16: 667-681

Giri AK (1997) Genetic toxicology of epichlorohydrin: a review. Mutat Res 386: 25-38

Girolamo F, Elia G, Errede M, Vigintino D, Cantatore S, Losusso L, Roncali L, Bertossi M, Ambrosi L (2006) In vivo assessment of epichlorohydrin effects: the chorioallantoic membrane model. Med Sci Monit 12: BR21-27

Hahn JD (1970) Post-testicular antifertility effects of epichlorohydrin and 2,3epoxypropanol. Nature 226: 87

Hellmér L, Bolcsfoldi G (1992) An evalutation of the E. coli K-12 uvrB/recA DNA repair host mediated assay. II. In vivo results for 36 compounds tested in the mouse. Mutat Res 272: 161–173

Hindsø Landin H, Osterman-Golkar S, Zorcec V, Törnqvist M (1996) Biomonitoring of epichlorohydrin by hemoglobin adducts. Anal Biochem 240: 1–6

Hindsø Landin H, Grummt T, Laurent C, Tates A (1997) Monitoring of occupational exposure to epichlorohydrin by genetic effects and hemoglobin adducts. Mutat Res 381: 217–226 Hindsø Landin H, Segerbäck D, Damberg C, Osterman-Golkar S (1999) Adducts with

Hindsø Landin H, Segerbäck D, Damberg C, Osterman-Golkar S (1999) Adducts with haemoglobin and with DNA in epichlorohydrin-exposed rats. Chem Biol Interact 117: 49–64

Hine C, Rowe VK, White ER, Darmer Jr KI, Youngblood GT (1981) Epoxy compounds. In: Clayton GD, Clayton FE (Eds) Patty's industrial hygiene and toxicology, ed. 2A, Wiley-Interscience, New York, 2141–2257

Holzer J, Voss B, Karroum S, Hildman H, Wilhelm M (2008) A comparative study of chemically induced DNA damage in isolated nasal mucosa cells of humans and rats assessed by the alkaline comet assay. J Toxicol Environ Health A 71: 936-946.

IARC (International Agency for Research on Cancer) (1999) IARC monographs on the evaluation of carcinogenicity of chemicals to man, ed. 71, IARC, Lyon, 603–628

Ippen H, Mathies V (1970) Die "protrahierte Verätzung". Berufsdermatosen 18: 144–165

- Jelks K, Berger T, Horner C, Miller MG (2001) α-Chlorohydrin induced changes in sperm fertilizing ability in the rat: association with diminished sperm ATP-levels and motility. Reprod Toxicol 15: 11–20
- John JA, Quast JF, Murray FJ, Calhoun LG, Staples RE (1983a) Inhalation toxicity of epichlorohydrin: effects on fertility in rats and rabbits. Toxicol Appl Pharmacol 68: 415–423

John JA, Gushow TS, Ayres JA, Hanley Jr TR, Quast JF, Rao KS (1983b) Teratologic evaluation of inhaled epichlorohydrin and allyl chloride in rats and rabbits. Fundam Appl Toxicol 3: 437–442

Jolanki R (1991) Occupational skin diseases from epoxy compounds. Acta Derm Venereol 159, Suppl: 1–80

van Joost T (1988) Occupational sensitization to epichlorohydrin and epoxy resin. Contact Dermatitis 19: 278–280

van Joost T, Roesyanto ID, Satyawan I (1990) Occupational sensitization to epichlorohydrin (ECH) and bisphenol-A during the manufacture of epoxy resin. Contact Dermatitis 22: 125–126

Kanerva L, Jolanki R, Alanko K, Estlander T (1999) Patch-test reactions to plastic and glue allergens. Acta Derm Venereol 79: 296–300

Kirkhart B (1981) Micronucleus test on 21 compounds. In: de Serres FJ, Ashby J (Eds) Evaluation of short-term tests for carcinogens. Report of the International Collaborative Program, Progress in mutation research, ed. 1, Elsevier, Amsterdam, 698–704

Klinefelter GR, Laskey JW, Ferrell J, Suarez JD, Roberts NL (1997) Discriminant analysis indicates a single sperm protein (SP22) is predictive of fertility following exposure to epididymal toxicants. J Androl 18: 139–150

epididymal toxicants. J Androl 18: 139–150 Knaap AGAC, Voogd CE, Kramers PGN (1982) Comparison of the mutagenic potency of 2-chloroethanol, 2-bromoethanol, 1,2-epoxybutane, epichlorohydrin and glycidaldehyde in Klebsiella pneumoniae, Drosophila melanogaster and L5178Y mouse lymphoma cells. Mutat Res 101: 199–208 C

- Kolman A, Dusinská M (1995) Comparison of propylene oxide and epichlorohydrin effects in two transformation tests (C3H/1013:511/2 and SHE cells). Toxicol Lett 81: 213-221 Kolman A, Spival I, Näslund M, Dusinská M, Cedervall B (1997) Propylene oxide and
- epichlorohydrin induce DNA strand breaks in human diploid fibroblasts. Erviron Mol Mutagen 30: 40-46
- Kolman A, Chovanec M, Osterman-Golkar S (2002) Genotoxic effects of ethylene oxide, propylene oxide and epichlorohydrin in humans. Update review (1990-2001). Mutat Res 512: 173-194.
- Konishi Y, Kawabata A, Denda A, Ikeda T, Katada H, Maruyama H, Higashiguchi R (1980) Forestomach tumors induced by orally administered epichlorohydrin in male Wistar rats. Gann 71: 922-923
- Kucerová M, Zhurkov VS, Políková Z, Ivanova JE (1977) Mutagenic effect of epichlorohydrin. II. Analysis of chromosomal aberrations in lymphocytes of persons occupationally exposed to epichlorohydrin. Mutat Res 48: 355–360

Laskin S, Sellakumar AR, Kuschner M, Nelson N, La Mendola S, Rusch GM, Katz GV, Dulak NC, Albert RE (1980) Inhalation carcinogenicity of epichlorohydrin in noninbred Sprague-Dawley rats. J Natl Cancer Inst 65: 751–757

Lawrence WH, Malik M, Turner JE, Autian J (1972) Toxicity profile of epichlorohydrin. J Pharm Sci 61: 1712–1717

Lefaux R (1966) Chemie und Toxikologie der Kunststoffe, Krauskopf-Verlag für Wirtschaft, Mainz

Luo JC, Cheng TJ, Kuo HW, Chang MJ (2004) Decreased lung function associated with occupational exposure to epichlorohydrin and the modification effects of glutathione

S-transferase polymorphisms. J Occup Environ Med 46: 280-286 Marks TA, Gerling FS, Staples RE (1982) Teratogenic evaluation of epichlorohydrin in the mouse and rat and glycidol in the mouse. J Toxicol Environ Health 9: 87–96

Milby TH, Whorton D (1980) Epidemiological assessment of occupationally related, chemically induced sperm count suppression. J Occup Med 22: 77-82

NTP [National Toxicology Program] (2002) Epichlorohydrin. Rep Carcinog 10: 113-114. Olsen GW, Lacy SE, Chamberlin SR, Albert DL, Acreneaux TG, Bullard LF, Stafford BA, Boswell JM (1994) Retrospective cohort mortality study of workers with potential exposure to epichlorohydrin and allyl chloride. Am J Ind Med 25: 205-218

Pallade S, Dorobantu M, Rotaru G, Gabrielescu E (1967) Étude expérimentale de l'intoxication par l'épichlorhydrine. Arch Mal Prof Med Trav Secur Soc 28: 505–516
 Picciano D (1979) Cytogenetic investigation of occupational exposure to epichlorohydrin.

Mutat Res 66: 169-173

Plna K, Osterman-Golkar S, Nogradi E, Segerbäck D (2000) ³²P-Post-labelling of 7-(3-chloro-2-hydroxypropyl)guanine in white blood cells of workers occupationally exposed to epichlorohydrin. Carcinogenesis 21: 275–280 Pozzani UC, Carpenter CP (1960) The toxicity of epichlorohydrin, AIHA, Abstract, Industrial

Health Conference, Rochester, New York

Prens EP, de Jong G, van Joost T (1986) Sensitization to epichlorohydrin and epoxy system components. Contact Dermatitis 15: 85-90

Prodi G, Arfellini G, Colacci A, Grilli S, Mazzullo M (1986) Interaction of halocompounds with nucleic acids. Toxicol Pathol 14: 438-444

Quast JF, Henck JW, McKenna MJ (1979) A 90-day inhalation toxicity study of epichlorohydrin in laboratory rodents (Abstract). Toxicol Appl Pharmacol 48: A 43

Rao KS, Betso JE, Olson RJ (1981) A collection of guinea pig sensitization test results grouped by chemical class. Drug Chem Toxicol 4: 331–351

Rebandel P, Rudzki E (1990) Dermatitis caused by epichlorohydrin, oxprenolol hydrochloride and propranolol hydrochloride. Contact Dermatitis 23: 199

Romano KP, Newman AG, Zahran RW Millard JT (2007) DNA interstrand cross-linking by epichlorohydrin. Chem Res Toxicol 20: 832-838

Rossi AM, Migliore L, Lascialfari D, Sbrana I, Loprieno N, Tortoreto M, Bidoli F, Pantarotto C (1983a) Genotoxicity, metabolism and blood kinetics of epichlorohydrin in mice. Mutat Res 118: 213-226

Rossi AM, Migliore L, Barale R, Loprieno N (1983b) In vivo and in vitro mutagenicity studies of a possible carcinogen, trichloroethylene, and its two stabilizers, epichlorohydrin and 1,2-epoxybutane. Teratogen Carcinogen Mutagen 3: 75-87

Salamone MF, Heddle JA, Katz M (1981) Mutagenic activity of 41 compounds in the in vivo micronucleus assay. In: de Serres FJ, Ashby J (Eds) Evaluation of short-term tests for carcinogens. Report of the International Collaborative Program, Progress in mutation research, ed. 1, Elsevier, Amsterdam, 686–697

Schultz C (1964) Fettleber und chronisch-asthmoide Bronchitis nach Inhalation eines Farbenlösungsmittels (Epichlorhydrin). Dtsch Med Wochenschr 89: 1342–1344

Singh US, Secker-Samuelian K, Solomon JJ (1996) Reaction of epichlorohydrin with 2'deoxynucleosides: cheracterization of adducts. Chem Biol Interact 99: 109-128 Slott VL, Jeffay SC, Dyer CJ, Barbee RR, Perreault SD (1997) Sperm motion predicts fertility in

male hamsters treated with α -chlorohydrin. J Androl 18: 708–716

Srám RJ, Cerná M, Kucerová M (1976) The genetic risk of epichlorohydrin as related to the occupational exposure. Biol Zentralbl 95: 451–462

C

Social Europ

Srám RJ, Zudová Z, Kuleshov NP (1980) Cytogenetic analysis of peripheral lymphocytes in workers occupationally exposed to epichlorohydrin. Mutat Res 70: 115–120

Srám RJ, Tomatis L, Clemmesen J, Bridges BA (1981) An evaluation of the genetic toxicity of epichlorhydrin. A report of an expert group of the International Commission for Protection against Environmental Mutagens and Carcinogens. Mutat Res 87: 299-319

Stoner GD, Conran PB, Greisiger EA, Stober J, Morgan M, Pereira MA (1986) Comparison of two routes of chemical administration on the lung adenoma response in strain A/J mice. Toxicol Appl Pharmacol 82: 19-31

Thorgeirsson A, Fregert S (1977) Allergenicity of expoxy resins in the guinea pig. Acta Derm Venereol 57: 252-256

Topham JC (1980) Do induced sperm-head abnormalities in mice specifically identify mammalian mutagens rather than carcinogens? Mutat Res 74: 379–387

Toth GP, Zenick H, Šmith MK (1989) Effects of epichlorohydrin on male and female reproduction in Long-Evans rats. Fundam Appl Toxicol 13: 16–25

Tsai SP, Gilstrap EL, Ross CE (1996) Mortality study of employees with potential exposure to epichlorohydrin: a 10 year update. Occup Environ Med 53: 299–304

Tsuchimoto T, Matter B E (1981) Activity of coded compounds in the micronucleus test. In: de Serres FJ, Ashby J (Eds) Evaluation of short-term tests for carcinogens. Report of the International Collaborative Program, Progress in mutation research, ed. 1, Elsevier, Amsterdam, 705–711

Van Duuren BL, Katz C, Goldschmidt BM, Frenkel K, Sivak A (1972) Carcinogenicity of haloethers. II. Structure-activity relationships of analogs of bis(chloromethyl)ether. J Natl Cancer Inst 48: 1431–1439

Van Duuren BL, Goldschmidt BM, Katz C, Seidman I, Paul JS (1974) Carcinogenic activity of alkylating agents. J Natl Cancer Inst 53: 695–700 Venable JR, McClimans CD, Flake RE, Dimick DB (1980) A fertility study of male employees

engaged in the manufacture of glycerine. J Occup Med 22: 87-91

Vogel E, Lee WR, Schalet A, Würgler F (1981) Mutagenicity of selected chemicals in

Drosophila in comparative chemical genesis. Environ Sci Res 24: 175–256 Weigel WW, Plotnik HB, Conner WL (1978) Tissue distribution and excretion of ¹⁴C-epichlorohydrin in male and female rats. Res Commun Chem Pathol Pharmacol 20: 275-286

Weil CS, Condra N, Haun C, Striegel JA (1963) Experimental carcinogenicity and acute toxicity of representative epoxides. Am Ind Hyg Assoc J 24: 305–325 Wester PW, van der Heijden CA, Bisshop A, van Esch GJ (1985) Carcinogenicity study with

epichlorohydrin (CEP) by gavage in rats. Toxicology 36: 325–339 Würgler FE, Graf U (1981) Mutagenic activity of 10 coded compounds in the Drosophila

sex-linked recessive lethal assay. In: de Serres FJ, Ashby J (Eds) Evaluation of short-term tests for carcinogens. Report of the International Collaborative Program, Progress in mutation research, ed. 1, Elsevier, Amsterdam, 666–672

Social Europ